A globe of Earth is shown from a perspective that includes North and South America. The globe is semi-transparent, revealing a grid of binary code (0s and 1s) underneath. The text is centered over the globe.

Introduction to Earth system modelling

Andy Ridgwell

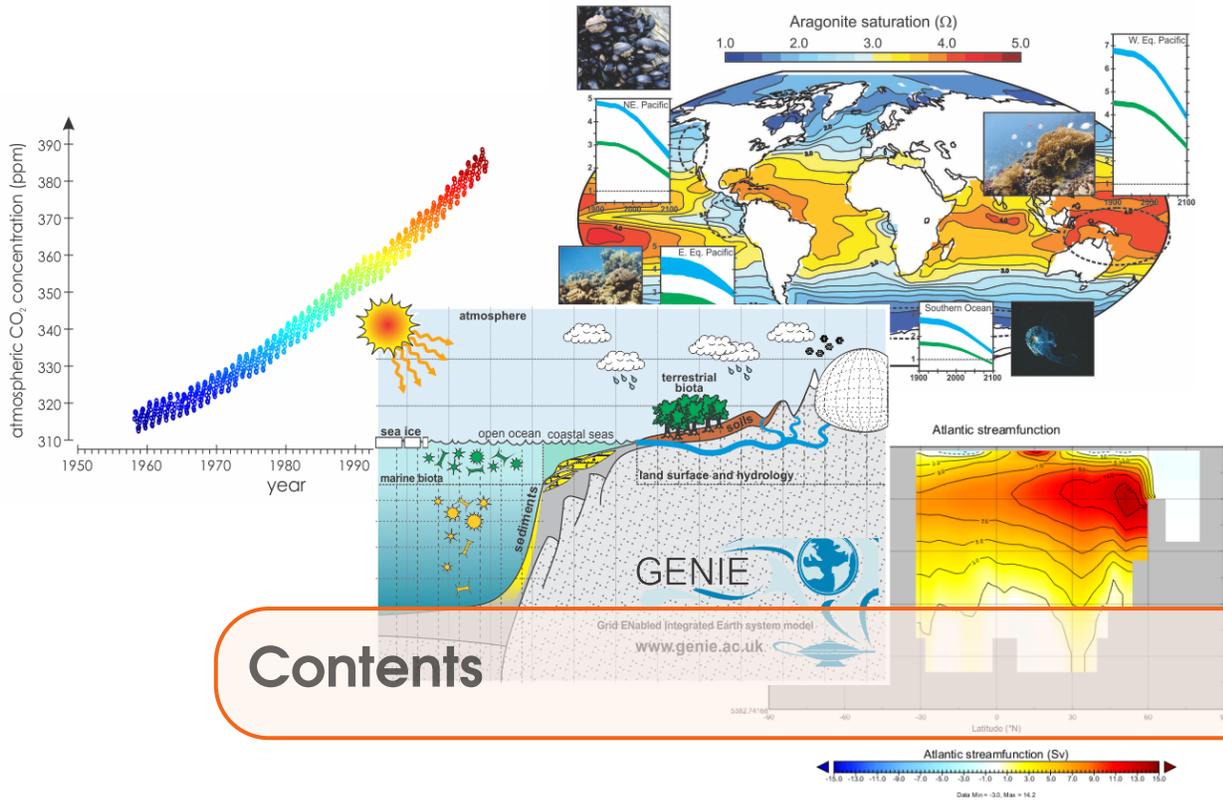
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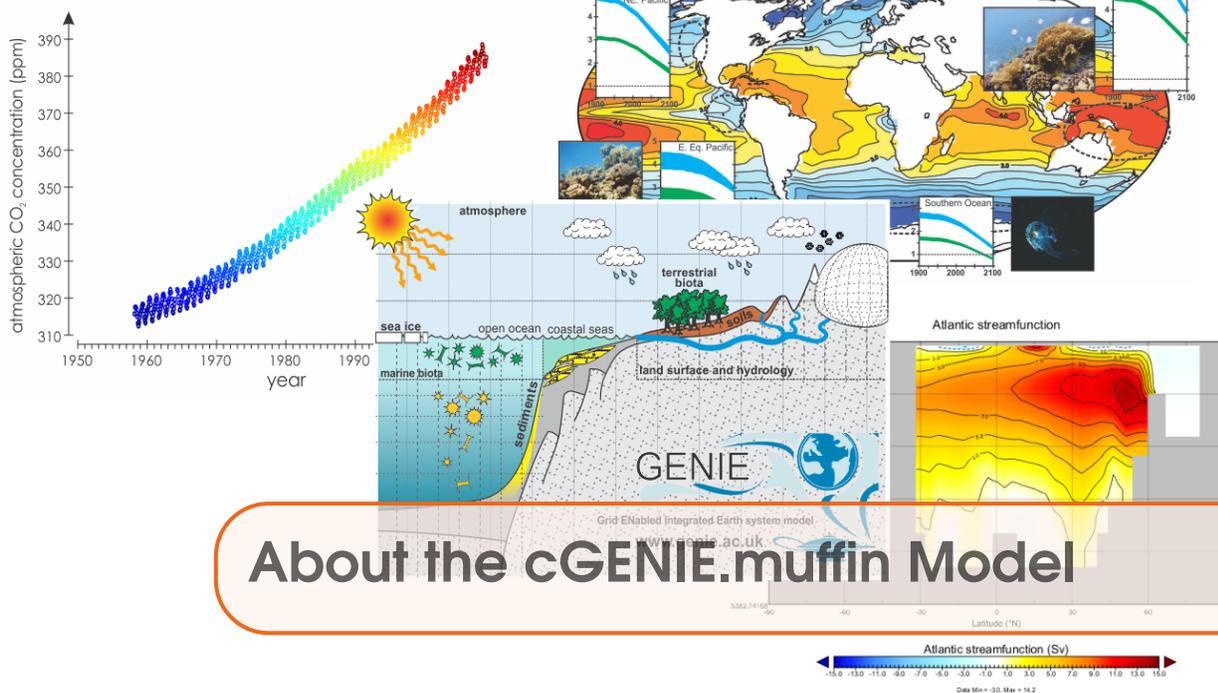
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About the cGENIE.muffin Model

Logistics

The labs are based around using and analysing the ‘**cGENIE**’ Earth system model¹. You will be working in groups of 2 (or 3) and will be remote accessing a computing cluster (where the model will actually run). You will hence need some means of accessing the remote computer. Unless you are some sort of wizard, I suggest a lab desktop, or your own laptop, connected to the internet.

In terms of group working dynamics – if everyone each brings a laptop, then in a group of 2 it becomes easier to be e.g. managing configuring and running the model on one laptop, and analysing results or displaying instructions/documentation on another other.

Required software

If you chose to use a lab desktop, which will be a Windoz 10 based machine, all the software you need will be pre-installed.

If you plan on using your own laptop to remote access the model and visualize results, you will need some specific software. The exact software will depend on your operating system, but everyone will need:

1. A terminal (‘shell’) window. This is no problem for linux and Mac users (you already have one built in). For Windows, either download a simple (and old) SSH client (ssh-client) from my website² or you can get hold of e.g. PuTTY (<http://www.putty.org/>).
2. A sftp (secure file transfer) client for convenience (i.e. dragging and dropping files between local and remote computers, and opening files directly on the remote computer cluster). If you have installed ssh-client (Windows, above) then a sftp client is already included as part of this software. If using PuTTY (Windows) you might try downloading WinSCP (<http://winscp.net/eng/index.php>). For the Mac – I am told that Cyberduck is OK (there are bound to be many other alternatives). For linux, maybe FileZilla.
3. A viewer for netCDF format spatial data. A Java viewer called Panoply is provided by NCAR for all platforms – <http://www.giss.nasa.gov/tools/panoply/> (Note that you will need Java installed!)
4. A simple text editor, except not the rubbish default Windows one – you need one that can display unix ASCII text without screwing it up. Options for Windows users are: notepad++ (<https://notepad-plus-plus.org/>) SciTE (<http://prdownloads.sourceforge.net/scintilla/Sc372.exe>) (linux and Mac users need no special/different editor compared with your standard editor – everything will display just fine).

Running cGENIE.muffin on a laptop

It is also possible to install and run the ‘**cGENIE**’ Earth system model on a linux box (e.g. Ubuntu) or a Mac. Sets of instructions (‘Quick-Start Guides’) are available on my website³ (in the ‘got muffin?’ box on the left).

Note that it is not possible at this time to run **cGENIE** (‘muffin’ version) under Windows (at least, not without near infinite pain).

Also note that if you have trouble installing and running **cGENIE** on your own linux box or Mac, there may not be time to sort out the problem (and in any case I have no clue at all about

¹<http://www.seao2.info/mycgenie.html>

²<http://www.seao2.info/cgenie/software/ssh-client.exe>

³<http://www.seao2.info/mycgenie.html>

Macs). If so, you'll have to access and run the model remotely. (There are also advantages to running on the remote cluster as you will see in due course.)

Before anything else ...

linux ...
ReadMe
Documentation
Version naming conventions
Go!

Starting (dozing?) off ...

Logging in!
Installing and testing the model code
(Notes on file editing)

Running the model #1

Model output

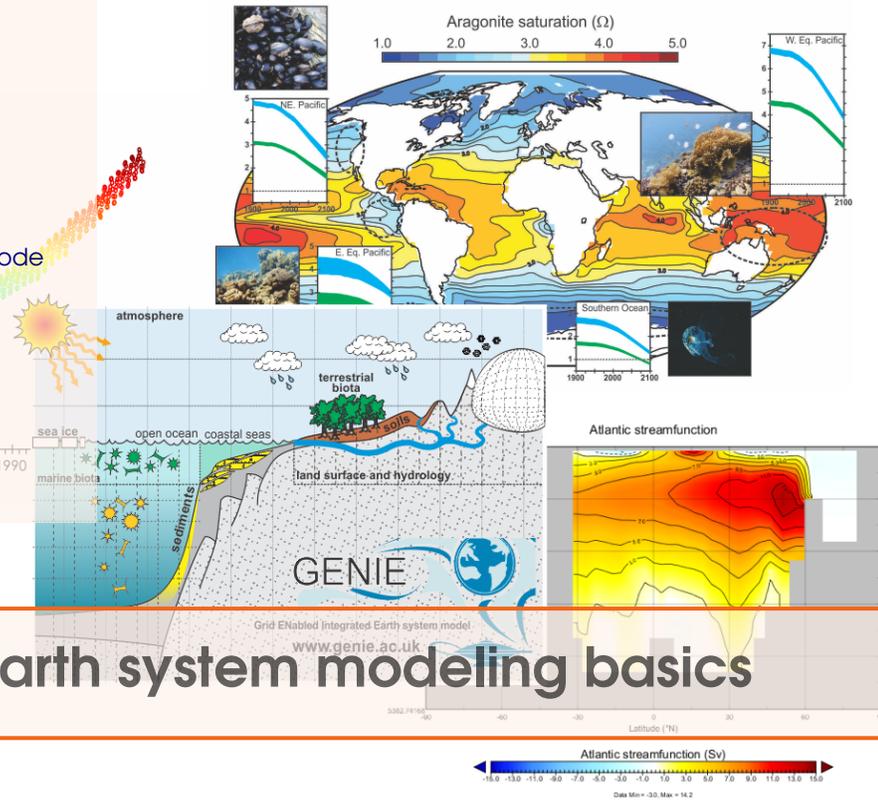
Time-slice output
Time-series output
File naming convention

Viewing model output

Time-series output
2- and 3-D time-slice output

Submitting experiment 'jobs'

'Restarts'



1. Earth system modeling basics

Stuff to keep in mind:

- **cGENIE** is a model. Models ARE NOT the 'real World'. (Don't get confused!)
- The very low resolution (for a 3-D ocean circulation model) of **cGENIE** limits its applicability for very short time-scale problems because the simplified atmospheric component does not simulate winds and there is no atmospheric dynamics or inter-annual variability.
- **cGENIE** is best thought of as a 'discovery and exploring' tool for learning how the Earth system (might) work rather than as a detailed 'simulation' tool.
- Have fun (or at least try).

1.1 Before anything else ...

1.1.1 linux ...

For some of you, the mechanics of running the model will be about as much fun as sticking your tongue in an electrical outlet (a popular hobby in England). (However, if you are an experienced linux/unix/tongue-in-electrical-socket user, you can skip onto the next Section and save yourself an entire 15 seconds of reading words.)

cGENIE has traditionally been configured and accessed (aka ‘run’) at the ‘command line’ of the linux (or Mac equivalent, which is sort of linux also) operating system. The command line is a place where you type text and when you press Return, something (hopefully, good!) happens. Typically the stuff you type started with a ‘command’ word, and often followed by one or more options and parameters. The command word and any options / parameters **MUST** be separated by SPACES.

The start of each line of the command line is indicated with something like: \$. The \$ is called the ‘prompt’ and is ... prompting you to type some input (commands, Tweets, swear words, etc.). See – the computer is just sat there waiting for you to command it to go do something (stupid?). Typically, you will also be informed (reminded) of the username, computer name, and current directory, e.g.:

```
[username@sprout ~]$
```

which in this example is user ‘username’ (yours will be different!) on computer ‘sprout’ (Sprout will eventually appear under ‘cat-of-the-day’ on my homepage if you press F5 enough times – all my computing clusters are named after my cats ...) and the current directory is the ‘home’ directory (~).

If you are not or not very familiar with the linux/unix command line – Appendix 1 contains a quick summary of some of the more important/useful Commands you can use at the command line. Appendix 2 is a version of Appendix 1 for anyone especially nervous about linux.

NOTE: Be **VERY CAREFUL** that spaces are not missed out when typing out example lines. Also be careful not to confuse the number one (1) for the letter el (l). Mis-spelling/typing will probably be the primary reason for any wailing and gnashing of teeth ...

NOTE-the-second: Text in grey, represents instructions specific to using Windoz PCs, and hence may or will differ on different operating systems, e.g. if you have a Macbook. Other software choices for editing files or viewing results are possible as are other ways of configuring software and file editing/transferring methodologies.

Do what suits you best – you can view the instructions as mostly representing an example methodology rather than a literal interpretation of the Constitution.

1.1.2 ReadMe

Some warnings and reminders are repeated over and over and over and ... over again. Some warnings and reminders are repeated over and over and over and ... over again. This is because you will forget immediately each time! ;)

1.1.3 Documentation

Additional documentation (of varying degrees of up-to-date-ness) lives on the interweb¹:

¹<http://www.seao2.info/mycgenie.html>

1. The **cGENIE** User Manual (the least up-to-date of all!)².
2. A set of Examples³.
3. A HOW-TO (explanations of how to get useful stuff done)⁴.
4. A table of model parameters⁵.
5. A Guide to installing **cGENIE**⁶.
6. A README that will inevitably remain un-read ... ⁷.

1.1.4 Version naming conventions

Note that you will be using the development branch of the model and a version code-named ‘muffin’ (if Apple can have ‘Leopard’, ‘Lion’ etc., I can haz baked goods, right?). The documentation may not be fully consistent in this respect ... and you may need to translate occurrences of e.g. a directory named ‘cgenie’ to ‘cgenie.muffin’. Also note that the instructions in general may not be bug-free – use your judgment.

1.1.5 Go!

OK – now we are ready to start ...

²cGENIE.muffin.User_manual.pdf

³cGENIE.muffin.Examples.pdf

⁴cGENIE.muffin.HOWTO.pdf

⁵cGENIE.muffin.Parameter_reference_guide.pdf

⁶cGENIE.muffin.QuickStartGuide.pdf

⁷cGENIE.muffin.README.pdf

1.2 Starting (dozing?) off . . .

You are going to be installing the model from scratch – why? Why not? Hell, it saves me installing it a dozen times. Actually, it will be a happy character-building experience for you – trust me.

1.2.1 Logging in!

Log in to the account that has been created for you on the sprout.ggy.bris.ac.uk computing cluster. To do this – first start the WinSCP program. Under ‘Host Name’, enter:

```
sprout.ggy.bris.ac.uk
```

The ‘Port number’ should be set to 22. Enter your computing cluster user-name on the line below this (‘User Name’) and then the Password. Click on Login. This is your file transfer client.

You will also need a terminal window. This can be opened by clicking on the ‘Open session in PuTTY’ icon on the top icon row, or pressing Ctrl+P.

You should now have TWO windows open – a ‘shell’ window (lines of text on an otherwise blank screen) and a file manager (transfer) window. Ensure that you have both these before moving on. It is recommended that you maximize both these windows to full screen. (But no-one will die horribly for not doing so. Probably ...).

1.2.2 Installing and testing the model code

The next step is to download a copy of the source code for the cGENIE model as follows (all this is done in the shell window of your sprout computer account and it installs the computer code in your sprout account rather than downloading it to your local computer/terminal):

To get a copy of **cGENIE**: from your home directory type:

```
$ svn co
https://svn.ggy.bris.ac.uk/subversion/genie/branches/cgenie.muffin
--username=genie-user cgenie.muffin
```

All this must be on a continuous line, with a S P A C E before `--username`, and before `cgenie.muffin`.

(If you get asked about a ‘certificate’ – enter ‘p’ to accept ‘permanently’.) (Note ... and you’ll ignore this reminder ... the user-name (and password) for the code server is not the same as the user-name and password for the cluster account ... different computers ...)

Depending on the weather and phase of the moon, you may (or may not!) be asked for a password. If so – it is: `g3n1e-user`. Again – don’t mix up the ONE (1) with an ‘el’ (l). (The mystery character in question is a ‘one’ if you must know ...) Watch out that under linux, the characters of the password are not displayed, nor replaced by a *. (It is OK then to ‘store the password unencrypted’ if asked.)

Some of the files that are installed are quite large, and at times, it may appear like it is ‘stuck’. Be patient! Installation should be complete within about 2-3 minutes (longer if everyone else is trying to do this at exactly the same time).

Change directory (see: Appendix I and also Figure 1.1) to:

```
cgenie.muffin/genie-main
```

If you are not 'linux-friendly' – maybe at first do this in steps – list the contents of the directory (`ls`) to check where you are (i.e. what directories are available to chance to), then change to `cgenie.muffin` (`cd cgenie.muffin`), then list again (`ls`) (and see what further directories are there), then change to `genie-main` (`cd genie-main`), and only then ... type:

```
$ make testbiogem
```

This compiles a carbon cycle enabled configuration of **cGENIE** and runs a short test, comparing the results against those of a pre-run experiment (also downloaded alongside the model source code). It serves to check that you have the software environment correctly configured.

There may be some 'Warnings' reported (== somepony's sloppy programming) but these are not detrimental to the ultimate science results (we hope!). 'Success' of this test is indicated by:

```
**TEST OK**
```

You can then be certain that the model you have installed is producing identical (within tolerance) results to everyone else in the World who has ever installed **cGENIE**. Note that the model will pause for a long time at the line:

```
./genie.job -t -k -f configs/eb_go_gs_ac_bg_test.xml -o /home/genie00/cgenie_output ...  
-c /home/genie00/cgenie -g ../../cgenie -m "" > testbiogem.out;
```

This is quite 'normal' – the model is thinking!

Ignore the compiler warnings ... (reflecting my lack of adequate software engineering skills).

That is it as far basic installation goes.

1.2.3 (Notes on file editing)

Later on you will be editing some configuration files. So now might be a good time to check that you can use the/an editor! (You will also be using the same editor to view some of the model output.)

You have two alternative options for editing and viewing text files, depending on whether you are a UNIX nerd with no life, or prefer anything to do with computers to be wrapped in cotton wool and covered with dollops of treacle. EITHER: Use the linux `vi` (`/vim`) application (or similar e.g. `emacs`) if you are familiar with it. I think that this pretty much sucks as a text editor and life is far too short and brutal ... so I will not *make* you ;) OR ... Use a suitable linux-friendly text editor (NOT Micro\$oft Notepad) in conjunction with the Secure File Transfer Client. For example: 'SciTE' <http://www.scintilla.org/SciTE.html> is suitable or Notepad++.

If you fiddle about with the settings under Options/Preferences in the WinSCP program and apply a little common sense, it should be possible to configure things so that you can simply double-click on a file in the remote (right-hand) window panel and it will open like magic (almost)! Saving the file after editing) should then result in the file being saved back to the cluster. Or you can select Edit With (and then SciTE) from right-mouse-button-clicking on the filename.

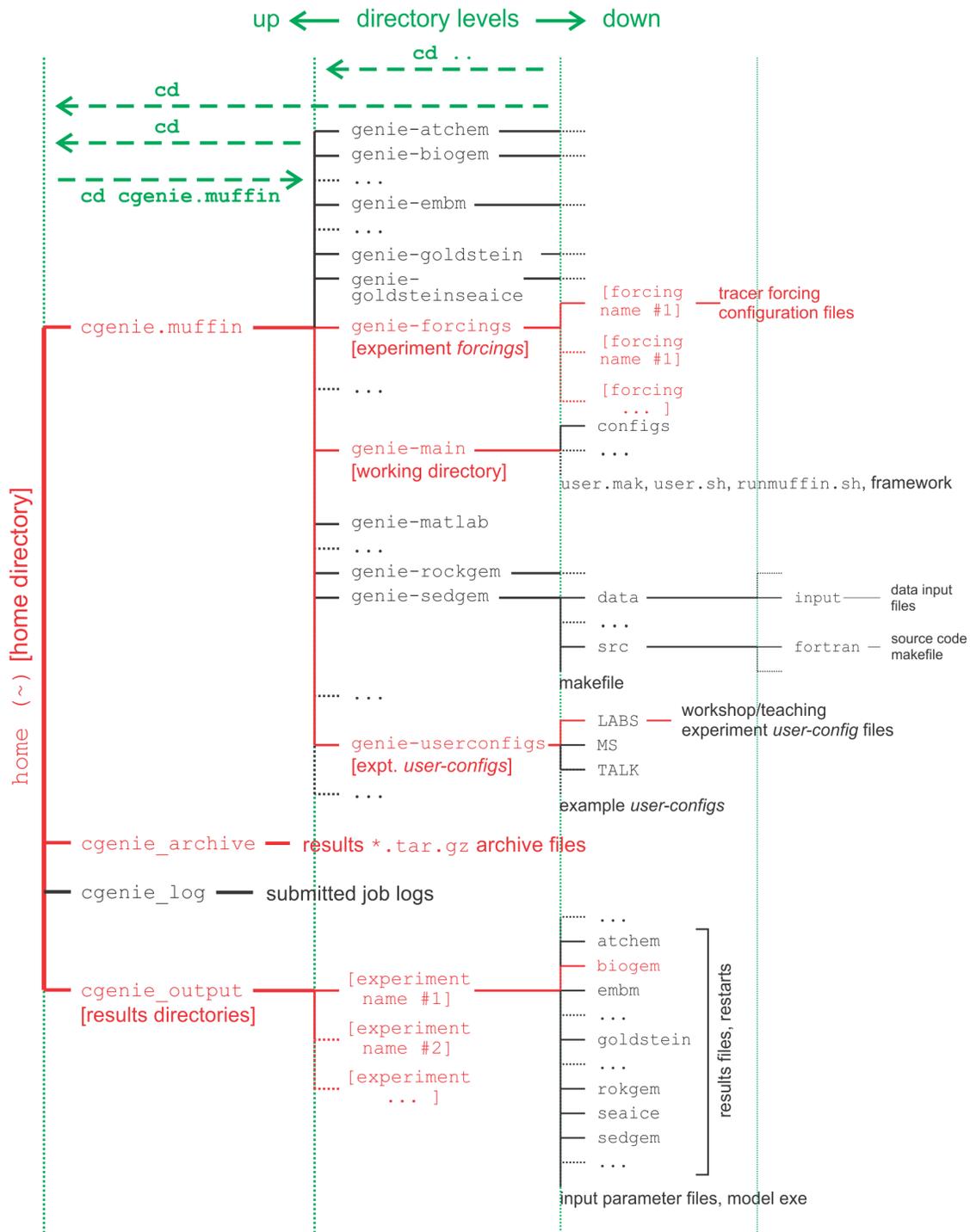


Figure 1.1: Directory structure of the cGENIE model. Highlighted in red are directories and sub-directories that you will need to access at some point. Vertical green lines designate directory levels, with example commands shown for moving between them.

1.3 Running the model #1

The overall sequence of configuring and running the **cGENIE.muffin** model, is shown in Figure 1.2.

At the command-line (\$) in the genie-main directory (not your home directory), you will be entering in a command (`./runmuffin.sh`) together with a list of parameters that will be passed to the model, and as if by magic the model will run (or sometimes not). The form of the command you are going to be issuing is:

```
$ ./runmuffin.sh #1 #2 #3 #4 (#5)
```

In which you must list at least 4 parameters after `./runmuffin.sh`, separated by `S P A C E S` and on a single continuous line (even if it ‘wraps’ around across 2 lines of the screen). These parameters are:

1. ... is the name of the required base (or ‘basic’) configuration (‘base-config’) of the model.
2. ... is the name of the subdirectory (if any) containing the user configuration (‘user-config’) file (i.e., the file containing the specification of a particular experiment).
3. ... is the name of the experiment itself. There must exist a file in the directory specified by parameter #2 (LABS) with exactly the same name as you enter here for parameter #3 (i.e. parameter #3 points to a file in the directory given by parameter #2).
4. ... is the run length of the experiment in years – this must be entered as an integer.

There is also one optional (5th) parameter (described later).

As an example of running the cGENIE Earth system model: parameter #1: The base config is: `cgenie.eb_go_gs_ac_bg.worbe2.BASE` parameter #2: The user config directory is: `LABS` parameter #3: The user config file (the experiment name) is: `LAB_0.EXAMPLE` parameter #4: Run the experiment for ten years: `10` parameter #5: (There is no restart file, and so no 5th parameter needs to be passed ...) The full command for your first example experiment, which you are going to issue from the `~/cgenie.muffin/genie-main` directory, then looks like:

```
$ ./runmuffin.sh cgenie.eb_go_gs_ac_bg.worbe2.BASE LABS LAB_0.EXAMPLE 10
```

REMEMBER: This must be entered on a single CONTINUOUS LINE. The (single) `S P A C E S` are vital. Take care not to confuse an el (‘1’) with a one (‘1’) when typing this in ... (it is a ‘one’ here).

What should happen is: First, you will end up twiddling your thumbs a while, as all the components of cGENIE are compiled from the raw source code (FORTRAN). When it has finished doing this, the model will initialize and carry out some brief self-checking. Only then will it start actually ‘running’ and doing something, starting with a header describing the columns of numbers that follow:

```
model year – ... guess!
ice(%) – global sea-ice fraction (%)
<SST> – global sea surface temperature (‘SST’) °C
<SSS> – global sea surface salinity ‘SSS’ (‰)
```

The choice of what information to display on screen as the model is running is rather arbitrary, but the chosen metrics do tend to summarize some of the main properties of the climate system and carbon cycle – for my own personal convenience rather than reflecting any fundamental scientific truth ... you may also see columns of information for:

$pCO_2(\mu atm)$ – mean atmospheric CO_2 concentration (in units of μatm)
 $\delta^{13}CO_2$ – mean $\delta^{13}C$ value of atmospheric CO_2 (‰) (NOTE: only if ^{13}C tracer is selected)
 $\langle DIC \rangle$ – global mean ocean dissolved inorganic carbon (DIC) concentration ($\mu mol\ kg^{-1}$)
 $\langle ALK \rangle$ – global mean ocean alkalinity (ALK) concentration ($\mu eq\ kg^{-1}$) and in experiments with a modern continental configuration, also:

- AMO (Sv) – Atlantic meridional overturning circulation (Sv)

This information is reported at the same intervals as time-series data (see later and/or refer to the User Manual) is saved and is indicated by:

```
>>> SAVING BIOGEM TIME-SERIES AVERAGE CENTERED @ year :
```

Interleaved between these lines are lines reporting the saving of time-slice data (the 2- and 3-D model states – more of which later as well as in the User Manual). These appear as:

```
>>> SAVING BIOGEM TIME-SLICE AVERAGE CENTERED @ year :
```

You can stop the model at any point (all data up to that time will have been saved) by hitting: $\langle Ctrl-C \rangle$ (CONTROL key + ‘C’ key).

Just from examining the screen output: how close to steady state does the system appear to have come after just 10 years? i.e., do SST and/or sea-ice extents appear to be converging towards stable (constant) values? This will be an important question to think about later on: ‘has the model reached steady-state (and does it matter)?’

In this example, the output should look something like the following:

```
*****
*** Initialisation complete: simulation starting ...
*****
```

	model year	* pCO2(uatm)	d13CO2	* AMO(Sv)	ice(%)	<SST>	<SSS>	* <DIC>(uM)	<ALK>(uM)	
\$N\$	0.00	278.000	-6.500	0.000	0.000	-0.000	34.900	2244.000	2363.000	
>>>	SAVING BIOGEM TIME-SLICE AVERAGE CENTERED @ year :						0.500			
>>>	SAVING BIOGEM TIME-SERIES AVERAGE CENTERED @ year :						0.500			
\$N\$	1.00	279.960	-6.598	13.613	0.744	2.509	34.901	2241.498	2363.111	
>>>	SAVING BIOGEM TIME-SLICE AVERAGE CENTERED @ year :						1.500			
>>>	SAVING BIOGEM TIME-SERIES AVERAGE CENTERED @ year :						1.500			
\$N\$	2.00	279.525	-6.580	12.828	3.499	4.471	34.901	2240.173	2363.135	
>>>	SAVING BIOGEM TIME-SERIES AVERAGE CENTERED @ year :						2.500			
\$N\$	3.00	279.258	-6.568	11.695	5.028	5.996	34.901	2239.169	2363.161	
>>>	SAVING BIOGEM TIME-SERIES AVERAGE CENTERED @ year :						3.500			
\$N\$	4.00	279.044	-6.558	10.444	5.929	7.209	34.901	2238.354	2363.191	
>>>	SAVING BIOGEM TIME-SLICE AVERAGE CENTERED @ year :						4.500			
>>>	SAVING BIOGEM TIME-SERIES AVERAGE CENTERED @ year :						4.500			
\$N\$	5.00	278.899	-6.551	9.380	6.191	8.156	34.902	2237.664	2363.220	
>>>	SAVING BIOGEM TIME-SERIES AVERAGE CENTERED @ year :						5.500			
\$N\$	6.00	278.777	-6.545	8.500	6.623	8.975	34.902	2237.069	2363.246	
>>>	SAVING BIOGEM TIME-SERIES AVERAGE CENTERED @ year :						6.500			
\$N\$	7.00	278.680	-6.541	7.922	6.629	9.637	34.903	2236.548	2363.267	
>>>	SAVING BIOGEM TIME-SERIES AVERAGE CENTERED @ year :						7.500			
\$N\$	8.00	278.601	-6.537	7.917	6.738	10.225	34.903	2236.087	2363.285	
>>>	SAVING BIOGEM TIME-SERIES AVERAGE CENTERED @ year :						8.500			
\$N\$	9.00	278.528	-6.534	7.952	6.740	10.732	34.904	2235.682	2363.301	
>>>	SAVING BIOGEM TIME-SLICE AVERAGE CENTERED @ year :						9.500			
>>>	SAVING BIOGEM TIME-SERIES AVERAGE CENTERED @ year :						9.500			
\$N\$	10.00	278.466	-6.531	8.025	6.694	11.176	34.904	2235.325	2363.314	

```
*****
*** Simulation complete: shutdown starting ...
*****
```

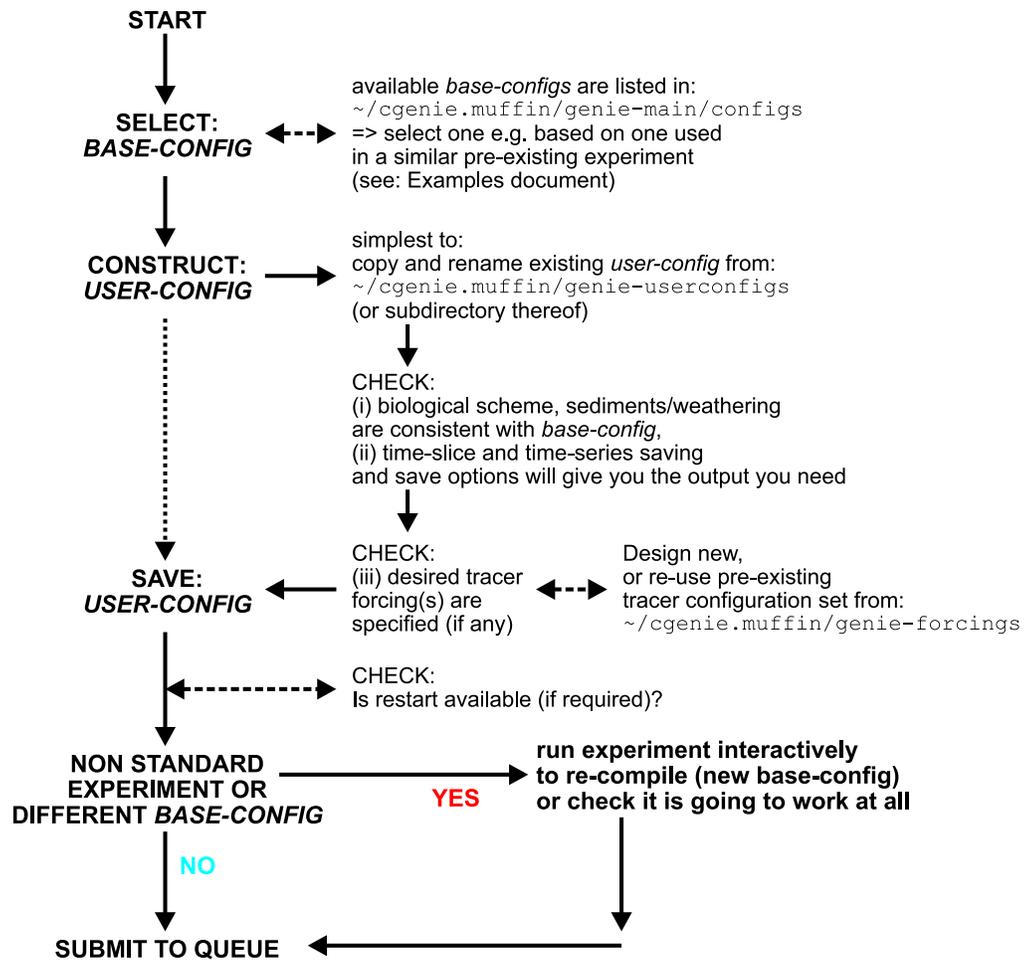


Figure 1.2: Schematic of the sequence-of-events in configuring and running an experiment.

1.4 Model output

The first thing to note about output (i.e., saved results files) from **cGENIE** is that every science module saves its own results in its own sub-directory (and sometimes in very different and difficult-to-fathom ways ...) – see Figure 1.1. All the sub-directories of results, plus copies of input parameters and the model executable, are gathered together in a directory that is assigned the same name as the experiment (= *user-config* file name). The experiment results directories all live in:

```
~/cgenie_output
```

and will be assigned a directory name something like:

```
LAB_0.EXAMPLE
```

(this being the results directory name for an experiment called LAB_0.EXAMPLE). Within this directory are each module's results sub-directories. We will primarily consider only results saved by the ocean biogeochemical module '**BIOGEM**' (subdirectory: biogem). The results files in this example will thus be found in:

```
~/cgenie_output/LAB_0.EXAMPLE/biogem
```

BIOGEM has a flexible and powerful facility of saving results by means of spatially explicit 'time-slices', and as a semi-continuous 'time-series' of a single global (or otherwise representative mean) variable. In contrast, **ATCHEM** does not save its own results (**BIOGEM** can save information about atmospheric composition and air-sea gas exchange) while **SEDGEM** essentially saves results only at the very end of a model experiment (**BIOGEM** can also save the spatial distribution of sediment composition as time-slices as well as mean composition as a time-series). Furthermore, in order to attain a common format for both ocean physical properties and biogeochemistry, **BIOGEM** can save a range of ocean results in addition to temperature and salinity, such as: velocities, sea-ice extent, mixed layer depth, convective frequency, etc

1.4.1 Time-slice output

One of the most informative data sets that can be saved is that of the spatial distribution of properties (such as tracers or physical ocean attributes). However, saving full spatial distributions (e.g. a $36 \times 36 \times 8$ array) for any or all of the tracers each and every time-step is clearly not practical; not only in terms of data storage but also because of the detrimental effect that repeated file access has on model run-time. Instead, **BIOGEM** will save the full spatial distribution of tracer properties only at one or more predefined time points (in units of years). These are termed time-slices. At the specified time points, a set of spatially-explicit data fields are saved for all the key tracer, flux, and physical characteristics of the system. However, rather than taking an instantaneous snapshot, the time-slice is constructed as an average over a specified integration interval (the default is set to 1.0 years, i.e. an annual average). **BIOGEM** then assumes that the specified time point represents the mid-point of the annual (by default) average with the results that output years end up being reported as e.g.

```
0.5  
1.5  
2.5  
4.5  
...
```

(the mid-points of averages made over the intervals: 0-1, 1-2, 2-3, 4-5 years, etc.).

1.4.2 Time-series output

The second data format for model output is much more closely spaced in time. Model characteristics must then be reducible to a single meaningful variable for this to be practical (i.e., saving the time-varying nature of 3-D ocean tracer distributions is not). Suitable reduced indicators would be the total inventories in the ocean and/or atmosphere of various tracers (or equivalently, the mean global concentrations / partial pressures, respectively). Like the time-slices, the data values saved in the time-series files represent averages over a specified integration interval (the default is set to 1.0 years (annual average) but the results are reported with respect to the mid-point of the average which is where the ‘.5’ bits come in again).

1.4.3 File naming convention

The **BIOGEM** results directory will contain files with names of the form:

- `_restart.nc` (is the re-start file created from the run you have just complete, and can be ignored).
- `biogem_series_*.res` – these are the time-series files (in ASCII / plain text format).
- `biogem_year*_diag_GLOBAL.res` – these contain (global diagnostics) summary information and are saved at the same frequency as the time-slices (also as ASCII / plain text).
- `fields_biogem_2d.nc` – 2-D fields of ocean and atmosphere properties, as NetCDF.
- `fields_biogem_3d.nc` – 3-D fields of ocean properties, as NetCDF.

1.5 Viewing model output

1.5.1 Time-series output

A descriptive summary of all the time-series (`biogem_series_*.res`) data files is given in the **cGENIE** User Manual if you are really that bored. The files of most immediate use/relevance are:

- `biogem_series_atm_humidity.res` - mean atmospheric (surface) humidity
- `biogem_series_atm_temp.res` - mean atmospheric (surface) air temperature
- `biogem_series_misc_opsi.res` - min/max overturning stream-function values (e.g. AMOC)
- `biogem_series_misc_seaice.res` - mean ocean sea-ice cover and thickness
- `biogem_series_ocn_sal.res` - mean ocean surface and whole ocean salinity
- `biogem_series_ocn_temp.res` - mean ocean surface and whole ocean temperature

One way of viewing the contents of files is the change directory to the experiment results directory and opening the file in the `vi` editor. But that is not so much fun.

Instead – change to the experiment results directory and then to the **BIOGEM** sub-directory in the Secure File Transfer Client, and try double-clicking (if you have set up the WinSCP preferences correctly) or right-mouse-button-clicking (the then Edit with) on one of the `.res` files (listed above). For `biogem_series_ocn_temp.res`, you should see 2 columns – time and mean (whole) ocean temperature (°C). (However, in subsequent exercises a fuller output will be created with additional columns, with one for mean surface ocean temperature ‘SST’ (°C) as well as mean benthic (bottom water) temperature (°C)). Other results files may differ in the numbers of columns but all should be identifiable from the header information.

Note: The Secure File Transfer Client does not automatically refresh the directory listing. If you cannot see the results sub-directory with the experiment name you have just run 99 times out of 100, it is because the display of the Secure File Transfer Client needs to be refreshed – there is an icon at the top of the program window or hit the ‘F5’ key.

For your information and edification (only): Excel, or **MUTLAB** if you prefer, can be used to graph the time-series results. Either way you will have to deal with the header line(s) that are present at the top of the file (and preceding the rows of data).

In Excel: Chose File then Open. You will want to select Files of Type ‘All Files (*.*)’. In the Text Import Wizard window you can request that Excel skips the first few lines to start the import on the 2nd or 3rd line of the text file. Alternatively: set an appropriate column width manually in Excel to ensure that the columns of data are correctly imported.

MUTLAB will ignore lines starting with a %, which the time-series starts with. However, it may be that the header line wraps-around and there is in effect a 2nd header line but without a %. In this case, extra care (or a quick edit of the header in the ASCII file) will be required to load the data into **MUTLAB**.

1.5.2 2- and 3-D time-slice output

For the time-slice NetCDF (*.nc) files you will be using a program called ‘Panoply’. If you want your own (FREE!) copy of this utility, you can get it here (and is available for: Windoz, Mac, and linux operating systems): <http://www.giss.nasa.gov/tools/panoply/>.

When you open the NetCDF file, you will be presented with a ‘Datasets and Variables’ window (on the left hand side of the application window). This contains a list of all the parameters available that you can display. You will find that the ‘Long Name’ description of the variable will be the most helpful to identify the one you want. Simply double-click on a variable to display. For the 3-D fields you will be asked first whether you want a ‘Longitude-Latitude’ or ‘Latitude-Vertical’ plot (for the 2-D fields, the plot display will immediately open). For the ‘Longitude-Latitude’ plots – there are multiple levels (depth layers) in the ocean of data that can be plotted, from the surface to the abyssal ocean. For the ‘Latitude-Vertical plots – there are multiple possible longitudes at which to plot slices. The default is the global mean meridional distribution. There is also an option for ‘Longitude-Vertical plots (which we will not use). For all three: there may be multiple time-slices (i.e., you can plot data saved from different years). You can interpolate the data or not (often you may find that it is clearer not to interpolate the data but to leave it as ‘blocky’ colors corresponding to the resolution of the model), change the scale and colors, overlay continental outline, change the projection, etc etc. Grey cells represent ‘dry’ grid points, i.e., continental or oceanic crust. NOTE: The default settings in Panoply can mislead: (1) By displaying the very 1st time-slice (often year mid-point 0.5) time-slice rather than the experiment end. (This can confuse as it can look like an experiment has not done anything!) (2) By interpolating the data (not always misleading). To remove interpolation, untick ‘Interpolate’ in the ‘Arrays’ tab. (3) By displaying a global zonal mean by default when selecting Latitude-Vertical plots. Then, to further confuse you, by plotting the output up-side-down (to invert: in the ‘Grid’ tab, hit ‘Swap B/T’ (for swap bottom/top). (4) By listing all ‘Plottable variables’ (option at the bottom of the window), when what you ideally want is to ‘Show’ is the shorter and less confusing list of ‘Georeferenced variables’. (5) In Longitude-Latitude plots, by overlaying the modern continental output. (GENIE land is marked in grey.) (6) By fitting a scale to the plot when the display window is opened, but not changing the scale when e.g. time or depth is changed. (The point of confusion is that you can quickly move outside the scale and end up with all model points dark blue or red.) Re-fit the scale, or manually set limited, in the ‘Scale’ tab. So be careful when opening a new plot that you are looking at what you *think* you are looking at ... All the defaults can be changed via the ‘Edit’ drop-down menu and ‘Preferences’.

Explore different data fields and play with different ways of displaying them. Aim for a set of display properties that show the information you are interested in / want to present in the clearest possible manner. Try different years (time-slice number), depth level (for a Latitude-Longitude plot), or longitude (for a vertical section).

To save plots in Panoply: File Save Image As ... Then select the location, filename, and graphics format.

1.6 Submitting experiment 'jobs'

This bit is no particular fun at all, but it is a very handy 'trick' for running the model in the background, and maximizes drinking time in the pub vs. sat bored watching a computer screen :)

Running jobs interactively is all very well, but there are three important limitations: (1) The connection between your terminal and the server computer running the model must remain unbroken. Anything more than a fleeting loss of internet connectivity may result in the experiment terminating. (2) You can only run one experiment at a time ... unless you want to have thousands of separate terminal open ...? I thought not ... (3) Any cluster or computer you are likely to be accessing using a shell will not have many computing cores itself, either because it is a single machine with only one or two processors, or if a cluster, by using a terminal you are running on the 'head node', which will have similar computing core limitations to running on a single machine. The more experiments you run simultaneously, the slower they will all run ...

The alternative is to submit your experiment as a 'job' to a queuing system which then manages what compute resources are used to run the model. Once you have submitted the experiment, that is it – you can go straight to the pub :)

Unless you want to read the cGENIE User Manual, for now, take the following suggested way of submitting jobs on trust: to run the same experiment to before (LAB_0.EXAMPLE) for maybe 100 years (or even longer if you wish – I am just pulling factors of 10 out of thin air here) by submitting the experiment as a job to the cluster queue, type:

```
$ qsub -j y -o cgenie_log -V -S /bin/bash runmuffin.sh
cgenie.eb_go_gs_ac_bg.worbe2.BASE LABS LAB_0.EXAMPLE 100
```

(Again: SINGLE, CONTINUOUS LINE.) Note that now you should omit the './' bit before runmuffin.sh. (If you are interested (I know that you are not): the options following qsub and before runmuffin.sh do things like re-directing screen output and error messaging to a file and specify which linux 'shell' to assume. It is even possible to receive an email when the job is done :)) The status of the cluster queue and how your experiment job is getting on (e.g., "Is it finished yet?") can be checked by typing:

```
$ qstat -f
```

(qstat -f -u "*" will show all jobs on the cluster.)

After submitting an experiment, you receive a job number. This number appears in the first column in the queue status information when you issue a qstat -f command. You should see your job appear on one of 6 compute nodes, numbered 0-0 to 0-5), although it might briefly reside as a 'PENDING JOB'. For each node, there are 24 processing cores, meaning that up to 24 instances of cGENIE can run simultaneously on each node (i.e., a total of 144 across the entire cluster). For an 8-level ocean based configuration of cGENIE, being run for 100 years, the job should remain there in the queue for a few minutes before 'disappearing' (your clue that it has finished, or died⁸ ...). If you periodically re-issue a qstat -f command you can follow your job's progress.

A rough rule of thumb is that 8-level ocean cGENIE @ a horizontal grid resolution of 36x36 will simulate about 1000 years per CPU hour. The 16-level version (which you will use later), runs at about 300-400 years per CPU hour.

⁸If your experiment appears on the queue but vanishes after a few seconds, it has most likely died :(

Cluster job submission in detail ...

For submitting jobs using Sun Grid Engine (SGE) on a cluster, a basic command⁹ would look like this:

```
$ qsub -S /bin/bash runmuffin.sh <options>
```

Here: the `-S /bin/bash` part is to ensure that SGE uses the BASH shell to submit the job because this is the language that `runmuffin.sh` has been written in.

Take care that the installed FORTRAN compiler can be seen by the cluster nodes. If not, the `cGENIE` executable must have already been built prior to submitting a job. The easiest way to do this is to run `cGENIE` interactively briefly (e.g., with a run length of just a couple of years or kill it) and then submit the full run to the cluster.

Other useful submission options for SGE:

- To redirect the standard output stream:

```
$ qsub -o genie_log ...
```

- To redirect the standard error stream:

```
$ qsub -e genie_log ...
```

- To merge standard output and error streams into standard output:

```
$ qsub -j y ...
```

- To specify particular resources, such as the nodes with 8 GB of RAM:

```
$ qsub -l mem_total = 8.0G ...
```

- To decrease the priority of a job¹⁰:

```
$ qsub -p 1 ...
```

- To submit a job from the current working directory:

```
$ qsub -cwd
```

- Request an email is sent when the job starts and/or when it finishes – see the main pages for `qsub` for the required syntax).

- A complete example for one of the University of Bristol clusters would be:

```
$ qsub -j y -o cgenie_log -V -S /bin/bash runmuffin.sh
      cgenie.eb_go_gs_ac_bg.worjh2.ANTH / EXAMPLE.worjh2.Caoetal2009.SPIN 10000
```

which merges standard output and error streams and redirects the resulting file to the directory `~/cgenie_log`. Note that to redirect output as per in this particular example, the directory `~/cgenie_log` **MUST** be present. I have no idea what happens if it is not ... but it can't be good ;)

You can check the status of the SGE job queue with the command:

⁹With a different queue management environment it may be necessary to place the call to `runmuffin.sh` together with its list of parameters into an executable shell, and submit that instead.

¹⁰The default priority is 0. A lower priority has a higher value ... !

```
$ qstat -f
```

and you can kill a job with the `qdel` command, the job numbers being given by the `qstat` command.

Depending on the cluster setup, it may be possible to graphically check what is going on via the `www`. In an X window, enter:

```
$ ssh almond.ggy.bris.ac.uk  
$ firefox --no-remote &
```

and follow the links: 'Cluster Status (Ganglia)', and then 'Job Queue' to get to the actual queue listing and cluster status.

NOTE: It may be that the FORTRAN compiler is not accessible by the computer nodes. The implication of this is that **the *cGENIE* executable must be already compiled BEFORE a job is submitted to the queue.**

In other words; if you have just changed the model resolution or continental configuration, or number of tracers (i.e. changed the *base-config*) or issued a `make cleanall` command you **MUST** briefly run your desired experiment (or equivalent) interactively (i.e., in the shell window) to ensure that everything is correctly compiled. For instance, either run the experiment for a couple of years or start the experiment for the desired full duration, but 'kill it' (Ctrl-C) once the experiment is running successfully.

1.7 ‘Restarts’

Not much fun here either ... but again – an important and time-saving (== increased drinking time!) modelling technique to learn to use.

By default, model experiments start from ‘cold’, i.e., the ocean is at rest and uniform in temperature and salinity while the atmosphere is uniform in temperature and humidity. All biogeochemical tracers in the ocean have uniform concentrations and/or are zero and there are no biogenic materials in deep-sea sediments. From this state it will take several thousand years (kyr) for the climate system to reach steady-state, and closer to 5 kyr (or more) for ocean biogeochemical cycles and atmosphere CO_2 to reach steady-state, and exceeding 100 kyr for sediment composition to re-balance weathering ... Reaching this the equilibrium state is called the ‘spin-up’ phase of the model. There is evidently little point in repeating the spin-up for each and every model experiment that are similar except in a single detail (e.g., testing a variety of different CO_2 emissions scenarios all starting from current year 2012 conditions). A facility is thus provided for requesting that a ‘restart’ is used – starting a new experiment from the end of a previous one, usually a ‘spin-up’ that has been run explicitly for the purpose of generating a starting point (restart) of the system at steady-state (equilibrium) for subsequent experiments to continue on from. It is important to note that there is nothing special about a restart – it is simply an experiment that you have already run. Equally, there is nothing special about the restarts you will download next – these you could have generated yourself – it simply saves time to have them provided.

To experiment with using a restart, you will first need to download a file that has been created (a pre-run 10,000 year spin-up). To fetch this: Change to the `cgenie_output` directory (perhaps by going ‘home’ first (`cd <RETURN>`), and then changing to `cgenie_output` – refer to linux commands crib sheet and Figure 1.1), and type:

```
$ wget http://www.seao2.info/cgenie/labs/AWI.2017/LAB_0.SPIN.tar.gz
```

This downloads an archived/compressed copy of the restart from a location on the interweb. Extract the contents of this archive by typing:

```
$ tar xfvz LAB_0.SPIN.tar.gz
```

Finally, change directory back to `cgenie.muffin` and then `genie-main` so that you are ready to run the model (the model is **always** run from `cgenie.muffin/genie-main`).

A restart can be requested in running an experiment by setting the 5th and last (optional) parameter when entering in the `runmuffin.sh` command. A spin-up of the climate state is provided: `LAB_0.SPIN` which you have just unpacked to the `cgenie_output` results output directory. Create a new (user config) experiment configuration file by using the given file `LAB_0.EXAMPLE` as a template (no parameter changes need to be made yet). You can make copies (`cp` command) of the experiment configuration files (e.g. `LAB_0.EXAMPLE`) and assign them different names, e.g., `twilight`, `rarity`, `applejack` ... (ideally, use a name that helps you remember what the experiment did). Or ... if you and the linux command lines are simply not BFFs, you can drag the user-config file (`LAB_0.EXAMPLE`) you want to use as a template to your local PC (/Mac) directory in the file transfer window, rename it either in the file transfer program or e.g. via the Windoz file-manager

(e.g. to LAB_0.NEW), and drag it back to the cluster directory again. You specify the use of the restart state by adding the restart experiment name as the 5th parameter, e.g.:

```
$ ./runmuffin.sh cgenie.eb_go_gs_ac_bg.worbe2.BASE LABS LAB_0.NEW 100 LAB_0.SPIN
```

The run-time output should now look noticeably different. There should be no (or perhaps just very little) drift in any of the various variable values outputted to the screen – this is because you have (re-)started from the end of a run that had already reached an equilibrium, steady-state.

Brrrrrrrrrrr – it's chilly on ... snowball Earth!

Further ideas

- Feedback loop analysis
- Continental configuration
- Geothermal heat input
- Seasonality



2. snowball Earth

Stuff to keep in mind:

- Models ARE NOT the 'real World' (it is going to be pretty obvious this is the case here).
- Don't believe what you read in Nature or Science.

Readme

You will need to download a new restart file prior to embarking on the snowball Earth experiments. To fetch this: change to the `cgenie_output` directory, and type (or copy and paste carefully from the PDF ...):

```
$ wget http://www.seao2.info/cgenie/labs/AWI.2017/LAB_1.SPIN.tar.gz
```

This downloads an archived/compressed copy of the experiment `LAB_1.SPIN` – effectively, just an experiment (spin-up) that has been run for 5,000 years for you. Extract the contents of this archive by typing:

```
$ tar xfvz LAB_1.SPIN.tar.gz
```

A new experiment results directly will then appear as if you had just run the entire 5,000 year experiment yourself, and you could in fact have done so (remember to refresh the **WinSCP** directory view window).

You'll then need to change directory back to `genie-main` to run the model.

2.1 Brrrrrrrrrr – it’s chilly on ... snowball Earth!

To illustrate how ‘easy’ it can be to configure an Earth system model such as cGENIE and explore the behavior of the Earth system and its response to perturbation – you are going to induce an extreme cooling of climate and see what happens. Solar output was weaker during the late Neoproterozoic, a time when the Earth experienced a series (2 ish) of extreme glaciations. Thus, having a mild climate state to start with must have been dependent on sufficient CO_2 and/or CH_4 in the atmosphere and hence presumably highly elevated compared to the modern World, so sort of the opposite of the problem we have today ...

You are going to be running experiments similar to before and using the restart:

```
$ ./runmuffin.sh cgenie.eb_go_gs_ac_bg.woreq1.NONE LABS LAB_1.EXAMPLE 100 LAB_1.SPIN
```

but rather than use the provided experiment configuration file `LAB_1.EXAMPLE`, why not get into the habit of creating new and uniquely named *user-config* files (no harder than copying it and renaming it!). If you keep using the same experiment name, the results will be over-written each time. Also, be especially careful not to have 2 (or more) experiments running simultaneously with exactly the same name as they try and over-write each other in a somewhat entertaining way.

Overall: your task in this exercise will be to determine the radiative forcing (or pCO_2 equivalent) threshold required to drive the climate system into a full ice-covered ocean (snowball Earth) state. (Read the *Hyde et al.* [2000] paper.)

Useful 2-D (netCDF—Panoply) variables to view are surface air temperature and sea-ice extent (and/or thickness). Ocean surface temperature and salinity can be viewed in the 3-D NetCDF results file (apologies for ocean temperature being in units of K ...).

Time-series (ASCII .res files) are useful for providing simple mean indicators of global climate such as global ocean fractional sea-ice covered.

Note that the model configuration of an idealized super-continent, positioned symmetrically about the Equator, is pretty unrealistic. But the further you go back in time, the more uncertain it becomes as to exactly where and in what orientation the continents were. Sometimes modelers have to resort to somewhat idealized experiments if the uncertainties are too great. In addition, one can conduct sensitivity experiments to test whether the continental configuration is important to the results. For instance, *Hoffman and Schrag* [2002] discuss the potential importance of continental configuration, while the entire hypothesis of *Donnadieu et al.* [2004] rests on specific details of the continental configuration being realistic.

For this configuration, the solar constant is set weaker than modern to reflect the fact that the Sun’s output has increased with time and during the Neoproterozoic the solar constant would have been ca. 5

```
ma_genie_solar_constant= 1285.92
```

(For reference, the modern value is 1368 W m⁻².)

Other questions to think about with regards to numerical modeling (and this experiment) are:

- (Is the model configuration and experimental design ‘realistic’ ... ?)
- What is ‘missing’ in the model and what might the implications for your predictions and conclusions be? For example, there is no land-surface scheme (and hence no concept of ‘snow’) in this particular configuration.

- Are the simulations being run for sufficiently long? Why not if not (i.e., justify your choices of parameter values and experimental assumptions)? How might the results and conclusions be biased (if at all)?
- How would you test model predictions and your overall conclusions?
- How could the experimental design be improved?
- Etc.

To search for the atmospheric CO_2 concentration (or rather, radiative forcing equivalent) that would lead to a ‘snowball Earth’ state in the Neoproterozoic and answer the question: ‘How low does CO_2 have to be to trigger a ‘snowball’?’ you are going to edit the file that controls the specific details of the experiment. This is the user-config file. From the `genie-userconfigs/LABS` directory, open one of the snowball experiments in the SciTE text editor. At the top of the file you should see something like:

```
#
#
# --- CLIMATE -----
#
...
# scaling for atmospheric \(\text{CO}_{2}\) radiative forcing, relative to 278 ppm
ea_radfor_scl_co2=20.0
```

Each line that is not commented out (i.e., no #) contains a parameter of the format:

PARAMETER=VALUE

The value of each parameter can be edited to form a new experiment. (Additional parameter value specifications can also be added, or existing ones deleted.) In this example, the line:

```
ea_radfor_scl_co2=20.0
```

specifies a radiative forcing of climate by CO_2 equivalent to x20 modern ($20 \times 278 = 2560$ ppm). If you instead wrote:

```
ea_radfor_scl_co2=1.0
```

this would give you a modern (x1, or $1 \times 278 = 278$ ppm) radiative forcing. (Technically: pre-industrial rather than ‘modern’ per se.) Note: CO_2 is not being explicitly modeled in this experiment, but the long-wave radiative forcing associated with a specified concentration of CO_2 (in ratio to modern) is being set instead.

Edit the value of `ea_radfor_scl_co2` (lower or higher) and save the file. Re-run the experiment to see whether sea-ice extent is approaching a new steady state. You may want to try even longer simulations (than 100 years) if it becomes clear that the model is still far from steady-state. You can judge how close to equilibrium things have got by following (and/or plotting) the evolution of e.g., global surface air temperature or sea-ice extent (both time-series files).

HINT: By submitting the experiments to the cluster will allow you to run many experiments (tries of different radiative forcing values) simultaneously.

For each experiment you want to be assessing how far towards the Equator the sea-ice limit encroaches through some of the time-series and time-slice files or even the on-screen summary lines (assuming running interactively rather than via a job submission to the cluster queue). Informative time-series variables include (but not necessarily be limited to: atmospheric temperature and sea-ice cover. (Sea-ice thickness, on account of the simple physics in the model, low resolution and long time-step, can fluctuate a little in area and volume at times.)

For the time-slice data: atmospheric and ocean surface temperature and sea-ice extent (2-D biogem NetCDF file) may be informative.

HINT: Be careful with the 'Fit to data' scaling feature in Panoply – at near complete sea-ice cover, you may find Panoply scaling min and max sea-ice between 99.1 and 99.9

In answering the question think about what an appropriate degree of accuracy might be for your experiments. Just because computer models generally calculate to around 16 significant places of precision, does not mean you have 16 significant figures of accuracy (or realism). For instance – how many significant figures is the solar constant quoted to and what do you think is the uncertainty in this? Harder to judge is how the assumed (incorrect) continental configuration creates additional uncertainty, or the simple physics assumed in the ocean or sea-ice, or lack of snow on land ...

Once you are happy about the controls on the snowball threshold try and answer:

How high does the (CO_2) radiative forcing have to be in order to escape from a snowball?

If you run the model with an appropriate radiative forcing to create a snowball, you can use that experiment as a restart and be able to carry out a series of experiments with increasing radiative forcing, all starting from the same snowball climate state you have created. Defining the radiative forcing / climate path going out of a snowball would complete the hysteresis loop of Hyde et al. [2000]. Note that a good restart is one for which the experiment did not sit too long in the snowball state before finishing (the more sea-ice thickness you create in the first experiment, the more you are going to have to melt in the next ...). You can fine-tune the number of years the experiment is run for to achieve this. (You cannot select when the re-start is saved – it is always saved at the end of an experiment.)

HINT: If you are having trouble deciding whether or not the snowball is heading in the right direction (towards an exit) because sea-ice is always reported at 100

Overall: think critically about the model configuration, the experimental design, and the nature of the scientific question (based on your background reading of snowball Earth). Some of the exploration/testing suggestions (above) may not necessarily give substantially different results. Such a finding would be as valid and interesting as determining an important dependence of a certain assumption, and would for instance indicate that the associated paleo uncertainties are not critical to model assessment of the question.

Always be prepared to justify all your choices for experimental design and model settings, e.g., range of radiative forcing assessed, continental configuration(s), solar forcing, use of re-starts (if any), run duration, etc. etc. etc.

2.2 Further ideas

2.2.1 Feedback loop analysis

In order to quantify the snowball Earth hysteresis loop in cGENIE as per Figure 2 in Hyde et al. [2000] you will need to extract from the model ‘meaningful’ measures of climate (e.g., global surface air temperature, fractional sea-ice coverage) as a function of CO_2 multiples, CO_2 concentration, or (better) radiative forcing. For the latter, in cGENIE, the radiative forcing for a doubling of CO_2 is set at: 5.77 W m^{-2} . See: Myhre et al. [1998] (Geophys. Res. Lett. 25, 2715–2718) and/or IPCC [2007] for more on what radiative forcing is and how it is related to a relative change in CO_2 concentration. Also, for making a comparison with Hyde et al. [2000] for going into the snowball, note that they plot the change in radiative with a ‘cooling’ as positive (a bit daft). Their baseline radiative forcing state (an anomaly of 0 W m^{-2}) you might assume is equivalent to 278 ppm and hence 130 ppm is an approximately halving of CO_2 and hence creates 5 W m^{-2} of cooling. (You might prefer to plot the radiative forcing change as warming being positive, which makes rather more sense ...)

For coming out of a snowball, because the CO_2 and hence radiative forcing threshold is so high as compared to going in, you may want to be creative in the plotting (assuming attempting to combine both thresholds into a single plot) and, for instance, one might break the scale between the low radiative forcing interval spanning going in and the high one spanning coming out.

Another example is as per Figures 3,4 in Stone and Yao [2004] (Clim. Dyn. 22, 815–822) (although here it is the solar constant rather than long-wave radiation forcing that is being varied). So in fact, you could try varying the solar constant as an alternative to radiative forcing and hence be able to come up with a plot directly comparable to Stone and Yao [2004].

2.2.2 Continental configuration

It was mentioned earlier that the position of the continents is an area of modelling uncertainty and might be important. You can test for this. Four alternative base-configs are provided which each define a different continental configuration:

1. `cgenie.eb_go_gs_ac_bg.wopol1.NONE`
– a single polar super-continent, with an ocean resolution of 36×36 with 8 vertical levels. (Note potential ‘1’ and 1’1 confusion in ‘wopol1’.)
2. `cgenie.eb_go_gs_ac_bg.wopol2.NONE`
– one continent at each pole, with an ocean resolution of 36×36 with 8 vertical levels.
3. `cgenie.eb_go_gs_ac_bg.woreq1.NONE`
– a single Equatorially-centred super-continent, with an ocean resolution of 36×36 with 8 vertical levels. [current configuration]
4. `cgenie.eb_go_gs_ac_bg.woreq2.NONE`
– two continents straddling the Equator, with an ocean resolution of 36×36 with 8 vertical levels.

You can use the given user-config file (LAB_1.EXAMPLE) as an experiment template and any of the alternative configurations can be run very similarly to as per before, i.e.:

```
$ ./runmuffin.sh cgenie.eb_go_gs_ac_bg.xxxxx.NONE LABS LAB_1.EXAMPLE 100
```

Note that you are using a different base-config file name: `cgenie.eb_go_gs_ac_bg.xxxxx.NONE` where xxxxx is one of: `wopol1`, `wopol2`, `woreq1`, or `woreq2`. Also note that no restarts are provided

for these configurations. You may (or may not) want to create some (you will need to judge for yourselves how long to run the restart experiments for to achieve as close to steady-state as you think is ‘sufficient’). Remember that restarts are just ‘normal’ experiments that have already been run. Be careful that when changing from one base-config to another, the model re-compiles. Simply running the new configuration briefly is sufficient to ensure this. Experiments can then be safely submitted to a cluster queue. i.e. do not try and submit an experiment using a different base-config straight to the cluster queue without having run it (or a short version of the experiment you want) interactively first (to ensure the model is re-compiled). This is also good practice – checking that a new sort of experiment and/or model configuration works as you intend and without hiccups.

2.2.3 Geothermal heat input

Finally, cGENIE will fairly happily build up sea-ice, apparently without limit (with the remaining wet ocean becoming progressively colder and more saline). In the real world, one might expect some sort of limit to the maximum thickness achieved as the heat diffusion across a progressively greater thickness of sea-ice approaches the heat input at the bottom of the ocean from geothermal energy. Different modes of ocean circulation are also possible if one considers heating from the bottom as well as cooling (and brine rejection) from the top and which might affect the entry into or exit from a snowball state.

In the experimental setup you have been given, a geothermal heat input is specified in the ocean circulation module via the following parameters:

```
bg_ctrl_force_GOLDSTEInTS= .TRUE.  
bg_par_Fgeothermal=100.0E-3
```

The first enables the temperature and/or salinity of the ocean to be modified by processes other than redistribution via ocean circulation and exchange with sea-ice and atmosphere (i.e. to impose external forcings). The second sets the geothermal flux in units of W m^{-2} . (Note that in the Neoproterozoic, the geothermal heat flux could have been somewhat higher than modern. How much? A question for Google...)

An appropriate research question might be to determine in radiative forcing vs. geothermal space (and requiring a 2D grid of parameter combinations to be created and submitted to the cluster): the equilibrium sea-ice thickness and region in which a snowball solution is not possible. However, more simply and suitable to a short workshop: How much of a difference to the estimated entry and exit thresholds of radiative forcing, does the inclusion of a geothermal input make? e.g. what happens if you set it to zero? What about 10 times modern (or more, although *extreme* seafloor heating can cause numerical instability)?

2.2.4 Seasonality

By default, this configuration is non-seasonally forced (by solar insolation). You can switch to a seasonally-forced to model by adding the following lines to the *user-config* file:

```
ea_dosc= .true.  
go_dosc= .true.  
gs_dosc= .true.
```

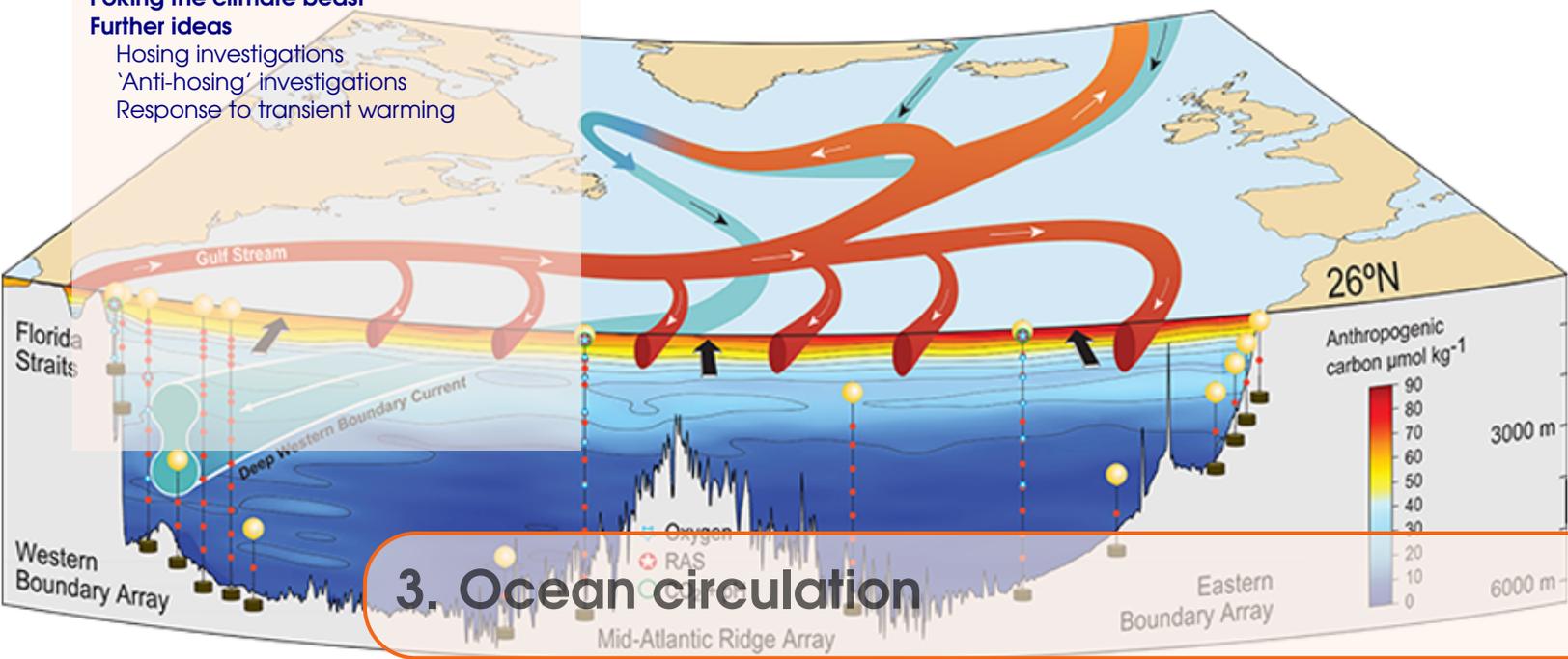
The scientific question here in trying this would be whether or not taking into account a seasonally-varying climate substantially affects the entry (and/or exit) thresholds for a snowball climate state.

(At least, whether it is important in the context of the resolution and physics of the model you are using.)

You can also save the data seasonally if you like – see Section 5.2.3 in the User Manual (your configuration has 24 time-steps per year for the BIOGEM module for reference).

Tracing ocean circulation
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Hosing investigations
'Anti-hosing' investigations
Response to transient warming



3. Ocean circulation

Stuff to keep in mind:

- Nothing at all – keep your mind completely empty and let the wonderful truths of **GENIE** permeate your entire being.

Background reading (Atlantic circulation and stability in GENIE):

- Hargreaves et al. [2004] (Climate Dynamics 23, 2004, Pages 745 – 760)
→Simple assessment of the likelihood of AMOC collapse.
- Marsh et al. [2004] (Climate Dynamics, 23 2004, Pages 761 – 777)
→Characterization of thresholds of AMOC collapse.
- Singaraye et al. [2008] (GRL 35, doi:10.1029/2008GL034074)
→Role of changing ocean circulation in atmospheric radiocarbon variability during the Younger Dryas.

Background reading (Miscellaneous (model) Atlantic circulation and stability):

- Rahmstorf et al. [2006] (In: Encyclopedia of Quaternary Sciences, Edited by S. A. Elias. Elsevier, Amsterdam)
→Provides the background to the Atlantic Meridional Overturning Circulation and hypothesized hysteresis.
- IPCC [2007] (e.g., Section 10.3.4)
→Future predictions of AMOC strength.
- Schmittner [2005] (Nature 434, 628– 633)
→Impacts on marine ecosystems and carbon cycling.
- Obata [2007] (J. Clim. 20, 5962–5976)
→Climate-carbon cycle model response to freshwater discharge.

README

You will need to download a new restart file prior to embarking on the experiments with modern ocean circulation. To fetch this: change to the `cgenie_output` directory, and type (or copy and paste carefully from the PDF ...):

```
$ wget http://www.seao2.info/cgenie/labs/AWI.2017/LAB_2.SPIN.tar.gz
```

This downloads an archived/compressed copy of the 10,000 year *spin-up* experiment `LAB_2.SPIN`. Extract the contents of this archive by typing:

```
$ tar xfv LAB_2.SPIN.tar.gz
```

You'll then need to change directory back to `genie-main` to run the model.

3.1 Tracing ocean circulation

The ocean biogeochemistry module (**BIOGEM**) in **cGENIE** provides a framework for applying time- and spatially-variable ‘forcings’ of the Earth system – fluxes or restored-to boundary conditions that can be prescribed for any gas, dissolved substance (including temperature and salinity), or particulate matter. Examples include freshwater input (== a negative salinity flux forcing) of the North Atlantic to alter ocean circulation, fossil fuel CO_2 emissions to the atmosphere (== a CO_2 gas flux forcing), or aeolian iron supply to the surface ocean (a 2-D dust flux forcing).

For example: view the user-config file: `LAB_2.colorinjection` – you will see the following lines (under the heading: ‘# -- FORCINGS --’)

```
bg_par_forcing_name="pyyyz_Fred"
bg_par_force_point_i=22
bg_par_force_point_j=33
bg_par_force_point_k=8
bg_par_ocn_force_scale_val_48=0.0
```

The first line points **cGENIE** to a directory located in `cgenie.muffin/genie-forcings` that contains a set of files that define what geochemical property is going to be altered plus information about how the magnitude of the forcing changes with time.

There are then three lines (`bg_par_force_point_i=20, ...`) that specify the location in the ocean of the geochemical forcing is going to be applied. The point sources are specified in (i,j,k) coordinates, which in this case is (22,33,08). For the ocean model resolution we are using, the grid is 36x36x16, longitude (i) is counted from left-to-right (1 to 36); latitude (j) is counted from bottom-to-top (1 to 36); level depth (k) is counted from downwards top-to-bottom (16 down to 1). Thus, (22,33,08) is a release of tracer in the North Atlantic, a little south of Greenland, and intermediate depth (level = 8 out of 16). Refer to the Figures for how the horizontal (Figure 3.1) and vertical (Figure 3.2) grid is specified.

Finally, there is a scaling parameter (`bg_par_ocn_force_scale_val_48`) which modifies the magnitude of the flux to be applied (in units of $molyr^{-1}$).

(Section 4 in the User Manual describes the original and most flexible provision for applying time-dependent *forcings*.)

You are going to run a brief experiment in which you will be injecting a conservative ‘dye’ tracer in the ocean. BIOGEM has two tracers defined for this purpose – ‘blue’ and ‘red’. Open the user-config file: `LAB_2.colorinjection` and edit the parameter controlling the flux of red dye to read:

```
bg_par_ocn_force_scale_val_48=1.0E12
```

which specifies a flux of 1.0×10^{12} (mol yr⁻¹) rather than zero as given.

The *base-config* you will be using is different from previously: `cgenie.eb_go_gs_ac_bg.worjh2.rb` – this specifies a 16 vertical levels ocean and also includes seasonality of solar insolation.

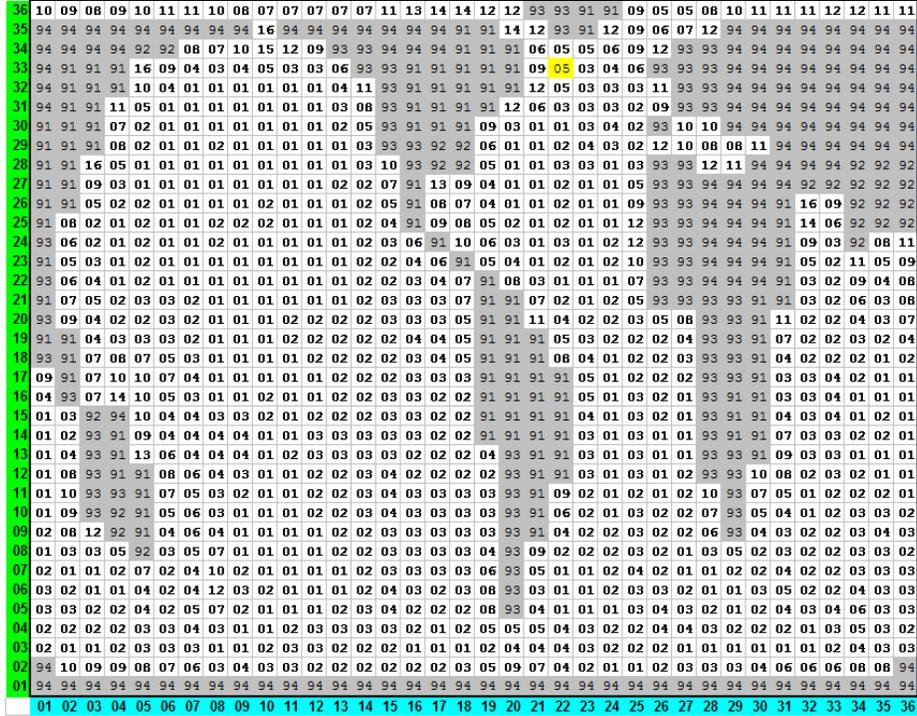


Figure 3.1: **cGENIE grid (36 × 36 ‘worjh2’ configuration).** Light blue numbers are the ‘i’ co-ordinates. Green numbers are the ‘j’ co-ordinates. The depth of the ocean at any location is indicated by its ‘k’ value – a number between 1 and 16, with 16 being the surface layer of the ocean, and 1 the maximum possible depth anywhere. Numbers > 90 (91, 92, 93, 94) and shaded grey are land (and specify the direction of run-off). Location (22,33,08) is highlighted in yellow. The longitude of the western edge of the grid is at 260W, and the increments are 10 degrees.

k	mid depth (m)	base of layer (m)
16.00	38.91	80.84
15.00	126.04	174.75
14.00	227.26	283.85
13.00	344.84	410.58
12.00	481.44	557.80
11.00	640.12	728.83
10.00	824.45	927.51
9.00	1038.59	1158.31
8.00	1287.35	1426.43
7.00	1576.33	1737.90
6.00	1912.04	2099.73
5.00	2302.02	2520.05
4.00	2755.05	3008.34
3.00	3281.33	3575.57
2.00	3892.71	4234.52
1.00	4602.92	5000.00

Figure 3.2: **cGENIE ocean vertical level definitions.**

Run the model for ... whatever, 20 years will do. Use the restart experiment you have just downloaded to start from:

```
$ ./runmuffin.sh cgenie.eb_go_gs_ac_bg.worjh2.rb LABS
  LAB_2.colorinjection 20 LAB_2.SPIN
```

View the results – how the Red tracer distribution evolves – in the time-slice files (full ocean (/atmosphere) properties saved in the netCDF format (.nc) files). You can follow the progress of the dye (and hence diagnose the properties of ocean circulation in the model) by plotting vertical and/or horizontal slices that go through (or near) the cell location in which you inject the dye tracer in the 3D netCDF file. Note that Panoply appears to ‘count’ the ocean layers in the opposite direction to the way in which the ocean model is actually counting them – the correct definition is with ‘1’ being very deepest level possible (and as displayed in the figure).

You can also view the tracer distributions in terms of a water-column integrated tracer inventory (netCDF variable name: `ocn_int_colr`; long name: `colr water-column integrated tracer inventory`) in the 2D netCDF output. (See: Sabine et al. [2004] for the use of water column integrals in the context of the distribution of anthropogenic CO_2 uptake and storage.) Changes in tracer inventory with time can be tracked in the time-series file

```
biogem_series_ocn_colr.res.
```

You can also plot the overturning circulation from the 2D netCDF file – variable `phys_opsi == global overturning streamfunction`, `phys_opsia == overturning in the Atlantic` to provide a visualization of the large-scale ocean circulation that drives tracer movement.

Spend a little while altering the flux (`bg_par_ocn_force_scale_val_48`) and/or location (`bg_par_force_point_i`, `bg_par_force_point_j`, `bg_par_force_point_k`) of tracer input. Note how you can use ‘tracers’ to help diagnose (and better understand) the circulation of the ocean.

An interesting (honest!) and illustrative exercise is to use the dye tracer to pick out the path taken by Mediterranean Intermediate Water. Despite the low resolution of the cGENIE ocean circulation model component and the highly restricted representation of the Mediterranean, the model does predict a salty Mediterranean as a consequence of P-E in this basin (and its catchments) being negative and this water makes its way out in the subsurface into the Atlantic.

Simply specify a dye injection somewhere in the Mediterranean (be careful with the restricted depth of the Mediterranean – if you inject too deeply (into the crust!) then you will not see anything (refer to the figure for the depth level (k) number of the maximum depth of the water column in each location), and it is better to inject it relatively close to the opening of the gateway (try some different locations and see which ones produce a reasonably instructive tracing of Mediterranean outflow). Run for e.g. 20 or 50 years (from the provided spin-up). Then:

1. View the dye-tagged plume of Mediterranean Intermediate Water by plotting a lat-lon slice (from the 3D netCDF file). This will give you the depth of the plume. How does this compare with salinity observations (salinity observations and appropriate global datasets can be found on the web with a little patience)? You can also view the water-column integrated distribution (2D netCDF).

2. Try viewing the plume via a lat-depth slice. Refer to the figure to determine the 'i' value up the Atlantic that will just graze the edge of what passes for Spain at this low model resolution. Which direction does it head after exiting the Mediterranean? Is this 'realistic'?

3.2 Poking the climate beast

Instead of adding a dye tracer, you could add fresh water to assess the sensitivity of the Atlantic Meridional Overturning Circulation (AMOC) to collapse in a classic ‘hosing’ experiment.

The user-config file for this is called: LAB.0001.hosing. The default (i,j) location of the flux input is the same (as the dye tracer), but now the injection at the surface (level: k=16). Note that the forcing of the salinity tracer is negative (freshwater = negative salinity compared to sea-water)!

To orientate you in freshwater forcing space: `bg_par_ocn_force_scale_val_2=-2.0E17` should be sufficient to make ‘stuff happen’ and quickly. BUT, this is a pretty extreme flux (see overleaf for a rough conversion between salinity forcing units (mol yr⁻¹) and fresh water flux (in m³ s⁻¹ or Sv). Much more than this and the model may crash or at the very least, you’ll be left with a large freshwater pond in the North Atlantic ... (See later (Section 1.6e) for some exciting discussion on units!)

To run the model for e.g. 20 years using the same restart:

```
$ ./runmuffin.sh cgenie.eb_go_gs_ac_bg.worjh2.rb LABS
LAB_2.hosing 20 LAB_2.SPIN
```

20 years should be long enough to see a collapse start to occur, but you might want to run the model for longer (and it can be submitted as a job, of course). Running for longer will also allow you to have a smaller, less extreme (and maybe more realistic) freshwater input flux.

The most obvious property of the Earth system to follow is the Atlantic overturning strength (`biogem_series_misc_opsi.res`). The AMOC stream-function (in `fields_biogem_2d.nc` 2-D time-slice netCDF results file, field: `phys_opsia`) is also illustrative. You can also try and identify the salinity anomaly (see below) due to freshwater input in the 3D salinity tracer field.

There are also important impacts on surface air temperatures and maybe sea-ice extent (in `fields_biogem_2d.nc`). Note the importance (sort of) of the AMOC in transporting heat to the N Atlantic region (the film *the Day After Tomorrow* was not entirely inaccurate in this particular respect). Be aware of the possibility of climate impacts far from the location of fresh water forcing. Look out for any significant-looking impacts on sea-ice extent, etc.

Note that as the model is running rather *s l o w e r* than in the snowball configuration, you might want to think carefully of making use of cluster queuing possibilities (i.e. running multiple experiments at once in the background).

To more easily assess some of these impacts (and for other sorts of analysis) it is possible to create an anomaly (difference) map in Panoply:

1. First open a dataset, e.g., `atm_temp` (surface air temperature) in the 2D netCDF file. You can either double-click the variable name, or, with the variable name highlighted, click the ‘Create Plot’ icon.
2. Now, with the `atm_temp` still selected (and the first plot window still open), click on the ‘Combine Plot’ icon. A dialogue box will appear and ask you to select a plot to combine the new one with. Make sure the name of your first plot window is selected/highlighted. Click ‘Combine’. OR, simply drag a second dataset into the plot window of the first dataset.

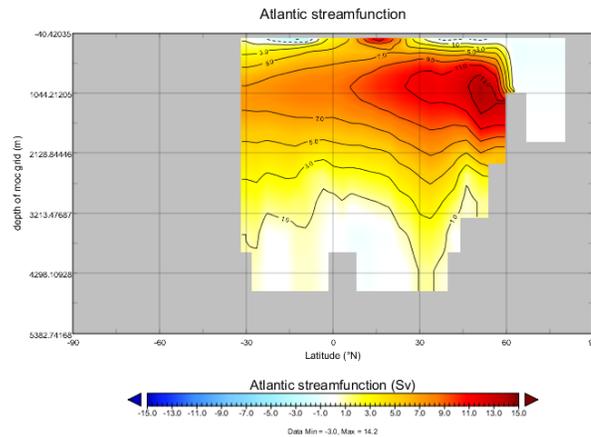


Figure 3.3: Example plot of (normal/default modern) overturning streamfunction (2D netCDF file). (e.g. for Atlantic: netCDF parameter name: phys_opsia, long-name: Atlantic streamfunction). Note that autoscaling has been turned off and the min and max plotting limits set manually. By convention, streamfunctions are plotted with their scale symmetrical around zero, giving red and ‘warm’ colors for positive value and clockwise overturning, and blues and ‘cold’ colors for negative values and anti-clockwise overturning. Also shown is the same for the Pacific. (The plots have been tart-ed up by overlaying solid contours plus contour labels.)

3. You now have a plot window that by default it is showing you the difference between two identical (in time) slices. The two different slices are labeled Array 1 (LH side) and Array 2 (RH side).

Keep one array (Array 1) fixed to the initial (year 1 (centered on 0.5)) and vary the year in the second array (Array 2). Note that you can select in Panoply whether Array 1 – Array 2 is plotted, or Array 2 – Array 1, or various proportional or relative differences.

Note that you can switch off the auto-scaling feature (Always fit to data) and center the scale so that no change is white, with positive deviations = red and negative = blue by clicking on Center on 0 (an often used convention in climate field plotting).

2 example plots (using Panoply) are shown in Figure 3.3 and Figure 3.4.

You can also plot ocean current fields which is sort-of fun and maybe even informative(!):

1. In the 3D netCDF file, the three components of ocean velocity are represented by the variables: ocean velocity – u (Eastwards), ocean velocity – v (Northwards), and ocean velocity – w (upwards). 2. Open up velocity – u. Chose ‘lon-lat’.
2. Select/highlight velocity – v. and click on the ‘Combine Plot’ icon (as per before).
3. Rather than a difference map, which is what you get by default, i.e. ‘Array 1 – Array 2’ – from the drop-down menu (next to the ‘Interpolate’ button) select ‘Vector Magnitude’.
4. You should have a color contoured (or not if you prefer plotting without contouring on) map of ocean current speed, with velocity vectors (direction and magnitude) overlain. You’ll need to re-scale the velocity vectors to properly see them – from the ‘Contours and Vectors’ tab – change the ‘Scale Length’ to e.g. 0.1. When fresh-water hosing – look out for impacts on the

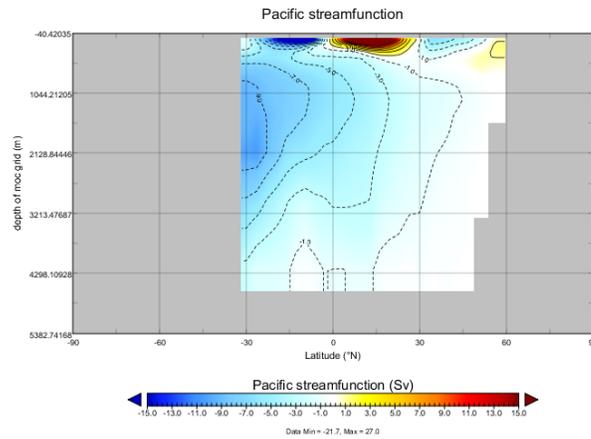


Figure 3.4: Example plot of collapsed AMOC.

N. Atlantic current system associated with the AMOC.

5. You can repeat this for deeper depth levels in the ocean – e.g. between about 1500 and 2000 m is a good place to go looking for the Western boundary current (and AMOC return) in the model (such as it exists at this low resolution) but you’ll need to re-scale the velocity vectors again (e.g. to 0.01 to less).

2 example plots (using Panoply) are shown in Figure 3.5 and Figure 3.6.

Finally, a brief note on units ... the freshwater forcing is implemented as negative salinity, just to really screw with your mind. The generic internal **cGENIE.muffin** model units for the forcing end up as $PSU kg^{-1} yr^{-1}$. Which sort of does not make much sense ...

Start, by thinking of a value of `bg_par_ocn_force_scale_val_2` of -34.9 as equivalent to taking all the salt out of $1kg$ of freshwater (since the mean global salinity is $34.9PSU$). Or equivalently, since the ocean volume is fixed, an applied forcing value of -34.9 is equivalent to adding $1kg$ of freshwater to a (surface) box. So, a value of `bg_par_ocn_force_scale_val_2` of -3.49×10^4 ($-3.49E04$) would be a flux of $1m^3 yr^{-1}$ ($1000kgm^{-3}$) of freshwater.

So, in the example earlier (`bg_par_ocn_force_scale_val_2=-1.0E18`), the freshwater flux is $1.0 \times 10^{18} / 3.49 \times 10^4 = 2.8653 \times 10^{13} m^3 yr^{-1}$.

The literature invariably gives freshwater fluxes in units of Sv ($10^6 m^3 s^{-1}$). So in the example, the freshwater flux is: $9.0797 \times 10^5 m^3 s^{-1}$ ($365.25 \times 24 \times 3600 = 31557600syr^{-1}$). Or $0.9Sv$. Read the literature ... but generally, fluxes of ca. $0.05Sv$ and larger (and to quite specific places) are applied in models in order to induce a collapse of the AMOC.

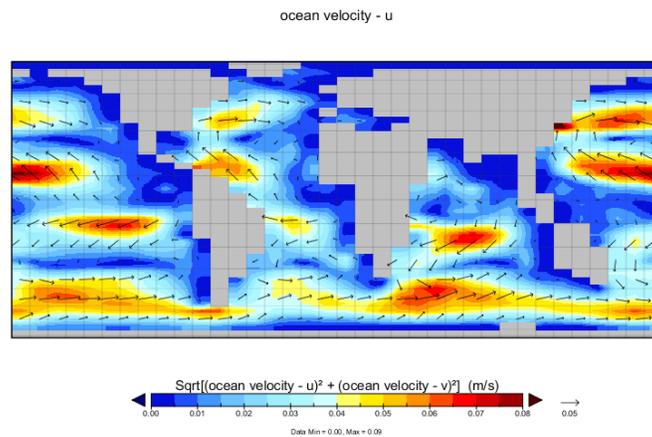


Figure 3.5: Example plot of (normal/default modern) ocean current fields (3D netCDF file). Again scaling has been set manually to create an easy-to-interpret axis scale. On the left is the surface field, and on the right an intermediate depth (illustrating what approximates the Deep Western Boundary current in the model in the Atlantic).

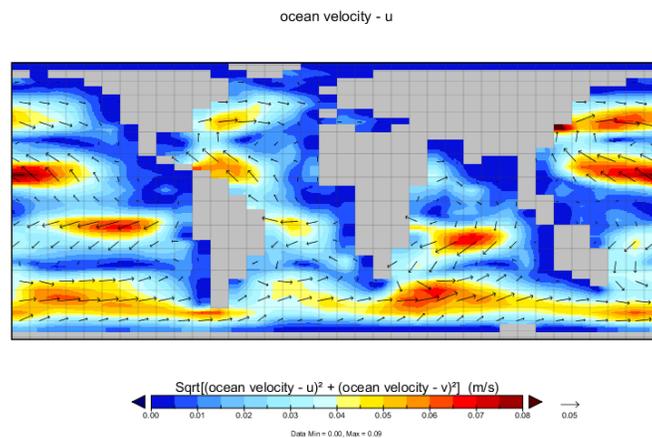


Figure 3.6: Example plot for collapsed AMOC.

3.3 Further ideas

3.3.1 Hosing investigations

What is the largest freshwater flux that can be sustained without ‘collapsing’ the AMOC? Is there a ‘threshold’ (‘tipping point’) of freshwater input, beyond which the AMOC rapidly decreases in strength?

Is the precise location of the freshwater input important (i.e., try tipping it in somewhere else)? What would you expect to see in the paleo (e.g., ice core) record of both hemispheres if such a shutdown occurred in the past?

Are any other major regions of deep water formation (where are they) sensitive to freshwater perturbation and what are the consequences (could it happen in the future)?

3.3.2 ‘Anti-hosing’ investigations

There are questions concerning past changes in the AMOC as to whether it is ‘pushed’ or ‘pulled’. i.e. if the AMOC shoals in depth and/or weakens, is it because its production has weakened, or as Antarctic Bottom water (AABW) strengthened and ‘pushed’ it out of the way (to shallower depths)?

What you might try then is to inject salt in the Southern Ocean as opposed to fresh water in the North Atlantic. All you need do is pick an appropriate grid point (this is worth thinking about carefully and maybe testing different locations) and rather than giving the parameter `bg_par_ocn_force_scale_val_2` a negative value, you give it a positive one. (Start by trying similar magnitudes of value as before and see what happens.)

Is the AMOC (for the same magnitude of forcing) more sensitive to being ‘pushed’ or ‘pulled’? (Obviously the answer will very much depend on where the perturbations are being applied.)

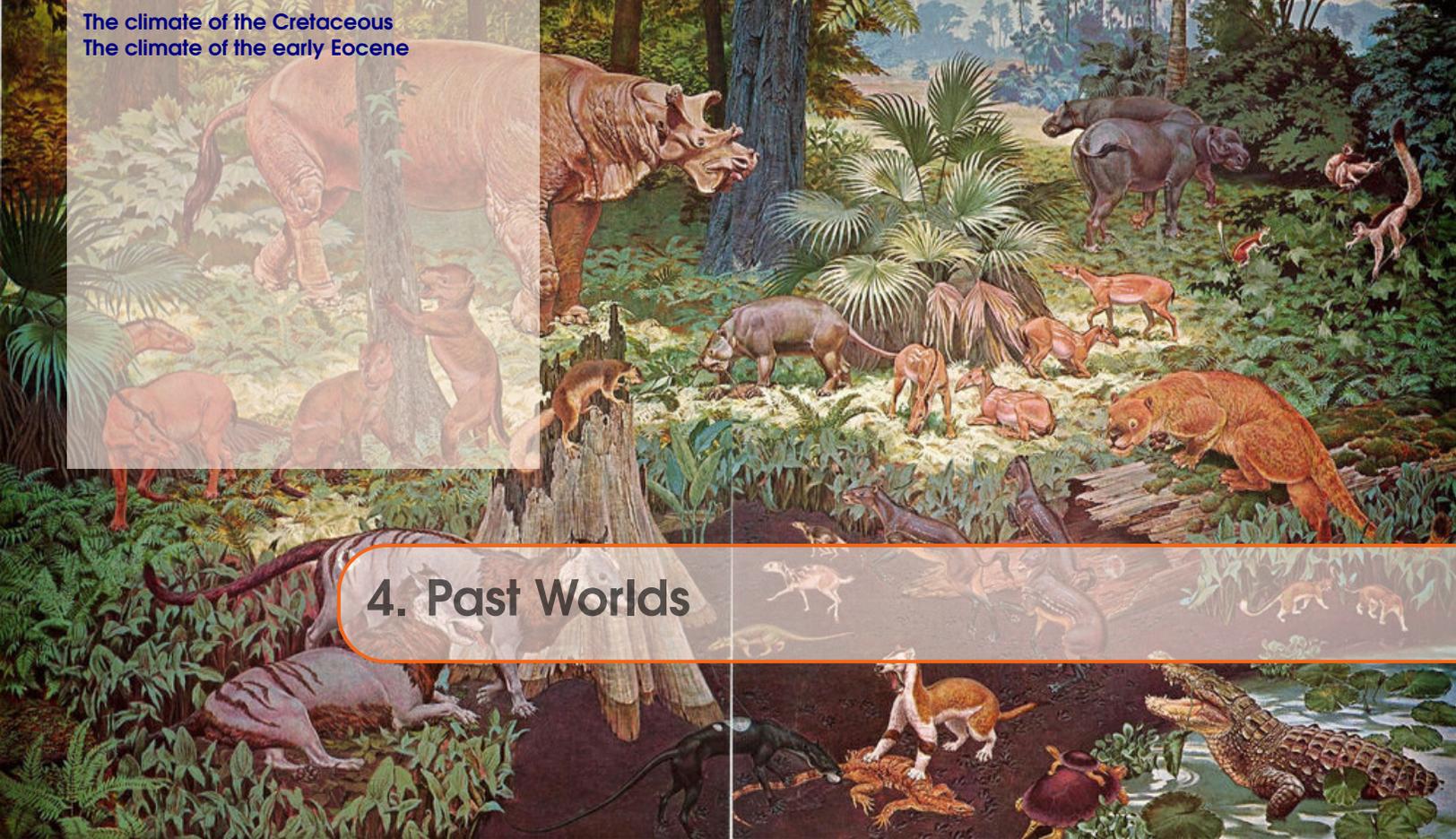
3.3.3 Response to transient warming

A current concern regarding global warming is the ocean circulation response to a strong warming of the surface, as it is assumed (and demonstrated in models) to result in surface stratification of the ocean, likely restricting the nutrient supply to phytoplankton and reducing ventilation of the ocean interior with dissolved oxygen.

You can explore the transient response of ocean circulation to warming by simply adjusting the radiative forcing parameter used in the snowball Earth experiments: `ea_radfor_scl_co2`. By default in the modern continental configuration it has a value of 1.0, corresponding to 278 ppm atmospheric CO_2 . A value of 2.0 would reflect warming equivalent to 556 ppm CO_2 . And 3.0 more like an end-of-the-century warming. Note that you are applying the warming instantaneously by manipulating the climate system in this way and hence the changes will be more extreme than those occurring over the time-scale of this century. Also note that a cooling could be applied instead. A user-config – `LAB_2.EXAMPLE` – is provided as a template for these experiments.

Potentially interesting properties of the Earth system to look at include sea-ice extent and AMOC strength (in the ASCII time-series files), and the overturning streamfunction and sea-ice extent (2-D netCDF output).

How much radiative forcing is required to collapse the AMOC? What atmospheric CO_2 value does this approximately correspond to?



4. Past Worlds

This Chapter comprises 2 different exercises, both employing 'realistic' reconstructions of past continental configuration and climate and designed to explore some of the key controls on global surface climate as well as the spatial pattern of surface temperatures.

The two exercises concern the late Cretaceous and early Eocene climate states, and are somewhat interchangeable.

Background reading (Cretaceous climate)

- Barron, E.: A warm equable Cretaceous: the nature of the problem, *Earth-Science Reviews* 19, 305–338, 1983.
- Bice, K. L., and R. D. Norris, Possible Atmospheric CO_2 extremes of the middle Cretaceous (late Albian-Turonian), *Paleoceanography* 17, doi: 10.1029/2002PA000778, 2002.
- Bice, K. L., B. T. Huber, and R. D. Norris, Extreme polar warmth during the Cretaceous greenhouse?: Paradox of the Late Turonian $d18O$ record at DSDP Site 511, *Paleoceanography* 18, doi: 10.1029/2002PA000848, 2003.
- Bice, K. L., D. Birgel, P. A. Meyers, K. A. Dahl, K. Hinrichs, and R. D. Norris, A multiple proxy and model study of Cretaceous upper ocean temperatures and atmospheric CO_2 concentrations, *Paleoceanography* 21, PA2002, doi:10.1029/2005PA001203, 2006.
- Donnadieu, Y., et al., Modelling the primary control of paleogeography on Cretaceous climate, *Earth and Planetary Science Letters* 248, 426–437, 2006.
- Huber, B. T., Norris, R. D., and MacLeod, K. G.: Deep-sea paleotemperature record of extreme warmth during the Cretaceous, *Geology* 30, 123–126, 2002.
- Hunter, S. J., et al., Modelling Maastrichtian climate: investigating the role of geography,

atmospheric CO_2 and vegetation, *Clim. Past Discuss.* 4, 981–1019, 2008.

- Jenkyns, H. C., Forster, A., Schouten, S., and Sinninghe Damste, J. S.: High temperatures in the Late Cretaceous Arctic Ocean, *Nature* 432, 888–892, 2004.

Background reading (Eocene climate)

- Dunkley Jones, T., D.L. Lunt, D.N. Schmidt, A. Ridgwell, A. Sluijs, P.J. Valdes, and M. Maslin, Climate model and proxy data constraints on ocean warming across the Paleocene-Eocene Thermal Maximum, *Earth-Science Reviews* 125 123–145 (2013).
- Lunt, D. J., Dunkley Jones, T., Heinemann, M., Huber, M., LeGrande, A., Winguth, A., Loptson, C., Marotzke, J., Roberts, C. D., Tindall, J., Valdes, P., and Winguth, C.: A model–data comparison for a multi-model ensemble of early Eocene atmosphere–ocean simulations: EoMIP, *Clim. Past*, 8, 1717-1736, doi:10.5194/cp-8-1717-2012, 2012.

READ.ME

You will need to download new *restart* files for this Chapter (both 10,000 year spin-ups). These are:

Cretaceous: \$ wget http://www.seao2.info/cgenie/labs/AWI.2017/LAB_5.SPIN.tar.gz

Eocene: \$ wget http://www.seao2.info/cgenie/labs/AWI.2017/LAB_5b.SPIN.tar.gz

Extract the results in the usual way and in the usual place ... and return to *genie-main*.

4.1 The climate of the Cretaceous

A previously spun-up state of Maastrichtian climate with **cGENIE.muffin** in a ca. 70 Ma configuration¹ (LAB_5.SPIN) is provided as a starting point. A *user-config* (LAB_5.EXAMPLE) that continues on this climate state is run:

```
$ ./runmuffin.sh cgenie.eb_go_gs_ac_bg.p0067f.NONE LABS
LAB_5.EXAMPLE 10 LAB_5.SPIN
```

Your task now is ... ‘simple’: Account for the Cretaceous reduced Equator-to-pole surface temperature gradient (compared to modern), particularly the apparently much warmer poles. Different data-based time-slices (including the Maastrichtian) are provided in *Huber et al.* [2002], although the low latitude $\delta^{18}O$ based temperatures are now not considered reliable. *Jenkyns et al.* [2004] contains high latitude (Arctic) data for the Maastrichtian. There are proxy-derived latitudinal temperature gradients and model-data studies for earlier in the Cretaceous – the problem is essentially the same.

The 2-D NetCDF results file contains the surface air temperature field (and sea-ice cover, if any). The 3-D NetCDF results file contains fields for ocean temperatures (and salinity). Both contain continental configuration and ocean bathymetry.

Panoply will plot the zonal average for you (as used in model-data comparisons – e.g., see Bice and Norris [2002]) – in the Array(s) tab, the Plot can be set to Zonal Averages rather than Map. You can get a smooth curve by selecting Interpolate. Remember you can set (and fix) scales rather than let Panoply auto-scale continually.

The following ‘controls’ over the climate system are provided to you in the form of a list of parameters at the bottom of the LAB_5.EXAMPLE *user-config* file for editing:

```
# --- MISC -----
#
# === ATMOSPHERE ===
# \{CO_{2}\} radiative forcing scaling factor [DEFAULT = 4.0]
ea_radfor_scl_co2=4.0
# CH4 radiative forcing scaling factor [DEFAULT = 1.0]
ea_radfor_scl_ch4=1.0
# Equator-to-pole different in planetary albedo [DEFAULT = 0.260]
ea_albedop_amp=0.260
# Baseline planetary albedo [DEFAULT = 0.200]
ea_albedop_offs=0.200
# atmospheric diffusivity of temperature (horizontal) [DEFAULT = 5.0e6]
ea_12=5.0e6
# === OCEAN ===
# ocean diffusivity of temperature + salinity (horizontal) [DEFAULT = 1494.4]
go_14=1494.4
# scaling for wind stress (set values of both identical) [DEFAULT = 1.531]
go_13=1.531
ea_11=1.531
```

Most of these parameters are associated with the radiative forcing of climate or atmospheric transports. The most useful ones are likely to be:

¹Note that for speed, no carbon cycle is selected in this configuration.

(i) The line:

```
ea_radfor_scl_co2=4.0
```

which specifies a radiative forcing of climate by CO_2 equivalent to 4 times modern CO_2 (i.e., $4 \times 278\text{ppm} = 1112\text{ppm}$) as per before (e.g. snowball Earth experiments).

The line:

```
ea_radfor_scl_ch4=1.0
```

specifies a radiative forcing of climate by CH_4 equivalent to 1 times modern CH_4 (i.e., $1 \times 700\text{ppb}$).²

(ii) cGENIE, as configured here, does not have a land surface scheme (no snow cover) nor clouds nor ice sheets, so a planetary albedo is prescribed (see Figure). This varies with latitude and is parameterized after a fully coupled GCM simulation. There is a parameter which controls how the albedo varies as a function of latitude, which can be adjusted:

```
ea_albedop_amp=0.260
```

However, if you vary this, why are you doing it? (What is the physical justification for giving the poles a higher or lower albedo relative to the Equator?) There is also a parameter that sets the baseline (minimum) albedo:

```
ea_albedop_offs=0.200
```

i.e., albedo is primarily a sum of the baseline value plus the Equator-to-pole slope (times the latitude).

(iii) The diffusivity of heat in the atmosphere (since it is a simple 2-D atmospheric model, with no atmospheric circulation, the atmosphere is made ‘diffusive’ to help capture heat and moisture transport) is:

```
ea_12=5.0e6
```

Note the maximum value the model can cope with is ca. $1.0E7$.

Run the model for however long you think is ‘necessary’ (/justified). The surface climate will approach equilibrium ‘relatively’ quickly. Deep ocean temperatures will typically take thousands of years to fully adjust ... You can assess how the model approaches equilibrium most easily from the atmospheric temperature time-series results file, and from the ocean temperature time-series results file (allowing to you to contrast surface and whole ocean temperature changes).

You can also try tracing and analyzing the patterns of ocean circulation in the Cretaceous world, in the same way as you did for the modern climate system. And in fact, it would be a useful exercise to directly compare your previous modern results vs. Cretaceous results, or even better, carry out a set of paired experiments (e.g. similar locations and fluxes of dye injection) for both modern and Cretaceous. (Note that if you do this and swap back and forth between modern and Cretaceous configurations, you cannot submit straight away to the cluster but must first briefly run

²But effectively, this is going to do exactly the same as changing radiative forcing due to CO_2 .

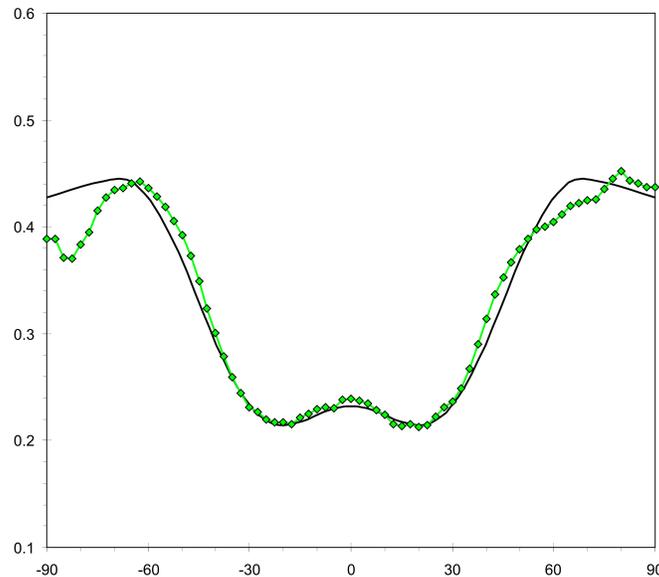


Figure 4.1: Prescribed planetary albedo. The latitudinal (from 90°S (-90°N) on the left, to 90°N on the right) profile of planetary albedo as calculated in a fully coupled GCM is given in green, and the cGENIE ‘fit’ in black.

the experiment at the command line in order that cGENIE is compiled into the new continental configuration.)

A new *user-config* (LAB_5.colorinjection) that continues on from this climate state, can be run:

```
$ ./runmuffin.sh cgenie.eb_go_gs_ac_bg.p0067f.rb LABS
LAB_5.colorinjection 10 LAB_5.SPIN
```

(Also note that the *base-config* is different as it now includes the definition of 2 dye tracers.)

The default locations for the dye release are set different compared to in the modern configuration. BUT, the default location is not necessarily ideal / particularly revealing ... so you’ll need to look count grid cells West-to-East (the *i* direction) and from South to North (the *j* direction) in order to determine a more suitable location for tracing circulation.

By means of dye tracing, looking at the (global only) overturning stream-function, and/or temperature and salinity (and density) profiles, see if you can identify where in the Cretaceous ocean deep water forms in the model. AND, more importantly, think about WHY does it form where it does?

Also: what about surface ocean circulation and gyres? Are these located where you would expect (e.g. based on your understanding of modern ocean circulation). Plot velocity vectors to help, and/or refer to the Barotropic streamfunction output in the 2D netCDF file.

4.2 The climate of the early Eocene

A previously spun-up state of early Eocene climate with cGENIE in a ca. 56 Ma configuration (LAB_5b.SPIN) is provided as a starting point. A user-config (LAB_5b.EXAMPLE) that continues on this climate state is run:

```
$ ./runmuffin.sh cgenie.eb_go_gs_ac_bg.p0055c.NONE LABS  
LAB_5b.EXAMPLE 10 LAB_5b.SPIN
```

(Note that once again, for speed, no carbon cycle is selected in this configuration.) Again, as per for the late Cretaceous configuration, a configuration that includes the ocean dye (color) tracers is also provided. The corresponding user-config (LAB_5b.colorinjection) that continues on this climate state can be run as follows:

```
$ ./runmuffin.sh cgenie.eb_go_gs_ac_bg.p0055c.rb LABS  
LAB_5b.colorinjection 10 LAB_5b.SPIN
```

What to ‘do’ with this? Again – start by tracing and analyzing the patterns of ocean circulation in the Eocene world, as per how you did for the modern and Cretaceous system. Where does deep water form in the Eocene? Is the surface ocean circulation as expected? Can you see any patterns emerging? At least in model world – what seems to be dictating the locations of deep water formation? Does overturning and/or surface circulation appear to significantly vary as a function of continental configuration or radiative forcing? If not, why not?

But how do you know how close, or not, the model climate is to ‘reality’? Available from the website, on the LH side of the page under ‘got data?’ at the bottom (item (7)), are 2 netCDF files containing proxy sea surface temperature (SST) data re-gridded to the cGENIE model grid. One is for pre-PETM SSTs, and one for peak PETM SSTs. (The data are from Dunkley Jones et al. [2013].)

These netCDF files can be opened and visualized in exactly the same way in exactly the same way as per the cGENIE output files. You’ll see a few (there are not many!) data points, in a sea of grey (which stands for ‘no value’). (Note that it is best to turn data interpolation OFF.)

More useful . . . is that the data distribution can be combined with the model ocean temperature output file, in a difference map (see earlier for notes on creating difference maps). You can thus visualize the pattern of model-data mismatch. (Note you might want to hit ‘Fit to data’ to autoscale the plot, or simply pick and manually enter a +/- limit for the data plotting, perhaps ensuring it is symmetrical about zero such that red anomalies could represent proxy SST values higher than min the model, and blue point data being lower than in the model.)

Furthermore, in the Arrays tab, you can switch to a Zonal Average view rather than a Map view. (It is probably easier to visualize the model-data misfit in this way.)

So, starting with the pre-PETM data and given model configuration, make some assessment of how the model fits the data (or not). What might be the reason for the misfit? You might test adding, and adjusting, some of the parameter values controlling surface SST from the Cretaceous climate exercise. Can you reduce the model-data misfit?

Finally, the PETM SST data given in the 2nd netCDF file are from the peak of the PETM. The ocean is rather warmer as compared to just prior to the PETM. Your task is to determine (either using the default model configuration, or your adjusted configuration) much radiative forcing (parameter `ea_radfor_sc1_co2`) and hence CO_2 is required to explain this warming?

Creating alternative Worlds

Using muffingen

Configuring cGENIE.muffin

Testing

Investigations

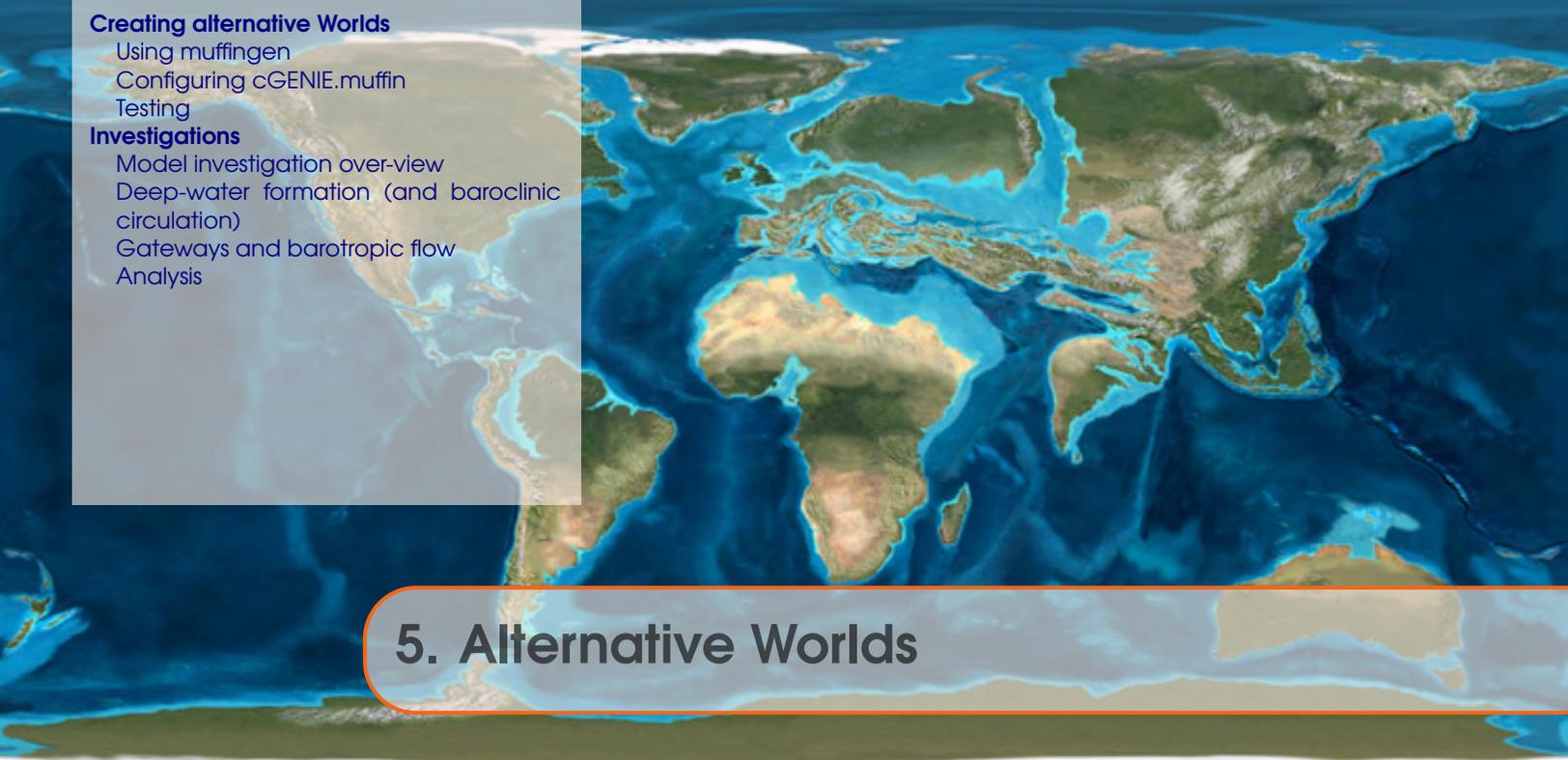
Model investigation over-view

Deep-water formation (and baroclinic circulation)

Gateways and barotropic flow

Analysis

5. Alternative Worlds



5.1 Creating alternative Worlds

5.1.1 Using muffingen

Refer to the **muffingen** *user-manual* as your primary source of information and tutorial on how to create new and adjust existing continental configurations of **cGENIE.muffin**.

Zonal wind-stress

For non GCM-based configurations, not prior wind fields exist. **muffingen** hence creates and configures an idealized zonal wind-stress field (from which wind velocity and wind speed is derived). The zonal wind-stress can take alternative strengths, depending on whether a high latitude gateway (in either hemisphere) exists. This can be prescribed directly, or **muffingen** can be enabled to 'choose' whether or not a high latitude gateway exists and hence whether or not to apply a strong or weak zonal flow. The parameter options (in the **muffingen** configuration file) are:

- `par_tauopt=0;`
muffingen chooses whether or not to apply a strong or weak zonal flow, and in which hemisphere.
- `par_tauopt=1;`
 A weak zonal flow is applied in both hemispheres.
- `par_tauopt=2;`
 A strong zonal flow is applied in both hemispheres.

For reference – the modern world has a mix of strong (southern) and weak (northern) hemisphere zonal flows.

The different EXAMPLE (.m) configuration files have a mixture of default choices:

- `muffingen_settings_BLANK,`
`muffingen_settings_drakeworld,` `muffingen_settings_eqpasworld,`
`muffingen_settings_ridgeworld,` `muffingen_settings_waterworld:`
`par_tauopt=0;`
- `muffingen_settings_wppcont1:`
`par_tauopt=2;`
- `muffingen_settings_modern:`
 n/a (wind-stress derived from GCM fields)

(No EXAMPLE configurations currently specify a strong zonal field, although `muffingen_settings_ridgeworld` will be prescribed one automatically in **muffingen**.)

5.1.2 Configuring cGENIE.muffin

Having created a new **cGENIE.muffin** configuration using the **MATLAB** function `muffingen`, carry out the following steps:

1. Firstly, copy/transfer the entire configuration subdirectory (the directory with the same name as whatever you called your 'world') and its contents, from the **muffingen** output directory, to:

```
cgenie.muffin/genie-paleo
```

2. Create a new *base-config* file. You do this by:

- (a) Taking one of the template *base-config* files:

```
CONFIG_template_081v1_R07.config
```

```
CONFIG_template_161v1_C09.config
```

The first being suitable for an 8-level (non seasonally forced ocean) and the second for a 16-level (seasonally forced) ocean.

Copy/rename the file to something ... 'appropriate'. (No spaces allowed in the filename.)

- (b) There is a highlighted (<<< >>>) line in the template *base-config* file:

```
# *****
# GRID & BOUNDARY CONDITION CONFIGURATION
# *****
# insert the automatically generated muffingen parameter list here
# *****
# <<<                                     >>>
# *****
```

Copy and past the contents of the **muffingen** output file:

`config_yymmdd.txt`

where indicated. (In the filename, `yymmdd` is the date of the configuration creation.)

- (c) Now copy this new *base-config*, to:

`cgenie.muffin/genie-main/configs`

3. (Create a new *user-config* file to complete the experimental setup.)

Note that in using one of the template *base-config* files, you have configured an ocean with just 2 tracers – temperature and salinity (i.e. there is no carbon cycle or ocean nutrients etc. enabled at this point).

5.1.3 Testing

5.2 Investigations

The following sub-sub-sections, details a set of (alternative) model-based investigations into fundamental controls on the nature of large-scale/global ocean circulation patterns.

5.2.1 Model investigation over-view

Whilst both investigations could in theory be based on the modern continental configuration (and climate) in **cGENIE.muffin**, you'd be much less likely to come by any particularly deep or fundamental insights into ocean circulation. In any case, the modern and future (and recent glacial past) ocean circulation states and dynamics has been picked at endlessly by 1000s of researchers, writing 100,000s of increasingly boring and un-revealing publications ... and still we do not fully understand modern ocean circulation, let alone that at the last glacial.

A better (more fun) approach is to generalise the problem and consider fundamentally different configurations of ocean basins and climate. This is what you will be doing. But first, some potted advice:

- Try and define a hypothesis, or hypotheses, to pursue and test. These need only be very rough at the outset – you'll likely find that you get new ideas and can either refine your original hypotheses, or come up with new ones, as you start to play with the model and see what is feasible and not feasible in terms of analysis.
- Create a plan of study – what sort of experiments, how many, how are they going to be analysed, how do they all fit together in addressing the overarching hypotheses? Again – it is very likely that your plan will evolve, but try and start out with something to guide you rather than wandering randomly in model world ...
- For creating and using different worlds – create a list or table of the configurations to be explored, and summarize what you are finding. It might be helpful to plan out on paper first, some of the main continental configurations you need to create and then test in the model. (Again, this will evolve.)
- Once you have some sort of idea what are going to do – plan the work. This is important because to run the model to steady state is not trivial. Perhaps plan on having to run the model for 5,000 years¹ in order to achieve a fully spun up ocean circulation state. You might get away with shorter runs, but know in advance, what sort of error this would include and is it 'important'? Once you have a list and know how long each might take, you can plan out when they will be run on the cluster (presumably on the queue) and when the results will be analysed. Note that it is a virtual certainty, that once you have analysed the results of the first set of experiments, you will want something different (and will then need to revise the plan and list of experiments and analysis etc.).

Overall – scientific investigations with simplified (and relatively fast) Earth system models tends to be a very iterative undertaking and can involve significant trial-and-error. Rarely, can you devise at the outset, a single run or set of experiments, and it turns out this is completely sufficient. If you do not see something you do not understand in the results of the initial model experiments, you are either God (e.g. the Spaghetti Monster or Invisible Pink Unicorn), or not doing it properly. Expect to see things that interest you and lead you off on a tangent (hopefully – this is how research should be).

Finally – note that in swapping between different continental configurations, in **cGENIE.muffin**,

¹Even so, you should try running the model much longer to be confident that 5,000 years is OK (and sufficiently close to final equilibrium).

the models needs (or really really wants to) re-compile.² This ... cannot happen on the compute nodes of the cluster if you submit an experiment using a different *base-config*. When changing to a new *base-config*: first, run the model briefly interactively (i.e. at the command line). Once it has compiled (and started running), the experiment can be killed (Ctrl-C) and now it is good to submit to the queue as a job. In any case, in utilizing a new configuration that you may never have used before, it is good practice to test it first (e.g. watch it run for a short period).

5.2.2 Deep-water formation (and baroclinic circulation)

The first investigation is to investigate the controls on the strength and large-scale structure of ocean circulation, particularly in terms of the global meridional overturning circulation (MOC), and associated with this – the primary sites of deep-water formation.

There are presumably 2 key controls to this:

1. Temperature.

This is presumably mostly controlled by latitude (i.e. the further North or South, the more suitable the site will tend to be for deep-water formation). There will also be an influence of the prescribed planetary albedo (which includes the effect of clouds) as well as of sea-ice (if it exists).

2. Salinity.

This will generally be controlled by P-E – the balance between precipitation to the ocean surface as well as fresh-water run-off from the continents, vs. evaporation from the ocean surface. Sea-ice may also be a key factor, in leading to the (seasonal) rejection of dense salty brine.

In turn, this suggests two sets of 'knobs' in the model that influence the patterns and magnitude of temperature and salinity across the ocean surface:

1. Continental Configuration.

The configuration of the continents and ocean basins will dictate the shape and location of the highest latitude ocean regions – presumably the locations where on average, deep-water formation will occur (and hence forming the downwards/sinking limb of the MOC).

The model knob is hence how the position and orientation of continental blocks has been set. This (creating or changing the continental configuration used by **cGENIE.muffin**) is all done using the **MATLAB muffingen** program.

2. Global Climate.

The mean and climate as well as the zonal temperature gradients, together with whether or not sea-ice forms (and how much), will modulate both temperature and salinity patterns at the ocean surface.

The model knobs here are primarily:

- (a) Atmospheric pCO_2 , or in the absence of an explicit carbon cycle, a prescribed radiative forcing:

```
ea_radfor_scl_co2=1.0
```

which here, specifies $\times 1 CO_2$ equivalent radiative forcing.

²Note that if the *base-config* is the same as the previous experiment, but you have changed a parameter value (in the *user-config*), you do not need to re-compile.

- (b) One could also adjust the value of the solar constant (here, given with its modern/present-day default value):

```
ma_genie_solar_constant=1368.0
```

which has a subtly different effect from changing CO_2 radiative forcing, as radiative forcing has a relatively spatially uniform impact, whereas changing the solar constant has a disproportionate impact towards the Equator (where the incident solar shortwave radiation is the greatest). So changing the solar constant is likely to impact the pole-to-Equator temperature gradient. e.g. see: Lunt, D. J., A. Ridgwell, P. J. Valdes, and A. Seale, Sunshade World.: a fully coupled GCM evaluation of the climatic impacts of geoengineering, *GRL* 35, L12710, doi:10.1029/2008GL033674 (2008).

- (c) One could also adjust the planetary albedo, particularly in respect of looking to modify the pole-to-Equator temperature. In the **muffingen** created configurations, there is an explicit 2D file specifying the zonal (with latitude) planetary albedo profile:

```
xxxxxxxx.albd.dat
```

where xxxxxxxx is the 8-character name of the **muffingen** created World. It would be a simple matter of editing the values in the file (between the range of 0.0 and 1.0, for perfectly absorbing, and perfectly reflective, respectively).³ Note that the file runs North-to-South (top to bottom).

5.2.3 Gateways and barotropic flow

The overarching question for this investigation is a little harder to define succinctly; it has to do with the conditions, of the degree of alignment or prevailing wind (stress) and open gateways, that give rise to strong zonal ocean flow. A good and obvious modern example is the existence of the Drake Passage, between the northern tip of the Antarctic Peninsular, and the southern tip of Southern America, how this aligns with the prevailing Westerlies in the Southern Hemisphere, and hence the nature and strength of the ACC (Antarctic Circumpolar Current).

So the question naturally arises: how misaligned does a gateway have to be relative to the prevailing wind stress maximum, before the circumpolar (or circum-equatorial) flow ceases. What about having multiple gateways and their relative alignment, including what happens if one gateway is aligned with Westerly wind stress, and a second with Easterly wind stress – who 'wins'? Also – what about sill depths? For the Drake Passage, the ocean floor, while tectonically messy, is generally relatively deep. What happens to ocean circulation with a progressively shallow sill depth?

The 2 key controls in this exercise are then:

1. Gateway alignment.

How does gateway, or gateways, align with the maximum of the zonal wind stress? For multiple gateways, what if one aligns with a wind-stress maximum of the opposite sign?

2. Sill depth.

The sill depth! This could be uniform in depth across the gateway (easiest/best) or could be varying (probably not so easy to learn anything).

The importance of the position of the gateways is primarily only in the context of the position of the maxima in the wind-stress field. In theory, the shape of the assumed zonal wind-stress could

³Best to make a copy of the original file before you modify it.

be altered ... but the wind stress needs to be defined on 2 different grids, and this gets messy.

As before, there are a number of changes that can be made in the model to explore the consequences of varying gateway position and sill depth (and knobs, turned):

1. Land-sea mask.

The model knob is hence how the width and location of gateways has been defined in the land-sea mask. There is also the question of how many gateways (and their respective position). As before, this (creating or changing the continental configuration used by **cGENIE.muffin**) is all done using the **MATLAB muffingen** program. Any of the given example **muffingen** configurations: **drakeworld**, **eqpasworld**, **ridgeworld**, or **waterworld**, could be taken as starting points – copying and renaming the configuration **.m** file, as well as the **.dat** file in the **INPUT** directory.

2. Ocean bathymetry.

The bathymetry (ocean floor depth) can be changed to alter the sill depth. The easiest way to do this is probably within the **muffingen** editor, using the setting:

```
opt_user=true; % [false/true] enable user input to grid
```

in the configuration **.m** file. (One might also then, 'draw' in the gateways by hand at the same time.)

Note that a gap between 2 land masses must be 2 or more cells wide, to count as a gateway (and allow barotropic flow).

3. Wind stress strength.

Although it is messy to attempt to edit the profile of the applied wind-stress field, it is possible to scale its impact on ocean circulation lower and higher. The **cGENIE.muffin** parameter for this is:

```
go_13=1.531013488769531300
ea_11=1.531013488769531300
```

Somewhat bizarrely ... it appears twice ... with different parameter names. Both parameters must be changed to the same value. Place these lines (of the new parameter value assignments) in the *user-config* file for the experiment. Higher values result in a stronger applied wind-stress on the ocean surface.

Note that although this is the simplest change to make, it may not have such a clear scientific question associated with it (other than the obvious and trivial).

5.2.4 Analysis

How do you 'judge' the strength and characteristics of deep-water formation, global overturning, and circulation patterns and strength, in general, or their climate (or biogeochemical) impacts? This is not a trivial question. Often, additional ocean tracers are employed in models to generate a quantitative measure of the age of a parcel of water (mean time since it last saw the surface), of some measure of the efficiency of ventilation, or large-scale transport at the surface or at depth.⁴ Or rather involved analysis of ocean physics and transport might be employed.

One starting point is to read some of the literature where the climate properties of various hypothetical worlds have been investigated. Such as:

⁴We'll see such tracers employed later in the course.

- *Marshall et al.* [2007] – ‘Mean Climate and Variability of the Atmosphere and Ocean on an Aquaplanet’, *Journal of the Atmospheric Sciences* **64**.
- *Enderton and Marshall* [2009] – ‘Explorations of Atmosphere–Ocean–Ice Climates on an Aquaplanet and Their Meridional Energy Transports’, *Journal of the Atmospheric Sciences*.
- *Ferreira et al.* [2010] – ‘Localization of Deep Water Formation: Role of Atmospheric Moisture Transport and Geometrical Constraints on Ocean Circulation’, *Journal of Climate* **23**.
- *Smith et al.* [2006] – ‘Global Climate and Ocean Circulation on an Aquaplanet Ocean–Atmosphere General Circulation Model’, *Journal of Climate* **19**.

In addition, the sub-sub-sections that follow outline some simple diagnostics and ways of going about some quantitative analysis.

Simple/global diagnostics

- In the biogem output results folder, there is a time-series file named:

```
biogem_series_misc_opsi.res
```

This contains a summary of the evolution with time, of the minimum and maximum (anywhere) global overturning stream-function values.

- Another simple property of the climate system that you might consider, is the pole-to-equator temperature gradient, both in terms of atmospheric temperature, and ocean surface temperature⁵ (although they should presumably be closely coupled).

Why? Because the large scale (overturning) circulation of the ocean should be transporting heat from the high latitude surface to the deep ocean. This presumably would act to reduce latitudinal temperature gradients. In contrast, a strong zonal slow might prevent latitudinal transport of heat.

Using Panoply

In a physics-only (T + S tracer) configuration of **cGENIE.muffin** you are more limited. A good starting point is the previous tutorial on ocean circulation and AMOC stability (in the context of the modern world, climate and continental configuration). For instance, you know already how to plot and visualize the MOC using **Panoply**. The Atlantic basin and hence the existence of an AMOC, is pretty specific and unique to the modern world, so likely you’ll need to focus on the global MOC. (A good starting point is to familiarise yourself with the pattern and intensity of the modern.)

For example – the drakeworld configuration gives rise to a global MOC pattern as shown in Figure 5.1.

This is stored as the variable `phys_opsi` in the netCDF file `fields_biogem_2d.nc`. Remember that the first time-slice plotted in **Panoply** is the first time-slice saved.

You can also plot the barotropic circulation using **Panoply** – the variable is called `phys_psi` – this is shown in Figure 5.3.

Finally in **Panoply**, you might plot the current field (surface or otherwise), as per Figure 5.2.

Using MATLAB ...

...

⁵**Panoply** has an option for plotting the zonal mean, from which you could read off pole-to-equator temperature gradients.

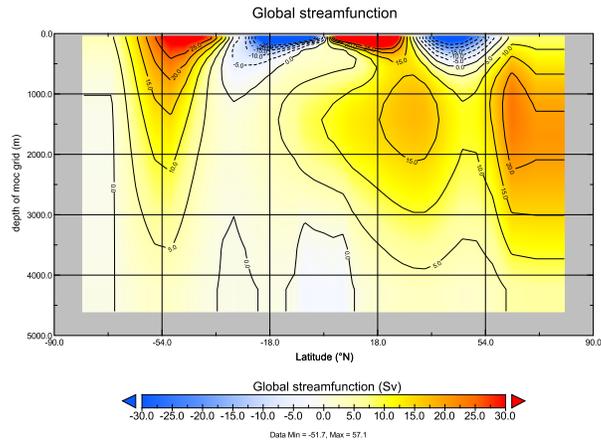


Figure 5.1: Drake work world overturning circulation.

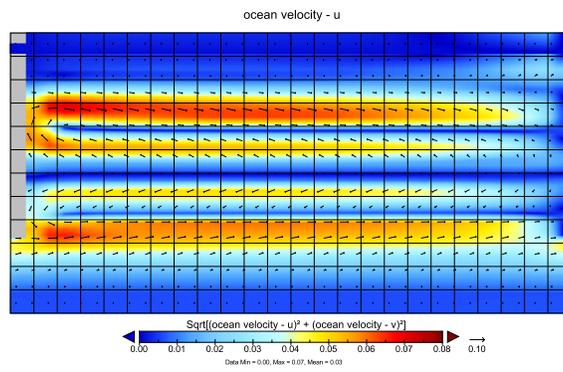


Figure 5.2: Surface current velocity field (arrows) plus speed (color scale).

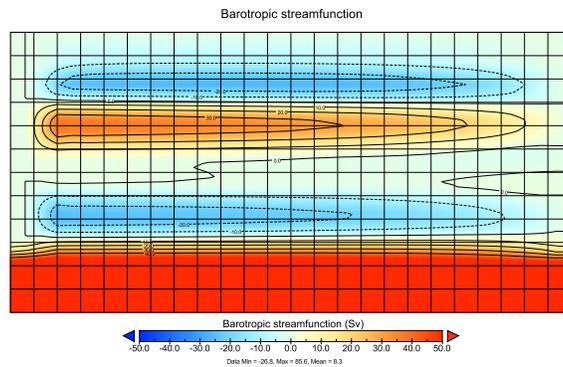


Figure 5.3: Drake work world barotropic circulation.

Via color tracing

Numerical color tracers can be used (as you have seen previously) to trace flow-paths. The default *base-config* files provided as part of the **muffingen** software, define a configuration with only 2 tracers in the ocean – T and S. To add red and blue dye tracers, in the *base-config* file, find the section marked:

```
# *****
# TRACER CONFIGURATION
# *****
# the total number of tracers includes T and S
# T and S do not need to be explicitly selected and initialized
# *****
# Set number of tracers
GOLDSTEINNTRACSOPTS='$(DEFINE)GOLDSTEINNTRACS=2'
# list selected biogeochemical tracers
# <<< >>>
# list biogeochemical tracer initial values
# <<< >>>
# *****
```

You need to make several (but simple) edits here:

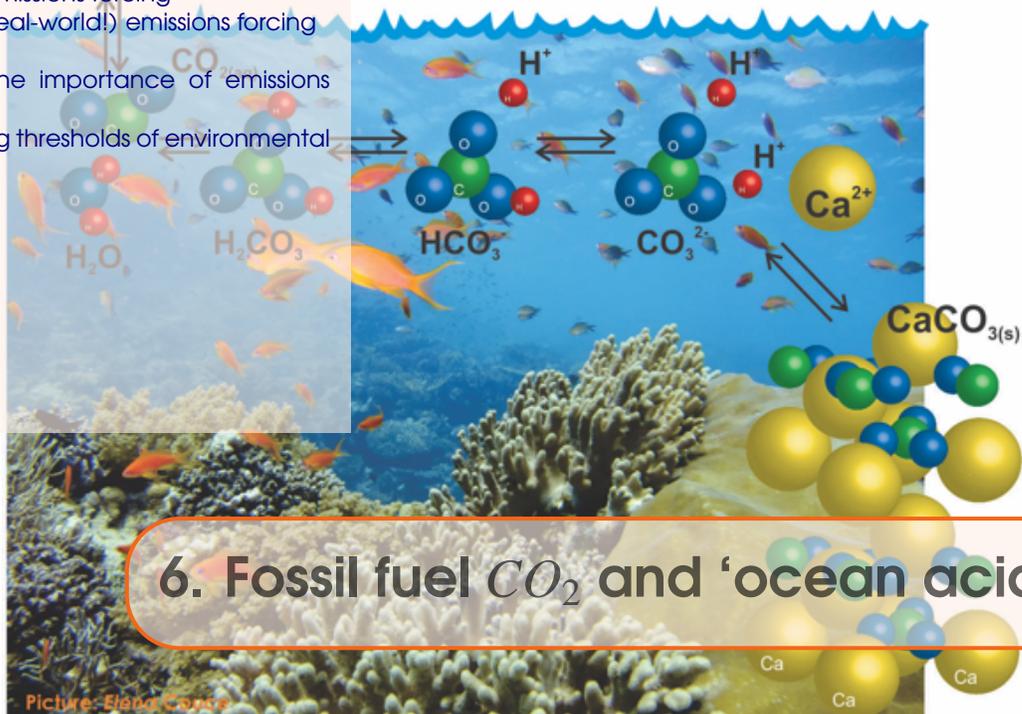
1. Firstly, change the number of tracers from 2 to 4, in line:
`GOLDSTEINNTRACSOPTS='$(DEFINE)GOLDSTEINNTRACS=2'`
2. Secondly, you need to list the additional list selected biogeochemical tracers. For this add the following code in place of the (<<< >>>) line:
`gm_ocn_select_48=.true.`
`gm_ocn_select_49=.true.`
3. Lastly, under the section list biogeochemical tracer initial values, and in place of the (<<< >>>) line:
`bg_ocn_init_48=0.0`
`bg_ocn_init_49=0.0`

Exploring the consequences of fossil fuel CO_2 emissions

Idealized emissions forcing
Historical (real-world!) emissions forcing

Further ideas

Assessing the importance of emissions rate
Determining thresholds of environmental impact



6. Fossil fuel CO_2 and 'ocean acidification'

Readme

You will need to download a new restart file prior to embarking on the experiments. This pre-industrial spin-up includes a basic ocean (-atmosphere) carbon cycle plus various diagnostic anthropogenic tracers, following *Cao et al.* [2009].

To fetch this: change to the `cgenie_output` directory, and type:

```
$ wget http://www.seao2.info/cgenie/labs/AWI.2017/LAB_3.SPIN.tar.gz
```

Extract the contents of this archive by typing:

```
$ tar xfvz LAB_3.SPIN.tar.gz
```

(change directory back to `genie-main` to run the model)

6.1 Exploring the consequences of fossil fuel CO_2 emissions

For the next experiment(s) you can chuck CO_2 into the atmosphere, just for the hell of it. As much as you want! Apparently, humans are actually doing this now. Imagine that!

The *user-config* for **cGENIE.muffin**, LAB_3.CO₂emissions is provided and configured with climate being responsive to CO_2 (i.e., it takes account of CO_2 -climate feedbacks):

```
# set \(\text{CO}_{2}\)-climate feedback
ea_36=y
```

as well as having a rate of calcification by plankton at the surface ocean that is responsive to ocean acidification and saturation state (i.e., it takes into account CO_2 -calcification feedbacks, which will additionally interact with climate – see *Ridgwell et al.* [2007b, 2009] from <http://www.seao2.info/pubs.html>). Anything could happen!!!

In this *user-config* file, a release of CO_2 to the atmosphere is prescribed, which by default is set for just 1 PgC over an interval of a single year. (Releasing CO_2 just over a single year is obviously rather unrealistic and many impacts will decay rapidly away, but represents a useful idealized experiment for assessing the time-scale(s) of fossil fuel CO_2 uptake by the ocean.) Additional **netCDF** output has also been prescribed (bg_par_data_save_level=10) so that more information relevant to assessing ocean acidification is saved.

Run the experiment for e.g., 20 (or more if you like) years, starting from the pre-industrial re-start experiment LAB_3.SPIN, i.e.:

```
$ ./runmuffin.sh cgenie.eb_go_gs_ac_bg.worjh2.BASE LABS
LAB_3.\(\text{CO}_{2}\)emissions 20 LAB_3.SPIN
```

As for what model results variables to consider ... think about the climate change and ocean acidification literature and which environmental properties are considered either critical for ecosystems or are simply helpful and illustrative. Refer to the Ocean acidification output supplement (S1) for a summary of some of the key ocean acidification variables saved by the model. In the 3-D **netCDF** time-slice file remember, for instance, that ocean surface waters in which aragonite becomes under-saturated (OHMEGA < 1.0) is regarded as a critical threshold for organisms making aragonite shells and skeletons and spells TROUBLE for some poor calcifying marine organism somewhere. (Temperature is also highly relevant to marine ecosystems under future global change.) Note that the calcification response is encoded in the model and described in *Ridgwell et al.* [2007a,b] (see: <http://pubs.seao2.org>) and may or may not reflect the real World.

For climate change ... the variables of particular interest should be obvious. Remember that there are both time-series outputs, as well as special 2D and 3D fields which might be more helpful for relating impacts to specific ecosystems.

6.1.1 Idealized emissions forcing

You can easily modify the experimental design to release more/less CO_2 very much as you did for the red dye tracer. In the *user-config* file, the line:

```
bg_par_atm_force_scale_val_3=8.3333e+013
```

scales the CO_2 flux given in the forcing file

biogem_force_flux_atm_p\ (CO_{2}) _sig.dat

which can be found in the directory:

cgenie.muffin/genie_forcings/pyyyyz.Fp\ (CO_{2}) _Fp13\ (CO_{2})

The format of this file is:

```
-START-OF-DATA-
  0.0  1.0
  1.0  1.0
  1.0  0.0
999999.9  0.0
-END-OF-DATA-
```

and defines an emission of 1 mol C per year over the first 1 year of the model experiment (between year 0.0 and 1.0), but which in the example user-config is then scaled by a value of 8.333×10^{13} (by the parameter `bg_par_atm_force_scale_val_3`) to give a total of 1 PgC yr⁻¹. (Year 999999.9 has no special meaning and is simply just way in the future ...)

Pause ... and note briefly how the final CO_2 flux is arrived at. **cGENIE.muffin** calculates it by multiplying the value in the forcing file (1.0) by a modifying parameter in the user config file ($8.3333e+13$). The total flux is hence: $1.0 \times 8.333 \times 10^{13} = 8.333 \times 10^{13}$ mol CO_2 yr⁻¹. If you set both values as 1.0, you’d get very little carbon released (a single mole!). If you screw up and multiply $8.3333e+013$ and $8.3333e+013$ as the total flux, you’ll soon know it as you cook the Earth ... But it does not matter which parameter has value 1.0 and which scales the units ($8.3333e+013$). For now, it is simply more convenient to be able to edit the forcing file with simple numbers (and leave the units conversion in the user config file).

Together, the scaling and forcing value gives a CO_2 release of 1 PgC yr⁻¹ for just a single year compared to current emissions are about 10 PgC yr⁻¹. So, do not expect anything exciting to happen at this point.

(The parameter: `bg_par_atm_force_scale_val_4=-27.0` specifies the carbon isotopic composition of fossil fuel carbon and can be ignored.)

Because ‘accidents can happen’ and the global environmental changes induced by the massive fossil fuel CO_2 release can obscure mistakes made in the experiment configuration (parameter values) and/or the re-start used, you are strongly advised to first (or in parallel, as a job submitted to the cluster – refer to Lesson Zero to remind yourself of the commend line syntax needed for this) set a control experiment going:

```
$ ./runmuffin.sh cgenie.eb_go_gs_ac_bg.worjh2.BASE LABS
  LAB_3.CONTROL 20 LAB_3.SPIN
```

Here – the user-config defining the control experiment (LAB_3.CONTROL) is identical to that for the actual experiment itself (LAB_3. CO_2 emissions) with the exception of the scaling of the CO_2 emissions that have been set to zero. (It is left completely to you to create the experiment configuration file LAB_3.CONTROL.)

If everything is OK, atmospheric CO_2 (and climate) should be stable and there should be little (or no) drift in any of the output variables (because the spin-up should have been run to an equilibrium state and you have not changed anything in the control experiment, right?).

It is good practice (i.e. do it!) to always run a control experiment for each different type of experiments – e.g. ideally run one control experiment for each set of CO_2 emissions experiments.

OK. You might want to run something a little more exciting now. For instance, rather than

```
-START-OF-DATA-
  0.0  1.0
  1.0  1.0
  1.0  0.0
999999.9  0.0
-END-OF-DATA-
```

you might have:

```
-START-OF-DATA-
  0.0 1000.0
  1.0 1000.0
  1.0   0.0
999999.9  0.0
-END-OF-DATA-
```

So now a total of 1000 PgC over a single year. Now you should see some policy-relevant impacts occur :o)

You can control the shape of the emissions profile as well as its magnitude. Between the start and end ‘tags’ in the text file, the data is arranged into 2 columns: the first contains a series of tie-points for defining the timing of changes in emissions, and the 2nd column contains flux information (units of PgC yr⁻¹ when scaled by the parameter `parameter bg_par_atm_force_scale_val_3` in the *user-config*). At each time-step of the model the CO_2 flux is interpolated between these time points.

The purpose of:

```
0.0  1.0
 1.0  1.0
 1.0  0.0
```

then specifies a uniform flux of 1.0 (scaled to PgC yr⁻¹) over the first full year of the model run, followed by a sharp turn-off to zero flux at the end of first year (and remaining zero thereafter). To extend the period of emissions – for example:

```
0.0  1.0
10.0  1.0
10.0  0.0
```

would result in a uniform flux lasting 10 years (i.e. once scaled by the parameter in the *user-config* – 1 PgC yr⁻¹ over 10 years – 10 PgC total emissions). In contrast;

```
0.0  0.0
10.0  1.0
```

would result in a linear ramp, starting from zero at the start of year 0.0 to 1.0 PgC yr⁻¹ at year 10.0 (a total CO_2 emission of $1 \times 10 \times 0.5 = 5$ PgC over 10 years).

6.1.2 Historical (real-world!) emissions forcing

Historical and future (SRES) emissions scenarios can also be prescribed explicitly. A historical emissions forcing (technically: a prescribed concentration profile of pCO_2 and other anthropogenic gases) can be specified by adding/substituting the following lines to the user-config:

```
bg_par_forcing_name='worjh2.historical2010'
```

Now, no additional scaling is needed because the forcing specification directly follows the observed change in atmospheric concentration with time (in units of atm CO_2) and the line containing the scaling parameter:

```
bg_par_atm_force_scale_time_3=1.0
```

should be deleted (or commented out with a # at the start of the line). An additional line is needed in the user-config because the historical pCO_2 transient starts in the 1700s (for which a nominal date of 1765 is often used) rather than year zero. For example, to start from year 1765, the start year parameter must be set:

```
bg_par_misc_t_start=1765.0
```

Because the start year has changed, it is convenient to specify save points that are consistent with the historical period, e.g.:

```
bg_par_infile_slice_name='save_timeslice_historicalfuture.dat'
```

```
bg_par_infile_sig_name='save_timeseries_historicalfuture.dat'
```

A *user-config* with these changes is provided for your convenience (or to double-check you were following it all) -- LAB_3.historical. A suitable experiment would then be one run for 245 years so that it reaches year 2010 (having started from year 1765):

```
$ ./runmuffin.sh cgenie.eb_go_gs_ac_bg.worjh2.BASE LABS
  LAB_3.historical 245 LAB_3.SPIN
```

WARNING! Ignore the ‘WARNING’s at the start – these are simply telling you that more tracer forcings have been specified than you have selected tracers for in the *base-config* (cgenie.eb_go_gs_ac_bg.worjh2.BASE). (A different *base-config* with additional selected tracers could have been specified to make use of other historical changes in atmospheric composition, such as of radiocarbon (14C) and CFCs.) Also: from year 1765 onwards, changes in atmospheric CO_2 only rise very slowly initially. Don’t expect to see anything happen in 10 seconds flat because relatively few people and countries in the 1800s could be bothered to burn much more than a little local coal. You could potentially start your experiment at year 1850, changing the value of `bg_par_misc_t_start` and specifying shorter experiment duration if you are desperate for the End of the World to come.

Don’t forget: you could submit this experiment to the cluster and do more (idealized emissions) ‘playing’.

Given that there is observationally-based information on the distribution of anthropogenic CO_2 taken up by the ocean (e.g. Sabine et al. [2004]) and having run a historical transient experiment with the model driven by observed increases in atmospheric pCO_2 , you are in a position to critically evaluate the model’s ability (or lack of) to represent the future-critical process of oceanic fossil fuel CO_2 uptake and transport by large scale ocean circulation.

In the 2D netCDF output, there is a variable for the water column integrated inventory of DIC – equivalent to the Sabine map except you will need to subtract the preindustrial background of DIC first, i.e. to create a DIC anomaly map representing only the added fossil fuel CO_2 component of ocean DIC. The data in the Sabine paper clusters around 1994. A time-slice centered on this year (1994.5) has been configured in the model exactly for this purpose. Your baseline state can either be from prior to CO_2 emissions commencing at any significant rate (e.g. 1750.5) or (better), from a control experiment. Note that similar comparisons could be (and are regularly) made with other tracers such as CFCs, which provide additional insights into the patterns and time-scales of trace gas uptake and ocean circulation. (See: Cao et al. [2009])

Observational data, re-gridded to the cGENIE grid and in netCDF format can be downloaded from the ‘usual place’ (<http://www.seao2.org/mycgenie.html>) from the ‘got data?’ box on the left. You could for instance, compare horizontal or vertical slices (3D netCDF) and create difference (anomaly) maps. Somewhat more representative of the entire ocean is to compare (or calculate difference maps) of zonal average profiles. Unfortunately, the observations are not in the form of water column integrals and hence you cannot create difference maps of model as per the Sabine paper ... unless you are MATLAB-friendly and you use the 3D BIOGEM MATLAB plotting scripts (genie-matlab) whose use is somewhat described in the cGENIE user-manual. Examples of MATLAB plotting of the model vs. observed anthropogenic anomaly are shown in Figure 6.1 (note the use of plotting un-interpolated model grid data as colors but with an interpolated contour overlain to help guide the eye and pick out features).

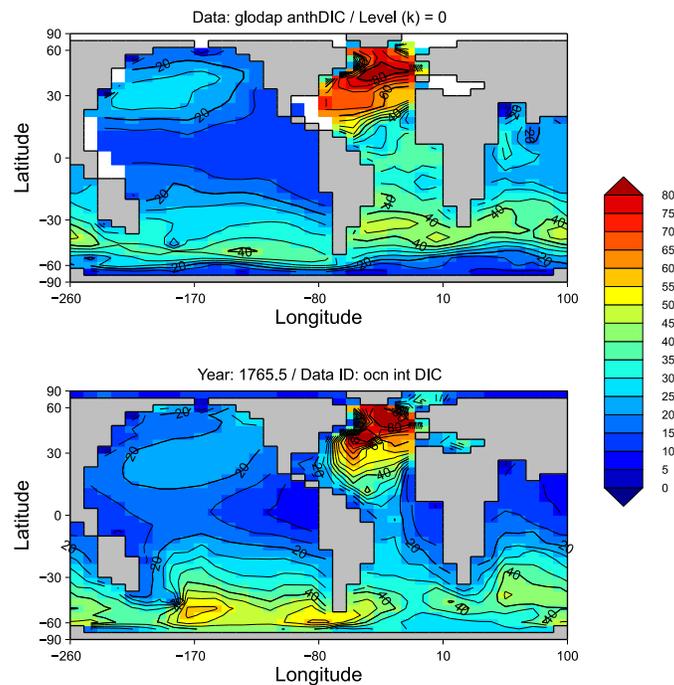


Figure 6.1: Observed (top) vs. Model (bottom) anthropogenic CO_2 inventories. Data and model water column integrals in units of $mol\ CO_2\ m^{-2}$ and are nominally with respect to year 1994.

Finally, and the closest to being slightly interesting: rather than applying highly idealized pulses of CO_2 emissions, IPCC SRES emissions scenarios can be used to make future projections with. An example forcing of this sort is provided and can be selected by changing the name of the forcing selection parameter (`bg_par_forcing_name`) to:

```
worjh2.FeMahowald2006.Fp\ (CO_{2}\) _Fp13\ (CO_{2}\) _A2_02180PgC
```

which gives you the IPCC ‘A2’ scenario (extended beyond year 2010 in this case to give a total cumulative fossil fuel burn of 2018 PgC, e.g. Ridgwell and Schmidt [2010]). Again, as this forcing has units of PgC yr⁻¹ in its time-series file, you will need to add a scaling parameter to the user-config file to turn units of PgC yr⁻¹ into mol yr⁻¹, i.e.

```
bg_par_atm_force_scale_val_3=8.3333e+013
```

For complete ‘realism’ you will need to run this experiment starting from the end of the historical transient experiment (Section 1.6), e.g.

```
$ ./runmuffin.sh cgenie.eb_go_gs_ac_bg.worjh2.BASE LABS
  LAB_3.future 90 LAB_3.historical
```

and with the start from year now set to year 2010 (the end year of the historical transient):

```
bg_par_misc_t_start=2010.0
```

Note that the *user-config* LAB_3.future is not provided for you – create this (or a file named whatever you like) by copying e.g. LAB_3.EXAMPLE and making the parameter changes described above (forcing specification parameter, emissions scaling parameter, and start year parameter).

You can also easily replace the details of the emissions with other SRES scenarios – simply find the year vs. emissions rate information from the interweb (e.g. http://sres.ciesin.columbia.edu/final_data.html) and edit or copy-and-paste) the flux values for each decade into the file `biogem_force_flux_atm_pCO2_sig.dat` in the forcing directory. **cGENIE.muffin** will then automatically interpolate between the decadal tie-points to give a continuous change in emissions. Now you are able to make a rather more realistic/plausible assessment of when and where potential ecological impacts (via assumed ocean chemistry criteria) might occur.

6.2 Further ideas

6.2.1 Assessing the importance of emissions rate

By editing the flux and/or timing information you can control the CO_2 emissions trajectory and total fossil fuel burn. Explore different CO_2 release assumptions and note their impact on climate and ocean biogeochemistry. Much more realistic and appropriate to our current global experimenting is a lower rate (order of 10 or 20 PgC yr⁻¹) released over a longer interval (order 100 years) compared to the conceptual 1000 PgC near instantaneous pulse. Because the experiments are getting longer to run in real time ... remember to make appropriate use of the cluster queuing facility – i.e. think about whether you want to sit around staring at the screen for 15 minutes waiting for a new line of numbers appear – if not: submit to the cluster queue. For instance, one might try and address the question: “For a given total release of CO_2 , is it safer to burn it slower?” The answer is maybe not completely obvious, as burning carbon resources slower will result in a small global impact, but perhaps one that persists for longer. You could conceive of an ensemble (set) of model experiments, maybe one of 100 PgC yr⁻¹ for 1 yr, one of 10 PgC yr⁻¹ for 10 years, and one of 1 PgC yr⁻¹ for 100 years, and run them all for e.g. 100 years. (As jobs submitted to the queue, all can be run simultaneously.) (Don’t forget the control experiment!) But note that you should create 3 new forcings based on the original if you are editing the forcing and expecting to run different ones at the same time. Really, this is little from copying and renaming user-config files, except it involves entire directories in genie-forcings. Remember that the forcing is specified by the directory name assigned to `bg_par_forcing_name` (enclosed in ’’).

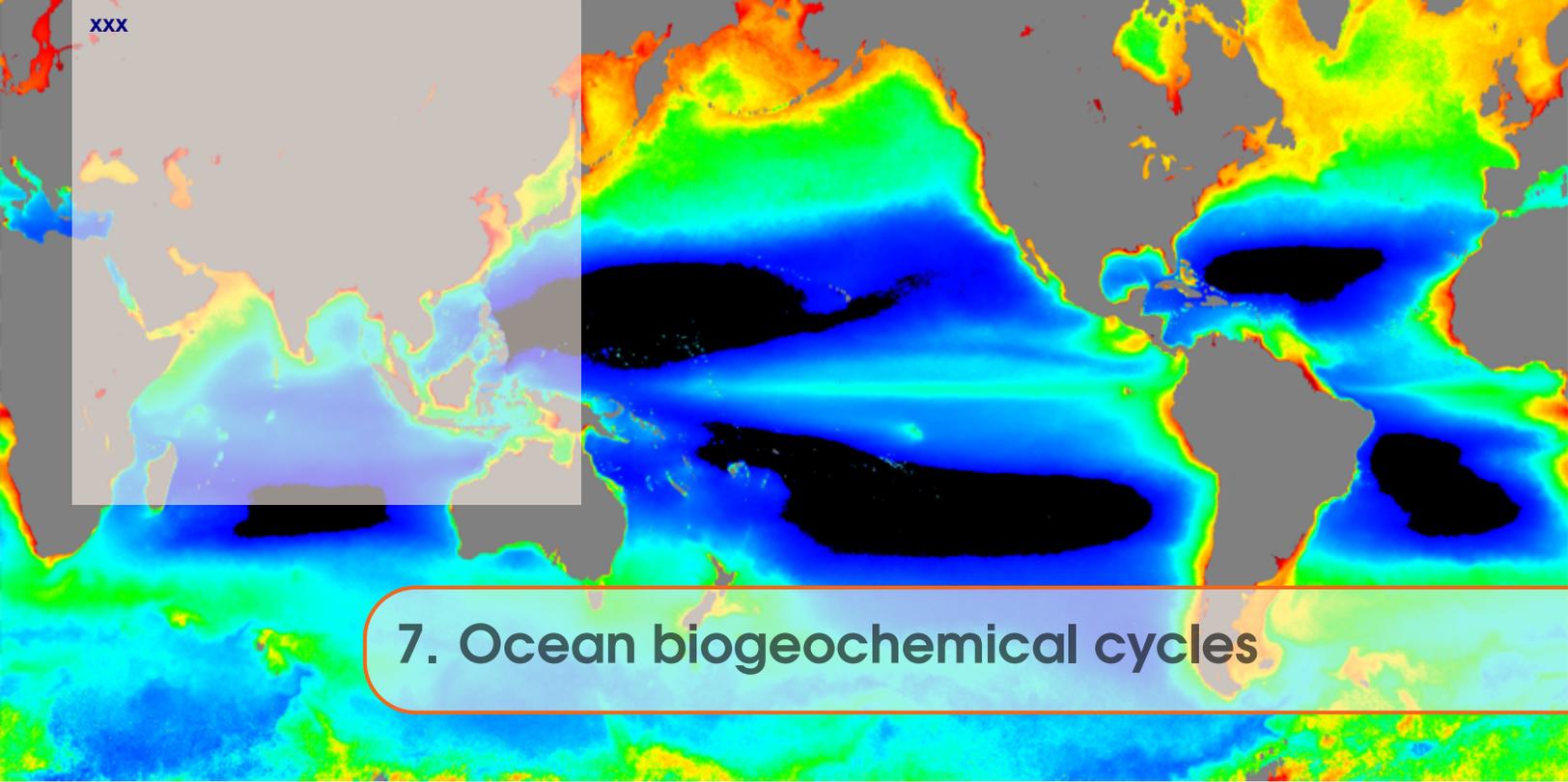
6.2.2 Determining thresholds of environmental impact

There are various concerns about the impacts of continuing fossil fuel CO_2 emissions and a number of proposed climatic (e.g. the 2 degree C global warming limit often mentioned in policy documents) and ecological ‘tipping points’. You can assess the maximum allowable CO_2 emissions to remain within particular global environmental limits in the model. For example:

- What is the maximum total CO_2 release that can be made without inducing aragonite undersaturation at the ocean surface anywhere (or any season – see Section 5.2.3 in the User Manual for seasonal time-slice data saving)? How important is the time-scale of emissions in determining this? For total emissions above this: where in the ocean does the surface first become under-saturated? How large would the emissions have to be in order to induce undersaturation at the surface in the tropics (home to socio-economically important reef systems). These are questions that can be addressed with simple CO_2 release experiments in ocean carbon cycle models and everyone seems to get a GRL paper out of it each and every time!
- How important are CO_2 -climate feedbacks in amplifying or diminishing future climate and ocean carbonate chemistry changes – e.g. is the same atmospheric pCO_2 value reached with and without climate feedback (and surface warming) – if not, why? You can investigate this by contrasting an experiment made including CO_2 -climate feedback with one made without. The CO_2 -climate feedback can be turned off by setting: `ea_36=n`.
- Also: How much CO_2 emission does it take to significantly ‘collapse’ the AMOC and over what time-scale? (Or alternatively: what is the atmospheric pCO_2 threshold for collapse?) If the AMOC weakens or collapses ... why in the absence of a prescribed freshwater perturbation does this happen? (Plotting appropriate ocean property anomalies between the CO_2 release experiment and a control experiment might help.)

Experiments could be hypothetical and consisting of CO_2 pulses or ramps (or exponentials)

and run on directly from a pre-industrial spin-up, or more 'realistic' and run on from the end of a historical transient experiment (e.g. starting in year 2010).



7. Ocean biogeochemical cycles

This Chapter ...

Background reading

-

READ.ME

You will need to download a new restart file prior to embarking on the experiments. This differs from previous provided restarts in that it now includes an iron cycle in the ocean and hence co-limitation of biological productivity by Fe.

```
$ wget http://www.seao2.info/cgenie/labs/AWI.2017/LAB_4.SPIN.tar.gz
```

Extract the results in the usual way and in the usual place ... and return to genie-main.

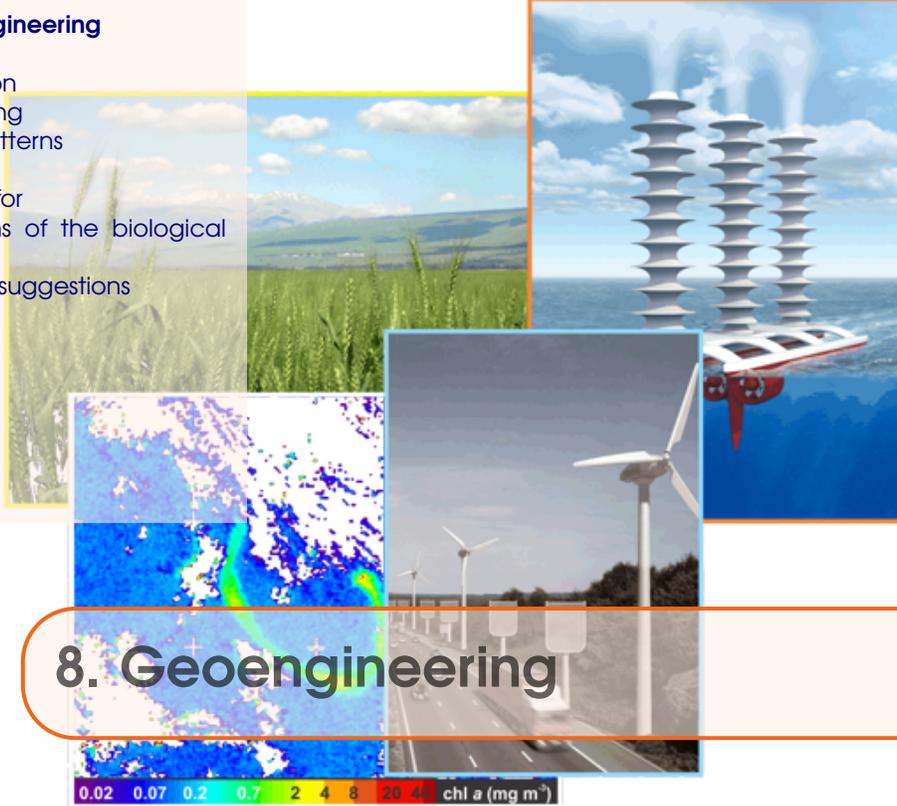
7.1 xxx

Ocean carbon geoengineering

Iron fertilization
Phosphate fertilization
Enhanced weathering
Modifying spatial patterns

Further ideas

Impacts to look out for
Further modifications of the biological pump in the ocean
Other thoughts and suggestions



8. Geoengineering

This Chapter ...

Background reading

-

README

You will need to download a new restart file prior to embarking on the experiments. This differs from previous provided restarts in that it now includes an iron cycle in the ocean and hence co-limitation of biological productivity by Fe.

```
$ wget http://www.seao2.info/cgenie/labs/AWI.2017/LAB_4.SPIN.tar.gz
```

Extract the results in the usual way and in the usual place ... and return to genie-main.

8.1 Ocean carbon geoengineering

In the following experiments you are going to explore some of the ocean biological controls on atmospheric pCO_2 (plus ocean acidification, and the distributions and intensities of oxygen minimum zones). Really, the ‘geoengineering’ focus is just an excuse to be looking at how the biological pump in the ocean works, how it regulates atmospheric pCO_2 , how sensitive it is to perturbation and what the consequences are of any changes in it. So if you are uncomfortable with ideas of large scale manipulating the Earth system, think about the relevance of the experiments to e.g. understanding why atmospheric pCO_2 was low at the time of the last glacial.

The overall idea of this lab is to run future CO_2 emissions scenarios and test whether ocean carbon geoengineering is an effective means for reducing future ocean acidification and marine ecological impacts (but keep in mind you are also exploring the basic natural operation of the system in doing so). You will require a pre-industrial spin-up and will need to create a new historical pCO_2 transient experiment because compared to the previous lab, you are now using a different *base-config* (`cgenie.eb_go_gs_ac_bg.worjh2.BASEFe`) that includes additional tracers for the marine iron cycle, i.e. you cannot simply use any of the experiments from previous labs as a restart.

Refer to the Supplement (final chapter section) for a guide as to what to ‘look for’ in the model results.

So to start with, go ahead and run a new historical transient experiment. A user-config is provided for your convenience (`LAB_4.historical`) ... but maybe check the settings for e.g. start year, as well as note that there are a number of new parameters to control the iron cycle (amongst other differences) as compared to before:

```
$ ./runmuffin.sh cgenie.eb_go_gs_ac_bg.worjh2.BASEFe LABS
LAB_4.historical 245 LAB_4.SPIN
```

(ignore the ‘WARNING’s at the start)

An example user-config is provided: `LAB_4.EXAMPLE`. This includes parameter settings for controlling any one of 3 different possible ocean carbon geoengineering schemes, described below (Section 1.3). By default, these are commented out (== ignored by the model) and only the forcing for the A2 emissions scenario (`worjh2.FeMahowald2006.FpCO2_Fp13CO2_A2_02180PgC`) with no geoengineering is set by default. You might regard this as a control (reference) experiment for all the with-geoengineering experiments you might run, i.e. the impacts of CO_2 emissions in the absence of any mitigation by geoengineering. To activate any particular geoengineering forcing: simply un-comment (delete the #) the appropriate pair of lines (the first line being the forcing specification, and the second one the total flux forcing used in the geoengineering scheme). If you have multiple (un-commented) settings of a parameter (e.g. `bg_par_forcing_name`) the value specified in the last occurrence is the one that is applied. This can get confusing, so if you un-comment out one set of parameter options, comment out (add a #) the ones you are not using.

The geoengineering (and control) experiments need to be run starting from the end of your historical transient experiment:

```
$ ./runmuffin.sh cgenie.eb_go_gs_ac_bg.worjh2.BASEFe LABS
LAB_4.EXAMPLE 90 LAB_4.historical
```

Because with a modern configuration and additional tracers in the ocean, the model is running rather slower than in some earlier exercises, you may not want to run beyond the end of the century (hence the 90 year experiment duration, starting from year 2010, as suggested above).

Each of the example geoengineering scenarios are delineated by its own specific forcing – a set of files that live in a uniquely named sub-directory within `genie-forcings`. The three forcings are:

- `worjh2.FeMahowald2006.Fp\CO2_Fp13\CO2_A2_02180PgC_FFe`
- `worjh2.FeMahowald2006.Fp\CO2_Fp13\CO2_A2_02180PgC_FPO4`
- `worjh2.FeMahowald2006.Fp\CO2_Fp13\CO2_A2_02180PgC_FALK`

Each forcing includes the A2 CO_2 emissions scenario, with the annual emissions (CO_2 flux) `biogem_force_flux_atm_pCO2_sig.dat` in units of PgC yr-1 (== GtC yr-1), hence requiring a units conversion setting in the user-config (`bg_par_atm_force_scale_val_3=8.3333e+013`) that is provided for you under the heading # CO_2 emissions scaling. (You can completely ignore the carbon isotope settings.)

Each forcing also includes a prescribed dust flux to the ocean surface (the FeMahowald2006 part of the directory name string). This is necessary because the model configuration you are using includes a co-limitation of biological productivity by iron (Fe) in addition to phosphate (PO₄). (The files associated with the dust forcing are: `biogem_force_flux_sed_det_sig.dat` and `biogem_force_flux_sed_det_SUR.dat` but you do not need to edit these files.) For the role of iron in controlling ocean productivity: possible starting points for background reading are: *Ridgwell and Kohfeld [2007]* (PDF available from my website) or *Jickells et al. [2005]* (Science).

The specific details of the 3 different example geoengineering scenarios are as follows:

8.1.1 Iron fertilization

Forcing: `worjh2.FeMahowald2006.FpCO2_Fp13CO2_A2_02180PgC_FFe` – a constant (with time) flux of dissolved Fe (in addition to whatever Fe dissolves into the surface ocean from the dust flux) is specified in: `biogem_force_flux_ocn_Fe_sig.dat`. The magnitude of the applied flux is then scaled in the *user-config* file by the setting:

```
bg_par_ocn_force_scale_val_9=1.0e+09
```

Note that this is simply an example total global flux. You might consider higher or lower fluxes, as well as potentially how ‘practical’ the annual production and supply of such quantities might be.

A spatial pattern of the flux is also defined, in the file:

```
biogem_force_flux_ocn_Fe_SUR.dat
```

An example pattern is set (see Section 1.4 for details on editing this pattern) – here a row of grid cells are marked at the same latitude in the Southern Ocean. You do not need to retain this pattern. In choosing an alternative: think about where in the modern ocean biological productivity is thought to be at least partly limited by the availability of dissolved Fe. Remember that the model may or may not correspond with reality, i.e. it may or may not predict Fe limitation in the correct regions, which may affect your choice of location for iron fertilization.

8.1.2 Phosphate fertilization

Forcing: `worjh2.FeMahowald2006.FpCO2_Fp13CO2_A2_02180PgC_FPO4` (‘macro-nutrient’ addition) – a constant (with time) flux of dissolved PO₄ is specified in: `biogem_force_flux_ocn_PO4_sig.dat`. The magnitude of the applied flux is then scaled in the user-config by the setting:

```
bg_par_ocn_force_scale_val_8=2.0e+12
```

Again, you should consider this as an example total flux. In choosing a total flux to apply, points of comparison include whatever the total weathering flux (via rivers) of P to the global ocean is. Also: global phosphate (fertilizer) production, which produces an interesting potential conflict between geoengineering and food production, although there are proposals for using fertilized ocean regions for enhanced fish production.

A spatial pattern of the flux is also defined, in the file:

```
biogem_force_flux_ocn_PO4_SUR.dat
```

An example pattern has been set – here, the Equatorial Atlantic. In choosing your regions(s), think about where in the ocean (again – there may be differences between real ocean and model) productivity is currently limited by PO₄. Also be aware of possible on-set of Fe limitation if you relieve the PO₄ limitation (i.e., you could potentially lose effectiveness if you supply too much PO₄ and instead productivity and CO₂ drawdown is capped by a second factor). You could potentially consider PO₄ and Fe addition at the same time ... ?

8.1.3 Enhanced weathering

Forcing: worjh2.FeMahowald2006.FpCO₂_Fp13CO₂_A2_02180PgC_FPO4 (alkalinity addition) – a constant (with time) flux of alkalinity is specified in: `biogem_force_flux_ocn_ALK_sig.dat`. The magnitude of the applied flux is then scaled in the user-config by the setting:

```
bg_par_ocn_force_scale_val_12=5.0e+13
```

Again, another example total flux. In choosing a total flux to apply, points of comparison include whatever the total weathering flux (via rivers) of alkalinity (often described in terms of the bicarbonate ion flux) to the global ocean is. Also: global cement (lime) production. (Note that in one mole of lime: CaO, you have 2 moles of alkalinity (Ca²⁺)).

A spatial pattern of the flux is also defined, in the file:

```
biogem_force_flux_ocn_ALK_SUR.dat
```

An example pattern has been set – here, bordering the major tropical coral reefs locations in the Western Pacific. In choosing your regions(s), you might think about mitigating specific ecosystem impacts of ocean acidification, or about the feasibility of transport and proximity to abundant limestone (CaCO₃ – the source of lime) and/or energy.

8.1.4 Modifying spatial patterns

The spatial patterns of an applied flux forcing to the ocean can easily be modified. The pattern is specified in a simple ASCII (plain text) file, in the file in the forcing sub-directory ending ‘_SUR.dat’. The file (in this example the default Fe pattern) looks like:

8.2 Further ideas

8.2.1 Impacts to look out for

- Impacts and ecosystems of interest could potentially be ones residing on the ocean floor, such as cold-water (deep water) corals, and are not necessarily planktic (/surface) only.
- Don't forget that different calcifying organisms employ different mineralogies (calcite vs. aragonite), with different saturation states and hence potentially susceptibility to ocean acidification. Hence thresholds of both aragonite and calcite saturation will be relevant, depending on the organism. Depending on the organism, saturation changes occurring in specific regions may be much more relevant than a global mean change. Also, it might be the seasonal minimum value reached, rather than annual averaged minimum, that is critical.
- Some of the arguments against some forms of ocean carbon geoengineering concern the potential for adverse impacts on marine organisms (and positive climate feedbacks) induced by decreases in the degree of oxygenation in the ocean, such as expanding and/or intensifying oxygen minimum zones. cGENIE saves 3D fields of O₂ concentrations that can be plotted in slices through the ocean of various orientations.

8.2.2 Further modifications of the biological pump in the ocean

Other manipulations of the biological pump and ocean carbon cycle are possible and potentially instructive and in the following examples may be of rather more relevance to past climates and carbon cycles and e.g. possible reasons for the low atmospheric CO_2 concentrations at the last glacial, as opposed to relevant to geoengineering (a good thing!). The first two of these may have profound effects not only atmospheric pCO_2 but also on dissolved oxygen concentrations in the ocean (and hence implications for the suitability of animal habitat such as for fish) and this is something that you will want to look at as part of your overall assessment of impacts.

Remineralization depth

In the model configuration that you have been using, the degradation of particulate organic matter sinking in the water column proceeds according to a fixed profile of flux with depth (there is no e.g. temperature control on the rate of bacterial degradation of sinking organic matter) with CO_2 and PO_4 released back to the seawater as the particulate flux decreases. The parameter that controls the (e-folding) depth scale of particulate organic matter is:

```
bg_par_bio_remin_POC_eL1=589.9451
```

Either edit this value (found under the heading: # -- REMINERALIZATION --) or add a new line at the end of the *user-config* file specifying the value you want. Units are *m*.

Read Ridgwell et al. [2007] for additional discussion of this parameter. See Figure 2-4 in Ridgwell [2001] (http://www.seao2.org/pubs/ridgwell_thesis.pdf) for an illustration of how the flux of particulate organic matter decreases with depth in the ocean, plus references therein.

There is also an associated parameter: `bg_par_bio_remin_POC_frac2`, which sets a fraction of organic matter that is assumed to settle through the water column completely un-altered (currently assigned a value of 0.045 == 4.5

Note that there may well be no simple parallel that can be found in geoengineering to this process. However, there are hypotheses that during the last glacial and as a result of colder ocean temperatures, the depth scale was longer. Conversely, there are ideas about that the warmer temperatures of the e.g. Eocene ocean and hence faster rates of bacterial metabolism led to a much shallower remineralization depth scale. So a remineralization depth scale that is responsive to

temperature may have importance in understanding ocean biogeochemical cycles during both past warm and cold climates as well as obviously, future global change. While you are not implementing a temperature-dependent parameterization explicitly, you can at least test for whether changes in temperature might have important impacts by simply changing the remineralization depth to be shallower (smaller depth-scale under a warming climate) or deeper (greater depth-scale in a colder ocean).

Macro nutrient inventory and uptake

Suggestions have been made that nutrients were used more efficiently during the LGM, meaning that for the same nutrient uptake at the surface more carbon was exported to depth in the ocean. See: Omta et al. [2006]. There are also a bunch of (relatively old) hypotheses concerning differences between glacial and modern ocean in how much nitrate (NO₃⁻) there was. There is no NO₃⁻ in this version of GENIE (just PO₄³⁻ and Fe), but an analogous change can be made to the phosphorous cycle.

For the nutrient-to-carbon ratio in organic matter, the relevant parameter is:

```
bg_par_bio_red_POP_POC=106.0
```

To change the default value (106.0), add a new line at the end of the user-config file specifying the value you want. A larger number means that PO₄ is being utilized more efficiently and more organic matter is being produced for the same nutrient consumption.

To test the effect of there being more PO₄ in the ocean, in addition to using the (surface) flux forcing as described earlier, it is also possible to simply increase the inventory of the ocean as a whole in one go:

```
bg_ocn_dinit_8=1.0E-6
```

which will add 1 $\mu\text{mol kg}^{-1}$ of PO₄ uniformly to the ocean. (A larger/smaller number will obviously increase the glacial nutrient inventory by more/less.)

In terms of geoengineering, changing the ‘Redfield’ ocean plankton might be difficult ... but not impossible, although we are presumably talking about releases of genetically modified organisms to the entire ocean to achieve this meaning there are obviously some severe ethical concerns. However, adding macro nutrients such as PO₄ (more often, NO₃ is talked about) may be more feasible.

CaCO₃:POC rain ratio

Kicked off by a classic 1994 Nature paper by Archer and Maier-Reimer (see: Kohfeld and Ridgwell [2009]), one potential means of changing atmospheric CO₂ naturally at the last glacial involves changes in the export ratio between CaCO₃ (shells) and POC (particulate organic matter). Such a change in ratio could come about through a variety of ways (e.g., via the ‘silica leakage hypothesis’ (see: Kohfeld and Ridgwell [2009]) and also through the direct effect of Fe on diatom physiology (see Watson et al. [2000] in Nature and also Supplemental Information). There are also ideas about an opposite ocean acidification effect, whereby the less acidic glacial (compared to modern) ocean led to increased calcification and CaCO₃ export. Note that this response (higher saturation == great calcification) is encoded into your model configuration – see Ridgwell et al. [2007b].

In GENIE, the CaCO₃:POC rain ratio is controlled (technically: scaled) by the parameter:

```
bg_par_bio_red_POC_CaCO3=0.04
```

The pattern of CaCO₃:POC rain ratio is not uniform across the ocean (why? (see: Ridgwell et al. [2007, 2009]), and its pattern can be viewed in the (2D BIOGEM) netCDF variable: misc_sur_rCaCO3toPOC.

(Note that it is unlikely that there is any parallel in a geoengineering context to this process.)

8.2.3 Other thoughts and suggestions

- If you want to combine forcings, you need to first update the file: `configure_forcings_ocn.dat` – this specifies which ocean flux forcing will be used – simply copy the relevant line from the equivalent file of the forcing to be added. You will also need to copy in the relevant ‘_sig.dat’ and ‘_SUR.dat’ files. Remember that in the user-config file, you will need to set the relevant flux scaling parameter for each different flux in the forcing.

- By default, the CO₂-climate feedback is ‘on’:

```
# set climate feedback  
ea_36=y
```

Should you want to assess the impacts of geoengineering independently of changes in climate – the option is there. (Note that under some of the high end CO₂ emissions scenarios, there may be a degree of collapse of the AMOC that will presumably affect the patterns of ocean acidification and oxygenation etc.)

- If you are having doubts that your experiment is actually ‘doing’ anything (different from the control) – remember to create anomaly maps (plots) to look for specific changes in e.g. saturation state, pH, or the water column inventory of anthropogenic CO₂. Even before this – plot anomalies of the flux you think you have applied, looking specifically at the region you think you have applied it to. For this, cGENIE saves the 3D distributions of dissolved Fe and PO₄. See Figures below.
- Always be aware of the caveats regarding this specific model (and models in general) – how much does it differ from the ‘real world’ for the modern ocean, particularly in terms of patterns of carbonate saturation? Does it even simulate anthropogenic CO₂ uptake adequately in the first place (e.g. see session #2)?

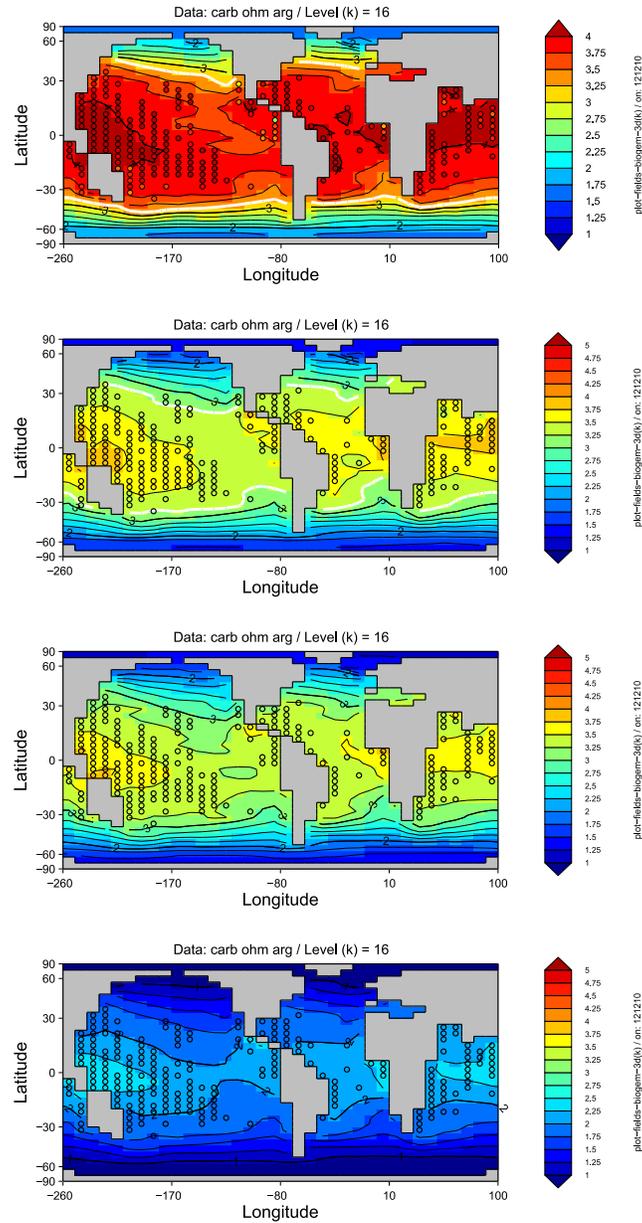


Figure 8.1: **Mean annual ocean surface saturation (aragonite) changes.** Top: pre-industrial model ocean surface saturation (aragonite) with ReefBase tropical coral reef locations re-gridded to the GENIE grid and color-coded with modern observationally-based saturation values. 2nd and 3rd down: Year 1994 and 2010 ocean surface saturation (aragonite) with ReefBase reef locations. Bottom: Year 2010 ocean surface saturation (aragonite) under the A2 CO_2 emissions scenario. The thick white line delineates the 3.25 saturation contour (inferred to reflect a limitation on corals). *Examples here produced using MATLAB (plotting scripts are located in genie-matlab) but equally do-able in Panoply with the exception of achieving a data overlay. These are provided simply to illustrate some of the impacts you might consider and possible ways of visualizing them.*

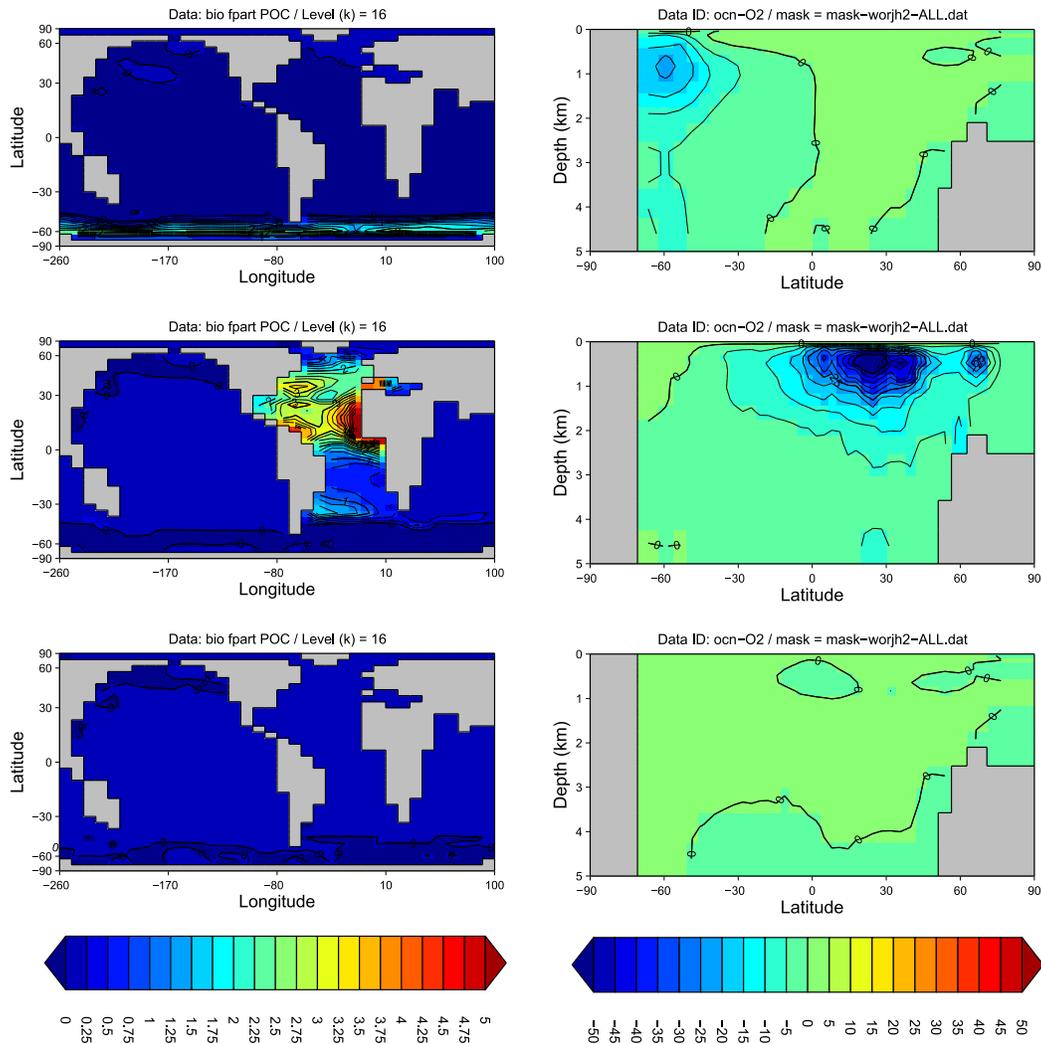


Figure 8.2: **Ocean surface export (particulate organic carbon) and zonal [O₂] anomalies.** Left: anomalies of global mean annual export production, for Fe fertilization (top), PO₄ addition (middle), and ocean liming (bottom). Right: Zonal mean anomalies of dissolved O₂ concentrations. *Examples here produced using MATLAB (plotting scripts are located in genie-matlab) but equally do-able in Panoply with the exception of achieving a data overlay. These are provided simply to illustrate some of the impacts you might consider and possible ways of visualizing them.*

Getting going with ECOGEM

- Running the model
- Viewing 2D time-slice output
- Comparing to observations

Ecosystem configuration

- Visualising composite data
- Iron limitation

Increasing ecological complexity

- Plankton size classes
- Viewing 2D time-slice output
- Create your own ecosystem

Build it up, tear it down

- A fully size-structured ecosystem
- Ecosystem characteristics
- Mixotrophy

Ecology in a paleo-ocean

- The PETM

9. Marine ecosystems and dynamics

The following is based on workshop material originally devised by Ben Ward <b.a.ward@bristol.ac.uk>

Stuff to keep in mind...

- We will be working with highly idealised ecosystems in a highly idealised ocean.
- The aim is to explore why the model behaves as it does.
- The assumption is that this will give us some insight into why the real world behaves as it does. Perhaps. (It is up to you to question the validity of this assumption.)

Background reading

- *Ward et al. [2017]* – manuscript submitted to GMD.

Read.me

Before you start, you need to retrieve some model experiment *restart* files. At the terminal, enter the five following commands (hitting ‘return’ after each command):

```
cd ~/cgenie_output
wget http://www.seao2.info/restarts/BENW.worjh2.P04FeTDTL_HYBRID_new.SPIN.tar.gz
wget http://www.seao2.info/restarts/EXAMPLE.p0055c.RidgwellSchmidt2010.SPIN1.tar.gz
tar xfvz BENW.worjh2.P04FeTDTL_HYBRID_new.SPIN.tar.gz
tar xfvz EXAMPLE.p0055c.RidgwellSchmidt2010.SPIN1.tar.gz
```

You will also need to carry out a code update:

```
cd ~/cgenie.muffin
svn update
```

and then return to the **GENIE** main directory:

```
cd genie-main
```

(Also before you start – read the *Ward et al.* reference.)

9.1 Getting going with ECOGEM

Previously, you have run the standard 'biogeochemical' version of **cGENIE.muffin**, and biological export fluxes calculated via simple mass balance (and limitation) considerations by the module **BIOGEM**. In this chapter, we are going to get started with the '**ECOGEM**' package. This will allow us to extend the capabilities of **cGENIE.muffin** to examine a range of questions relating to the role of physiology and community structure in regulating the biological pump.

First, we need to understand the relationship and differences between **BIOGEM** and **ECOGEM**. In **BIOGEM**, the biological pump is driven by an implicit (i.e. unresolved) biological community. As in the real ecosystem, the biological uptake of carbon and nutrients (such as phosphorus and iron) is limited by light, temperature and nutrient availability. Unlike the real ecosystem, any uptake is *directly* and *instantly* returned to dissolved organic matter (DOM) and inorganic nutrients in the ocean interior.

- surface inorganic nutrients $\xrightarrow[\text{and export}]{\text{production}}$ DOM and deep inorganic nutrients

In **ECOGEM**, biological uptake is again limited by light, temperature and nutrient availability, but here it must pass through an explicit and dynamic intermediary plankton biomass pool, before being returned to DOM or dissolved inorganic nutrients.

- surface inorganic nutrients $\xrightarrow{\text{production}}$ plankton biomass $\xrightarrow{\text{export}}$ DOM and deep inorganic nutrients

9.1.1 Running the model

We will start with the simplest possible configuration of **ECOGEM**, with just a single phytoplankton class. You can run this model at the command line, by entering the following command (note that this should be one continuous line) . . .

```
./runmuffin.sh cgenie.eb_go_gs_ac_bg_eg.worjh2.BASESFdTDL / BSS.NPD.SPIN 10
BENW.worjh2.P04FeTDL_HYBRID_new.SPIN
```

As before, we have five input parameters after the run script (`./runmuffin.sh`) is invoked:

1. `cgenie.eb_go_gs_ac_bg_eg.worjh2.BASESFdTDL.benw` → The '*base-config*' file. Note the additional '`_eg`' that marks the inclusion of the **ECOGEM** package.
2. `/` → The *user-config* directory where the *user-config* file resides.
3. `BSS.NPD.SPIN` → The *user-config* file (experiment name). Here, 'NPD' stands for 'Nutrients, Plankton, Detritus'.
4. `10` → The run duration (for this particular example experiment), in years.
5. `BENW.worjh2.P04FeTDL_HYBRID_new.SPIN` → The *restart* filename. Here we are telling the model to start from the endpoint of a previous experiment (a 10,000 year run of **BIOGEM**). Note that the restart here only applies to the biogeochemical part of the model. The ecological community will be initialised from some very low biomass.

The model will run as before, except that now we have the **ECOGEM** debug option enabled, and you will get a lot of extra information about how the ecological model is configured.

The extra lines, e.g.

```
>>> SAVING ECOGEM TIME-SLICE AVERAGE CENTERED @ year : 0.500
```

confirm that **ECOGEM** is writing ecological time-slices at the same time as **BIOGEM** is writing its own time slices. (**ECOGEM** is not currently set up to save time-series data in the same way as **BIOGEM**.)

9.1.2 Viewing 2D time-slice output

Following the same convention as for **BIOGEM**, **ECOGEM** *time-slice* output will be saved in the directory `~/cgenie_output/BSS.NPD.SPIN/ECOGEM/`

1. Open the `fields_ecogem_2D.nc` file by locating it in the correct directory, and double clicking on it in the file transfer window. Alternatively, you can open **Panoply** by selecting File → Open, and clicking through.
2. You should now see a list of 2D arrays that were output by **ECOGEM**. Looking at the Long Name description, simply click on a variable of interest. If a menu window pops up, just click on Create or hit the Return key.
3. Check the Panoply settings to make sure you really know what you are looking at.
 - (a) Which time-slice (i.e. simulation year) are you looking at?
 - (b) What is the data range (i.e. colour scale)

NOTE: You can change the default settings in **Panoply** to avoid changing things every time you open a new file. First click on Panoply → Preferences.... Here you can switch off interpolation and the grid overlay under the General menu. You can disable the spuriously precise coastline under Lon-Lat Plots.

9.1.3 Comparing to observations

Models are usually intended as an approximation of the real world (whatever that is). It might, therefore, be useful to check if our approximation is in anyway realistic. We can do this by comparing the model output to observations.

1. You can download a compilation of key biogeochemical variables from the mycenie [web-page](#).
2. You can open the `GEnIE_observations.nc` file in **Panoply** in the same way as you opened the `fields_ecogem_2D.nc` file.
3. You can now compare the model output to key biogeochemical variables, such as surface chlorophyll or phosphate. (Also, e.g. create difference maps.)
4. Does the model perform well or poorly with respect to reproducing these variables?

NOTE: **ECOGEM** only saves a limited number of surface (2D) data arrays. You can look at other variables (in 2D and 3D) by opening the corresponding **BIOGEM** files, `fields_biogem_2D.nc` and `fields_biogem_3D.nc` in **Panoply**. They can be found in the directory:
`~/cgenie_output/BSS.NPD.SPIN/BIOGEM/`

Bear in mind that we restarted the model from a previous simulation with **BIOGEM**. Some of the state variables may take a long time (i.e. $\gg 10$ years) to adjust to the new model configuration (especially at depth).

9.2 Ecosystem configuration

In the last section you ran a very simple version of the ecosystem model, and compared it to observations. In this section we are going to add a bit more ecological realism, with the aim of improving model performance. We will start by adding a zooplankton population that should bring a degree of ‘top-down’ control to the phytoplankton population.

Details of the original ecosystem are specified in the *user-config* file, `BSS.NPD.SPIN`.

1. Locate the *user-config* file (in `~/cgenie.muffin/genie-userconfigs/.`), and open it in your preferred text editor.
2. The *user-config* file can be used to configure the model to your liking. One of the most important amendments to note straight away can be seen on line 11, `bg_par_bio_prodopt="NONE"`. This effectively disables the biological pump in **BIOGEM**, replacing it with the explicit biology of **ECOGEM**. This is a necessary step whenever running **ECOGEM**, because we do not want the implicit and explicit biological schemes to be implemented in the same simulation.
3. We can also see a load of other model parameters. Any that begin with ‘bg_’ correspond to **BIOGEM**, while ‘eg_’ corresponds to **ECOGEM**. The **ECOGEM** parameters begin after line 71.
4. One of the most important parameters specifies the *ecosystem configuration* file:

```
eg_par_ecogem_plankton_file = 'NPD.eco'
```

This points to a file (located in `~/cgenie.muffin/genie-ecogem/data/input/`) that specifies every plankton population that is included at the start of the model run. If you open that file in the text editor, you will see something akin to the following:

```
01          02    03
\          \    \
```

```
-START-OF-DATA-
Phytoplankton    10.00    1
-END-OF-DATA-
```

```
/\          /\    /\
01          02    03
```

```
DATA FORMAT AND ORDER
-----
```

```
COLUMN #01: plankton functional type name
COLUMN #02: plankton diameter (micrometers)
COLUMN #03: number of randomised replicates
```

```
INFO: TRACER ASSIGNMENT RULES
-----
```

```
Plankton functional type one of: Prochlorococcus
                                Synechococcus
                                Picoeukaryote
                                Diatom
                                Coccolithophore
                                Diazotroph
                                Phytoplankton
                                Zooplankton
                                Mixotroph
```

5. The first thing to note is that only the lines in between the `-START-OF-DATA-` and `-END-OF-DATA-` tags are read by the computer. The rest is there solely for your guidance.

Each line that is entered in the computer-readable area tells the model to put a distinct plankton population in the model. The “plankton functional type” of this population is specified in the first column, while the plankton diameter specified in the second column. A ‘1’ must always be placed in the third column (it doesn’t do anything, but the model still needs it).

In this ‘NPD’ configuration, we only have a 10 micron generic phytoplankton. The ecological and physiological traits of this population are assigned automatically according to the size and the functional type.

NOTE: The only PFTs available at the moment are Phytoplankton, Zooplankton and Mixotroph. The other groups currently have no real functionality associated with them. (If you have some good ideas, we may be able to add them in later in the course).

6. We can increase the ecological complexity of the model by adding another plankton population. Save the *ecosystem configuration* file under a new and highly intuitive name (such as NPZD.eco), and add another line specifying a 100 micron zooplankton. It is important that the zooplankton is 10 times larger than the phytoplankton in terms of diameter. This is the optimal predator-prey length ratio in the default configuration. (You could maybe think about changing this value later on in the course.)
7. To run the model with this new configuration, change the name of the *ecosystem configuration file* in the *user-config* file...

```
eg_par_ecogem_plankton_file = 'NPZD.eco'
```

8. Save the new *user-config* file under a different name (e.g. BSS.NPZD.SPIN). You can now execute the model at the command line. Don’t forget to change the name of the *user-config* file here as well ...

```
qsub -j y -o cgenie_log -V -S /bin/bash runmuffin.sh
  cgenie.eb_go_gs_ac_bg_eg.worjh2.BASESFcTDTL
  / BSS.NPZD.SPIN 10 BENW.worjh2.P04FcTDTL_HYBRID_new.SPIN
```

9. Once you have completed the new simulation, compare the new results to the old simulation, in terms of its ability to reproduce observations. Has the addition of zooplankton to the model improved its behaviour?

Look also at the global distributions of carbon biomass in the phytoplankton and zooplankton populations (again, a log scale might help).

How have the zooplankton interacted with the phytoplankton to change the model dynamics?

9.2.1 Visualising composite data

We can perhaps get a better handle on this question by looking at the ratio of phytoplankton-to-zooplankton biomass. Such ratios can, however, be difficult to assess simply by eyeballing two maps. Instead we can use Panoply to combine data arrays.

1. First close all your **Panoply** plot windows. Then open a new one for C Biomass - Popn. 001 (10.00 micron phytoplankton). Next, select C Biomass - Popn. 002 (100.00 micron zooplankton), and click the Combine Plot icon at the top of the **Panoply** window.
2. A box will open up asking you In which existing plot should I combine the variable. As you now only have one plot available, this should be a straightforward choice. Click Combine.
3. A new map should appear showing the total zooplankton carbon biomass minus the total phytoplankton carbon biomass (see the label on the colour scale). This is not what we want.

Below the map, under the Array(s) tab, there is a drop down menu showing the range of different ways the two arrays can be combined. We want to look at the Z:P biomass ratio, so select Array 2 / Array 1.

4. You now need to make sure that you are looking at the right year (you can time-lock the two arrays by clicking on the chain icon). You may also find it helpful to look at the data on a log scale, with a scale range of 0.1 to 10. You might also like to change the Color Table: option to GMT_polar.cpt.

Questions:

- What does this plot say about the relationship of zooplankton and phytoplankton in different regions of the ocean?
- In what regions do zooplankton or phytoplankton dominate?
- What affect does a high Z:P ratio have on the size of the phytoplankton population? (For example, in terms of the chlorophyll concentration.)

9.2.2 Iron limitation

Up to this point, we have only considered phosphate as a limiting nutrient. (Iron was included in the model, but it was not limiting to phytoplankton growth.) You can switch on iron limitation by modifying two lines in the *user-config* file:

```
eg_useFe=.true.
```

and

```
eg_fquota=.true.
```

Give the *user-config* file a new name (e.g. BSS.NPZD_Fe.SPIN), and reevaluate the model. Again, don't forget to change the name of the *user-config* file in the command ...

```
qsub -j y -o cgenie_log -V -S /bin/bash runmuffin.sh
cgenie.eb_go_gs_ac_bg_eg.worjh2.BASESFeTDTL
/ BSS.NPZD_Fe.SPIN 10 BENW.worjh2.P04FeTDTL_HYBRID_new.SPIN
```

1. Examine the effect of iron limitation in the new model. What has changed?
2. We can get a more exact picture of the nutrient limitation terms through `eco2D_xGamma_Fe_001` and `eco2D_xGamma_P_001`.

These two variables take values of between 0 and 1. A 1 indicates that the factor is not limiting to growth. A 0 indicates the factor is completely preventing growth.

- In what regions are iron and phosphorus more or less limiting to growth?
 - In regions where neither is limiting, what other factors might be important?
3. Plankton stoichiometry plays a critical role in determining which nutrient is most limiting to growth. You can increase the plankton Fe:C ratio by increasing the minimum and maximum iron quotas. Look at the parameters `eg_qminFe_a` and `eg_qmaxFe_a` in the *user-config* file.
 - What happens to the ecosystem if you increase these parameters by a factor of 2, 5 or 10?
 - How does a change in these parameters affect the model behaviour?
 - What has changed in terms of the patterns of nutrient limitation?

- What has happened to the concentration of the limiting and non-limiting nutrient?

NOTE: It can be risky to just change the parameter value in place, as you might forget what you started with. Instead copy/paste a new version of the line in question, and comment out the original by placing a '#' at the beginning of the line. For example:

```
#eg_qminFe_a = 3.0e-6  
eg_qminFe_a = 6.0e-6
```

4. Nutrient supply ratios are also important in determining the limiting nutrient.
The `bg_par_det_Fe_sol_exp` parameter determines the solubility of atmospheric iron inputs in seawater. Decreasing the value of `bg_par_det_Fe_sol_exp` will therefore decrease the iron-to-phosphorus supply ratio.
 - What happens to the ecosystem if you decrease `bg_par_det_Fe_sol_exp` by 10, 20 or 50%?

9.3 Increasing ecological complexity

In the last lesson we looked at the results of some simulations based on ‘NPD’ and ‘NPZD’ type ecosystem models. Here we will begin to incorporate a bit more ecological complexity.

9.3.1 Plankton size classes

We are going to add a few more plankton size classes, so we have small, medium and large phytoplankton and zooplankton.

1. Save the *ecosystem configuration* file under a new name (e.g. 3P3Z.eco), replacing the existing plankton populations with the ones described in Table 9.1.
2. To run the model with this new configuration, change the name of the *ecosystem configuration file* in the *user-config* file:

```
eg_par_ecogem_plankton_file='3P3Z.eco'
```

3. Save the new *user-config* file under a different name (e.g. BSS.3P3Z.SPIN), and you can now execute the model at the command line:

```
qsub -j y -o cgenie_log -V -S /bin/bash
  runmuffin.sh cgenie.eb_go_gs_ac_bg_eg.worjh2.BASESFeTDTL / BSS.3P3Z.SPIN
  10 BENW.worjh2.P04FeTDTL_HYBRID_new.SPIN
```

Table 9.1: Plankton functional groups and sizes.

<i>j</i>	PFT	Diameter (μm)	<i>j</i>	Functional Type	Diameter (μm)
1	Phytoplankton	0.6	4	Zooplankton	6.0
2	Phytoplankton	6.0	5	Zooplankton	60.
3	Phytoplankton	60.	6	Zooplankton	600.

9.3.2 Viewing 2D time-slice output

Open up the 2D time-slice data for the new model, following the same procedure as in the previous lesson. You will now see a lot more time-slice variables have been loaded into Panoply. We have all the same diagnostics as before, plus some new ones relating to the new plankton populations you have just added. We also have a load of other arrays describing the size distribution and diversity of the photosynthetic community (non-phototrophic populations are ignored in these metrics). These were not included before, because there was only one phytoplankton population.

Size fractions

Variables beginning “eco2D_Size_Frac_...” give the chlorophyll biomass in three size fractions:

1. picophytoplankton (diameter $\leq 2 \mu\text{m}$)
2. nanophytoplankton ($2 < \text{diameter} \leq 20 \mu\text{m}$)
3. microphytoplankton (diameter $> 20 \mu\text{m}$)

Size metrics

Any other variables beginning ‘eco2D_Size_...’ give metrics describing the phytoplankton size distribution.

- eco2D_Size_Mean: Geometric mean¹ phytoplankton diameter, weighted by carbon biomass
- eco2D_Size_Stdev: Geometric standard deviation² of phytoplankton diameter, weighted by carbon biomass.
- eco2D_Size_Minimum: Diameter of smallest phytoplankton contributing >0.1% of the total phytoplankton carbon biomass.
- eco2D_Size_Maximum: Diameter of largest phytoplankton contributing >0.1% of the total phytoplankton carbon biomass.

Diversity metrics

Any variables beginning 'eco2D_Diversity_...' give metrics describing the phytoplankton diversity.

- eco2D_Diversity_Threshold: the threshold diversity index. The number of species contributing >0.1% of the total phytoplankton carbon biomass [Barton *et al.*, 2010].
- eco2D_Diversity_Berger: the inverse Berger(-Parker) index [Berger and Parker, 1970]. The proportion of carbon biomass made up by all but the single most dominant population. For example, if the dominant population accounts for 40% of the total carbon biomass, inverse Berger (-Parker) index is 0.6.
- eco2D_Diversity_Simpson: the inverse (Gini-)Simpson index [Simpson, 1949]. This is effectively the probability that two samples taken at random from the community will be from a different species (note that the probability of selecting a population is dependent on carbon biomass, not cell abundance). If we define the proportional biomass of each species as its relative contribution to the total carbon biomass in the community, the inverse Gini(-Simpson) index is calculated as one minus the sum of the squares of the proportional biomasses of each species.
- eco2D_Diversity_Shannon: the Shannon(-Wiener or -Weaver) index [Shannon, 1948]. With the proportional biomass defined as above, the Shannon index is defined as the sum of [the proportional biomass multiplied by the logarithm of the proportional biomass] for each species.

NOTE: The threshold index is a fairly crude measure of the total number of species in the community, relative to a small and arbitrary threshold of relative biomass. This index is not very sensitive to the relative biomass of individual species (although one very successful species can raise the absolute value of the threshold, thus lowering the diversity). The other three indices do more to quantify the evenness of the community. The more unequal the proportional abundances, the smaller the value of the index. If almost all the abundance is concentrated into one type and all the other types are very rare, the latter three indices can become very small. A community with fewer species, but with more evenly distributed biomass, may well have higher values for these three diversity indices.

Have a look at some of these metrics, but bear in mind that they summarise the diversity of a phytoplankton community that includes just three species. They are probably not that revealing,

¹ We use the geometric mean and standard deviation, because phytoplankton biomass is approximately log-normally distributed across the phytoplankton size range.

² The geometric standard deviation describes the *relative* size range of the phytoplankton. For a geometric standard deviation of σ , ~68.2% of the phytoplankton carbon biomass will be in cells no more than σ orders of magnitude smaller or larger than the geometric mean size.

so we will come back to them later. Instead, have a look some of the other metrics describing the model ecosystem.

- What are the global distributions of the different size classes?
- How do the global biomass distributions compare to variables such as temperature³, or primary production (Uptake Fluxes C)?
- How does nutrient, light and temperature limitation vary between the size classes?
- How can you explain the distribution biomass according to the different limiting factors?

9.3.3 Create your own ecosystem

Save the *ecosystem configuration* file under a new name and add some more plankton populations. Update `eg_par_ecogem_plankton_file` in the *user-config* and save this under a new name. Run the new model.

- How many populations can you get to coexist?
- What effect do the new populations have on the community as a whole?
- What effect, if any, do they have on the strength of the biological pump? (look at `bio_fpart_POC` in `fields_biogem_3D.nc`.)

Just bear in mind that the more populations you put in, the slower the model will run!

References

- A D Barton, S Dutkiewicz, G Flierl, J Bragg, and M Follows. Patterns of diversity in marine phytoplankton. *Science*, 327(5972):1509–1511, 2010.
- W H Berger and F L Parker. Diversity of planktonic foraminifera in deep-sea sediments. *Science*, 168 (3937):1345–1347, 1970.
- C E Shannon. A mathematical theory of communication. *The Bell System Technical Journal*, 27(379-423 and 623-656), 1948.
- E H Simpson. Measurement of diversity. *Nature*, 163:688, 1949.

³Ocean temperature is saved in `fields_biogem_3D.nc`

9.4 Build it up, tear it down

9.4.1 A fully size-structured ecosystem

We are now going to switch to a more diverse version of the size-structured ecosystem model. This configuration has 8 size classes of phytoplankton, and 8 size classes of zooplankton, as shown in Table 9.2.

1. Save the *ecosystem configuration* file under a new name, replacing the existing plankton populations with the ones described in Table 9.2.
2. Update the *user-config* to point to the new *ecosystem configuration* file, and save again under a new name. (It is generally a good idea to make a note of the name and goal of each experiment as you set it up.)
3. Run the new model for at least 20 years (this will probably take about 10-15 minutes).

Table 9.2: Plankton functional groups and sizes.

j	PFT	Diameter (μm)	j	Functional Type	Diameter (μm)
1	Phytoplankton	0.2	9	Zooplankton	0.6
2	Phytoplankton	0.6	10	Zooplankton	1.9
3	Phytoplankton	1.9	11	Zooplankton	6.0
4	Phytoplankton	6.0	12	Zooplankton	19.
5	Phytoplankton	19.	13	Zooplankton	60.
6	Phytoplankton	60.	14	Zooplankton	190.
7	Phytoplankton	190.	15	Zooplankton	600.
8	Phytoplankton	600.	16	Zooplankton	1900.

9.4.2 Ecosystem characteristics

We can now begin to look at the size and diversity metrics of the phytoplankton community in a meaningful way.

1. Look first at (1) the total carbon biomass, (2) the carbon uptake flux (i.e. primary production) and (3) the geometric mean size. Make sure in each case that you are looking at the last year of model output. You may also find it useful to adjust the colour scale, or to change to a logarithmic colour scale (e.g. try a logarithmic scale from 2 to 20 microns for the geometric mean size).

Looking at the maps, we can perhaps pick out three different “biomes” in terms of their community properties:

- (a) The low-latitude oligotrophic gyres are relatively unproductive, and support some of the lowest annual mean biomasses in the surface ocean. In these regions the mean phytoplankton size is very small.
- (b) Subpolar latitudes between 40° and 50° N or S are much more productive, and support very high annual mean biomass. These communities also have the highest mean sizes of any region.
- (c) The polar oceans are also highly productive (except perhaps the high Arctic), and support relatively high annual mean biomasses. These communities are made up (in the model, at least) of slightly smaller phytoplankton than we see in the subpolar regions.

2. What can we find out about the community structure in these regions? Open up some of the other metrics describing the community (standard deviation of size distribution, size fractionation, diversity, limiting factors). What can you find out about the community structure within each region, in terms of coexistence and exclusion?
 - Does the community span a broad or narrow size range?
 - How many size classes are coexisting in each biome?
 - What is the smallest and largest size class in each biome?
 - How much biomass is concentrated in each size fraction (picoplankton, nanoplankton and microplankton)?

What factors do you think are most important in terms of dictating the global distribution of each size class?

To find out the answers to these questions, you are going to pull the model apart, and then put it back together. At each stage the aim is to bring in a different limiting factor, so that you can see its effect on the model behaviour.

The fundamental niche

The first step is to find out the impact of abiotic factors on the distribution of different phytoplankton sizes. In other words, we need to find out what the distribution of the phytoplankton would be in the absence of any ecological interactions, such as resource competition and predation. This is effectively their ‘fundamental niche’.

The fundamental niche is fairly abstract, and not something that can be measured in the real world. In model world, however, we can get a useful estimate of the fundamental niche by making a few simple changes to the model.

1. First of all, you can remove all predation, simply by removing the zooplankton from the *ecosystem configuration* file. Once again, you will need to save a new and appropriately named *user-config* file.
2. Next, you also need to remove all competition for nutrients and light. This involves tweaking the model equations so that the phytoplankton are not nutrient limited, and do not attenuate light. To do this, all you need is to add the following line to the *user-config* file.

```
eg_fundamental = .TRUE.
```

3. If you now run the model (just 10 years should do in this case), you should have a community of eight phytoplankton size classes that are growing solely as a function of the incoming light and the temperature. This growth will be balanced by the basal (i.e. non-grazing) mortality. As there is no feedback between the ecosystem and the environment, populations that can survive will grow exponentially and without limit, potentially reaching astronomical abundance in very little time. Populations that cannot survive will rapidly decline to almost nothing.

The regions in which each plankton shows positive growth defines its fundamental niche. This is a function of abiotic conditions only, and is the absolute limit of its geographical range. Look at the carbon biomass distribution in each size class (set the data range in each case from 0 to 1 mmol C m⁻³)

- How and why does the fundamental niche vary with size?
- Could the limits of the fundamental niche explain some of the patterns seen in the full model?

Resource competition.

The next step is assess the impact of resource competition. We are first going to do this in the absence of any zooplankton grazing.

1. All you need to do at this stage is to re-enable nutrient and light competition. To do this, simply delete 'eg_fundamental = .TRUE.' from the *user-config* file, and save under a new name. Leave the *ecosystem configuration* file as it is.
2. You should have a community of eight phytoplankton size classes that are competing for nutrients and light, again as a function of temperature. This is a much more realistic simulation, as feedbacks between the ecosystem and the environment serve to limit the size of the phytoplankton populations.

Examine the model to find out:

- What size classes are able to persist when resource competition is enabled?
- Why are different size classes more or less abundant in different areas?
- How does the distribution of each size class compare to the fundamental niche?
- What are the reasons for any differences?

Phytoplankton biogeography at this stage begins to approximate the realised niche, which defines the range of conditions that support a population in the presence of ecological interactions. Note that at this stage, however, we have ignored the effects of any predator-prey interactions, as the zooplankton grazers are still missing.

Resource competition + one generalist zooplankton

The previous simulation is clearly unrealistic (although, hopefully, informative). You are now going to add back in just a single zooplankton class, that grazes equally on all plankton (including itself).

1. Add a 100 micron zooplankton into the *ecosystem configuration* file, and save under a new name. Also update the *user-config* file to reflect the change, and save under a similar name.
2. You need to modify the model so that the zooplankton eats all prey with equal preference. This can be done by adding the following lines to the *ecosystem configuration* file.

```
eg_ns=1
eg_pp_sig_a=1.0e99
```

NOTE: For aficionados, the first parameter disables prey-switching (i.e. predators no longer preferentially attack the most abundant prey). The second parameter increases the width of the grazing kernel (i.e. predators can attack a range of prey across a huge size range with equal preference).

3. The addition of zooplankton to the model community should give a more accurate approximation of the realised niche.
 - Does the addition of a single zooplankton grazer enable more or less coexistence?
 - What factors might be responsible for any shifts in biogeography?

Resource competition + one “switching” zooplankton

You began with a full food-web containing 8 phytoplankton and 8 zooplankton size classes. The diversity of zooplankton clearly has an effect on the phytoplankton community that is not seen in the previous experiment. This effect can be imitated with just one generalist zooplankton if we instruct it to graze preferentially on the most successful prey.

Re-enable this 'prey switching' effect by changing the following control parameter to a 2:

```
eg_ns=2
eg_pp_sig_a=1.0e99
```

Compare this simulation to the first experiment (8 phytoplankton and 8 zooplankton) to see how the inclusion of prey switching increases coexistence through the ‘kill-the-winner’ mechanism.

- How does nutrient limitation change with phytoplankton size, and how might zooplankton be affecting this?
- Look at the C:P biomass ratio in the community as a whole, and compare to your estimates from the NPZD model (Lesson 1).
- How does the C:P ratio vary with size? How does having a diverse community affect the coupling of carbon and limiting nutrients?

Further questions to answer

- What sets the fundamental niche, and how does it change with size?
- How is the fundamental niche modified by resource competition?
- What species are favoured in terms of nutrient competition?
- How is the outcome of competition affected by...
 - Abiotic conditions?
 - Increased mortality (through generalist grazing)?
 - Density-dependent mortality (through specialist grazing)?
- Do these experiments tell you all you need to know?
What other modifications can you think of making?

9.4.3 Mixotrophy

Try adding some mixotrophs to the phytoplankton and zooplankton already present in the community. These will have exactly half the nutrient uptake traits of phytoplankton of a similar size, and half the prey capture traits of zooplankton if a similar size.

1. Save the previous *ecosystem configuration* file under a new name.
2. Edit the new *ecosystem configuration* file and add an additional line to add a mixotroph:

```
Mixotroph    xxx    1
```

where xxx is the class size of the mixotrph. (And remember to save it.)

3. Update the *user-config* to point to the new *ecosystem configuration* file, and save again under a new name. (It is generally a good idea to make a note of the name and goal of each experiment as you set it up.)
4. Run the new model for at least 10-20 years.

Questions to explore/answer:

- How does this effect the mean and standard deviation of cell size?
(Size and diversity metrics will be calculated for phytoplankton and mixotrophs together)
- How does mixotrophy effect the C:P ratio of organic matter?
- How does the realised niches of mixotrophs compare to the fundamental niches of phytoplankton?

Also: try replacing all the phytoplankton and zooplankton with a range of sizes of mixotrphs. How does the simulation differ from one with the same size range of separate phytoplankton and zooplankton classes?

9.5 Ecology in a paleo-ocean

9.5.1 The PETM

In this practical we are going to look at the ocean as it *might* have been just over 55 million years ago, at the Paleocene-Eocene Thermal Maximum (PETM) – in short a lot warmer, and with a somewhat different continental configuration and hence ocean circulation.

We are going to make the rather strong assumption that the ecosystem is structured according to exactly the same rules as in the modern ocean, and simply run the model in the new (i.e. old) environment.

To do this, simply evaluate the following text at the terminal command line:

```
./runmuffin.sh cgenie.eb_go_gs_ac_bg_eg.p0055c.BASES / BSS.p0055c.8P8Z.SPIN 1
EXAMPLE.p0055c.RidgwellSchmidt2010.SPIN1
```

which will force a re-compile of the model, and then to run longer:

```
qsub -j y -o cgenie_log -V -S /bin/bash runmuffin.sh
cgenie.eb_go_gs_ac_bg_eg.p0055c.BASES / BSS.p0055c.8P8Z.SPIN 10
EXAMPLE.p0055c.RidgwellSchmidt2010.SPIN1
```

NOTE: This simulation does not include iron as a limiting nutrient. The spatial distribution of bioavailable iron sources is highly uncertain even in the modern ocean, and we simply do not have enough information to justify using iron in this ancient world. Bear that in mind when thinking about your results.

Using what you have learnt so far, and your own resourcefulness, see what you can find out about the Paleocene-Eocene ecosystem.

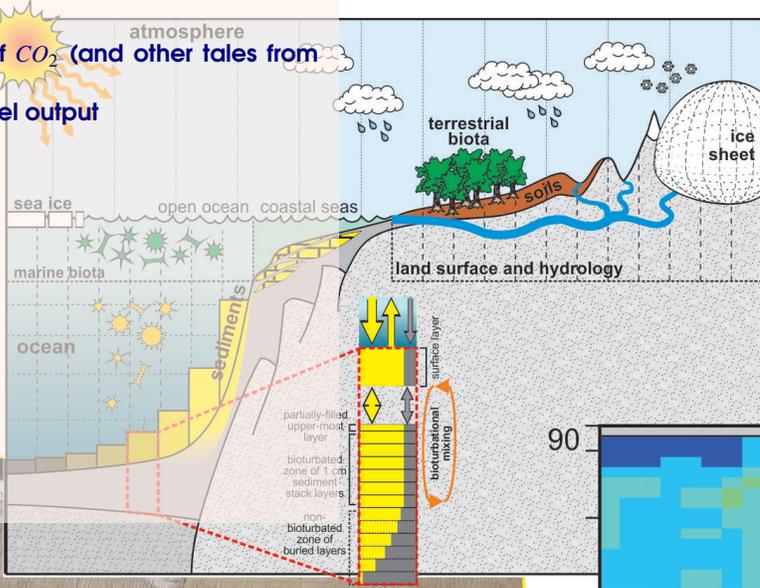
In this warmer world:

- What has happened to the mean plankton size in different regions?
- What has happened to the fundamental niches in different size classes?
- What has happened to the realised niches?
- Is the system more or less productive?
- Has carbon export gone up or down?

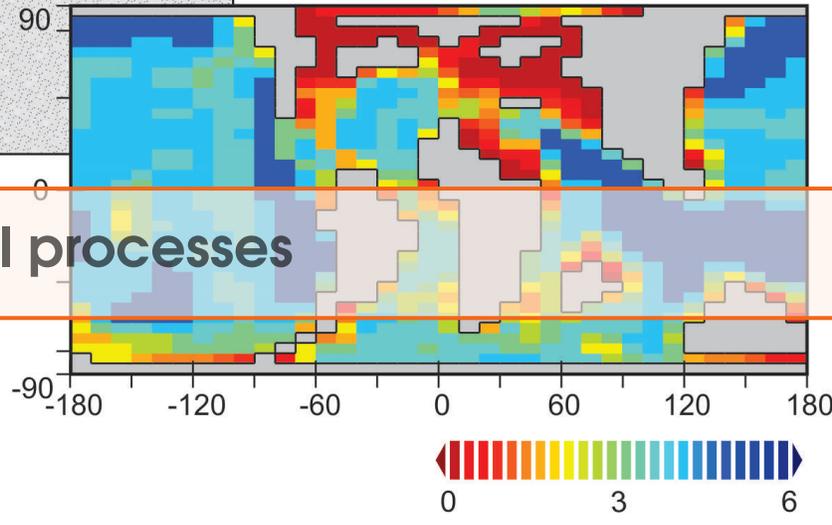
See what you can find out about the two systems and think about the mechanisms that might be responsible for the differences ...

README

The long tail of CO_2 (and other tales from the sediments)
Sediment model output
Further ideas



10. Geological processes



10.1 README

If you have not been through (and completed!) Session #0100 ('Fossil fuel CO_2 release and ocean acidification') will need to download a restart file prior to embarking on the experiments with modern ocean circulation. To fetch this: change to the `cgenie_output` directory, and type: `$ wget http://www.seao2.info/cgenie/labs/Yale.2016/ EXAMPLE._rwlma.PO4_S18x18.SPIN2.tar.gz` Extract the contents of this archive by typing: `$ tar xfv EXAMPLE._rwlma.PO4_S18x18.SPIN2.tar.gz` You'll then need to change directory back to `genie-main` to run the model.

10.2 The long tail of CO_2 (and other tales from the sediments)

You will be using a 'modern' configuration of cGENIE, but ... it is rather more idealized than you have seen for the modern Earth in previous Labs (although not quite as idealized as for the snowball Earth experiments). It also differs in that in addition to having an ocean carbon cycle (which was omitted from many of the previous Lab experiments) it includes a representation of deep-sea sediments and interaction between the preservation and burial of $CaCO_3$ and ocean chemistry plus balance between weathering and sedimentary burial. For an over-view of the sediment model and what time-scales and nature of carbon cycle interaction between ocean and sediment you can expect, read: Ridgwell and Zeebe [2005] and Ridgwell and Hargreaves [2007].

Take the new model for a test drive by running on from the restart provided:

```
EXAMPLE.\_rwlma.P04\_S18x18.SPIN2.tar.gz
```

This is a steady-state climate+carbon cycle experiment that includes the deposition of $CaCO_3$ in deep-sea sediments and the balance between weathering (solute input to the ocean) and burial (output). Try running ('briefly', but 100 years would not be too tedious for this faster configuration!):

```
$ ./runmuffin.sh cgenie.eb_go_gs_ac_bg_sg_rg_gl._rwlma.BASES
LABS LAB_x.sediments 100 EXAMPLE._rwlma.P04_S18x18.SPIN2
```

Note that the base-config

```
cgenie.eb_go_gs_ac_bg_sg_rg_gl._rwlma.BASE
```

specifies the use of a sediment model 'sg' in GENIE for instance. The degraded resolution (and fewer time-steps per year) is helpful in being run cGENIE on geological time-scales within a reasonable time. It also utilizes a conceptual/idealized continental configuration (similar to as in the snowball Earth experiments).

The user-config LAB_x.sediments is set up with the global carbon cycle is 'open' – that is to say, that there is an input of carbon (and alkalinity) to the ocean from weathering, and a loss due to preservation and burial of $CaCO_3$ in deep-sea sediments. Depending on the state of ocean chemistry (and biology) and weathering, these two fluxes (input and output) do not have to balance, and hence ocean carbonate chemistry (and in turn, atmospheric pCO_2) can change with time. The spin-up may not have the two fluxes (input and output) perfectly balanced and hence before you run any experiments you might want to confirm whether the spin-up provided really is adequately 'spun-up'. (Note that a residual drift can be dealt with if it is relatively small and near linear and you have a control experiment, because any experiment you carry out will likely also incorporate (or be biased) by the same residual drift. Hence running a control gives you something to directly contrast with your experiment minus the control (e.g., a difference map or simple subtraction of global numbers) will give you the effect of whatever parameters you changed in the experiment and corrected for any drift. In previous exercises we were a bit lazy, and difference maps were often created with respect to year 1 of an experiment – strictly, they should have been created relative to the same year of a parallel control experiment, i.e., results at year 100 should have been contrasted with the year 100 results of the control.)

Because you are now considering a rather more complex carbon cycle than before (i.e. now including a number of additional, mostly sediments/weathering processes), it is worth conducting a number of idealized perturbations of the global carbon cycle to get a feel for the sensitivity and time-scale of the system response.

For instance – one illustrative experiment, and which has a parallel to experiments you have conducted previously, is to add a pulse CO_2 release to the atmosphere and track the consequences for atmospheric pCO_2 and ocean chemistry (particularly ‘alkalinity’), and now also e.g. deep sea sediments. To the user-config, add the lines:

```
bg_par_forcing_name=' pyyyz.Fp\ (CO_{2}\) _Fp13\ (CO_{2}\) '
bg_par_atm_force_scale_val_3=8.3333e+016
bg_par_atm_force_scale_val_4=-27.0
```

in order to achieve a 1000 PgC total release in a single year (or multiply by 5 to be comparable-ish to Ridgwell and Hargreaves [2007]) with an isotopic composition of -27‰ – appropriate for a fossil fuel carbon source. Methane derived carbon (e.g. as from hydrates) would be more like -60‰.

Run the model for as long as you dare (or can be bothered) – 1,000 or 2,000 years might be just enough as a minimum to start to see impacts on deep-sea sediments, but 5,000 or 10,000 years would be much better. (You can always submit this to the cluster queue and get on with something else.) FYI: 10,000 years is going to take something like an hour.

Plot the time-series of e.g. atmospheric pCO_2 and compare to the (much shorter experiments) you have carried out before with a simple ocean+atmosphere only system. Compare how quickly atmospheric pCO_2 decays compared to previously GENIE papers (e.g. Ridgwell and Hargreaves [2007]) or other models (e.g. Archer et al. [2009]) (see: <http://www.seao2.info/pubs.html>).and e.g. how the sediments respond (e.g. the time-series of sediment $CaCO_3$ content).

To properly (quantitatively) appreciate the role of ocean-sediment interaction (and weathering) and controlling atmospheric pCO_2 , you need to contrast these experiments with as similar a model configuration as possible, except one having no sediments (or weathering). You can achieve this quite simply: create a new user-config and edit the lines:

```
# set an 'OPEN' system
bg_ctrl_force_sed_closedsystem=.false.
```

to:

```
# set a 'CLOSED' system
bg_ctrl_force_sed_closedsystem=.true.
```

(You do not have to edit the comment line (#) but it will help you remember what you have done.)

What this line does is to force the model to always balance $CaCO_3$ preservation and removal in marine sediments with $CaCO_3$ weathering – i.e. the balance (or imbalance) between sediments and weathering is not allowed to change ocean chemistry and hence acts exactly the same as a configuration without any sediments or weathering (as used before).

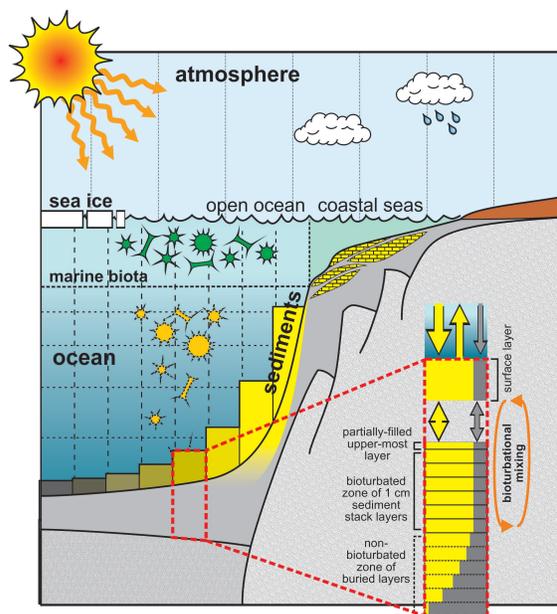


Figure 10.1: Schematic of SEDGEM sediment component.

By comparing the two experiments: can you deduce the effect of the sediments in modulating the atmospheric pCO_2 decline? E.g. compare the pCO_2 time-series. Also view the sediment distribution (of $CaCO_3$): what are the impacts on sediment composition for the experiment with an ‘open’ system? The time-series file of mean global sediment composition `biogem_series_sed_CaCO3.res` (wt% $CaCO_3$) may help illustrate what is going on here. Note that the way the ‘closed’ system is constructed; a response of the sediments is predicted and saved in the output, even though it is not allowed to affect chemistry or atmospheric pCO_2 .

10.3 Sediment model output

There is a whole new set of additional outputs from this configuration of cGENIE, including sediment output (from the ‘SEDGEM’ module). For instance, the composition of the sediments only at the very end of a model experiment (hence unlike BIOGEM, which saves a series of time-slices long) is saved by the SEDGEM module – kill a run before this, and you will get no (or little) output. 2D (e.g. surface sediment properties) results can be found in the `sedgem` subdirectory of your experiment directory and in a netCDF file called `fields_sedgem_2d.nc`. (Note that there is some duplication of results saving, because a series of time-slices of sediment composition are also saved in the 2D biogem netCDF file `fields_biogem_2d.nc` alongside with sea-ice extent etc.). The time-series file: `biogem_series_sed_CaCO3.res` also contains information about how the mean $CaCO_3$ content of surface sediments evolves with time.

For instance, the 2D distribution of wt% $CaCO_3$ – which is the weight fraction of calcium carbonate ($CaCO_3$) in the surface sediments of the deep ocean (i.e., how much plankton carbonate shell material is there compared to other stuff in the mud at the bottom of the ocean?) is saved under a variable called: `sed_CaCO3`. How much carbonate material there is tells you both something about how many carbonate shell secreting plankton were living at the ocean surface above and what

is the chemistry of the deep ocean like that these tiny shells were preserved and did not dissolve. To gauge to what degree the faster configuration of cGENIE might provide an adequate representation of the interaction between ocean chemistry and sediment composition (e.g., in CaCO₃ buffering of CO₂ release and 'carbonate compensation'), the output should be contrasted to observational-based maps as well as (higher resolution) model results (e.g., as presented by Ridgwell and Hargreaves [2007]). Appreciate that cGENIE does not reproduce reality ... particularly at this lower resolution, but does it get the broad patterns right (is it more right than wrong, or more wrong than right)? Do you think the model-data misfits might be important? (Note that there is a reconstruction of the glacial pattern of CaCO₃ in sediments that is available and may be of use in constraining your glacial CO₂ hypothesis, or at least testing it against data.)

The model also generates artificial sediment 'cores' (e.g. see: Ridgwell [2007]) and hence what one might expect to see of your applied perturbations recorded in a sediment core recovered from the ocean floor. In the sedgem results sub-directory, there is a netCDF file which contains all the locations selected (if any) – sedcore.nc. These are not really aligned with latitude as the Panoply display might suggest – the locations are in fact distributed from all over the ocean (Panoply is being fooled in order to display them together). In the SEDGEM 2D netCDF file, these locations are marked in the netCDF variable grid_mask_sedcore. The locations of these cores are stored in a little ASCII 'map' of the ocean in the directory:

```
~/cgenie.muffin/genie-sedgem/data/input
```

and the file for this experiment is given by the parameter: sg_par_sedcore_save_mask_name. Simply by editing (using the ASCII text editor) a '0.0' to a '1.0', you can get the model to generate and save a sediment 'core' at that location.

The sedcore.nc. variables are displayed as a function of 'latitude'. Variables include:

- phys_layer – sediment layer number (counting down).
- phys_depth – (cumulative) depth below surface, measured from the sediment surface to the mid-point of each sediment layer (cm).
- th (cm) – thickness of each sediment layer (cm).
- age_CaCO₃ – the mean age of CaCO₃ particles in a sediment layer. Note that this will not be defined if there is no CaCO₃ preserved.
- ... then some alternative ways of assigning a chronology to a sediment core ... (ignore).
- phys_porosity – sediment layer density (as if you cared!).
- sed_POC and sed_POC_13C – mean organic matter content of each sediment layer and its $\delta^{13}C$. But note: in this configuration no organic matter is preserved (hence all zeros for POC).
- sed_CaCO₃ and sed_CaCO₃_13C – mean CaCO₃ content (wt%).
- sed_det and sed_ash – the wt% detrital and 'ash' contents of a layer (ash is used as a conservative numerical sediment tracer in order to mark the depth of the start of the experiment).

Obviously – you could plot e.g. CaCO₃ (or its $\delta^{13}C$) as a function of depth and/or age across and see how your carbon release experiment might be recorded in the marine geological record. How does this compare with e.g. the PETM? Note that the sediment cores reflect not only the material which has accumulated (or not, if it has dissolved ...) during the course of your experiment, but also the material that accumulated during the 50,000 year spin-up. AND, whatever material the sediment core was initialized with to start with. The large interval of first 100% detrital material below which everything is zero simply reflects the initialization of the sediment array in the model. Also note the ash 'peak' near the bottom of the stack (filled) sediment layers – this is a tracer to 'tag' the start of the model spin-up. If you look at the spin-up results (not your recent perturbation

experiment) – the ash peak lies in a sediment layer with age 50,000 years. But why is there any ash deeper than the age corresponding to the start of the spin-up? How can it get there?

10.4 Further ideas

You can also explore the role of weathering and sensitivity of e.g. atmospheric pCO_2 and climate to this (as well as to volcanic CO_2 outgassing). Because the time-scale for seeing interesting stuff happen is now going to be long ... you need some help.

The *user-config* file: `LAB_x.weathering` is provided which uses a ‘trick’ to accelerate the run-time (the new section `# -- GEOCHEM ACCELERATION --`) ... see Lord et al. [2015].

Parameters of ‘interest’ that might be adjusted in the context of weathering and long-term controls on atmospheric pCO_2 (listed under `# -- WEATHERING --`), include:

- `rg_par_outgas_CO2` -- CO_2 outgassing rate in mol yr⁻¹
- `rg_par_ref_T0` -- reference land surface temperature for weathering (C)

You might investigate other facets of the nature of the relationship between ocean and sediments (and weathering) as how climatic (biogeochemical) signals are encoded in the marine geological record. For instance, you could explore the effect/importance of sediment ‘bioturbation’ (e.g. see *Ridgwell* [2007]). Whether the surface sediment layers are bioturbation or not is set by the parameter:

```
sg_ctrl_sed_bioturb=.true.
```

Simply change to `.false.` in order to ‘turn off’ bioturbation.

Rather than driving an initial dissolution of $CaCO_3$ in deep sea sediments, the opposite (initial response) can be obtained by removing CO_2 from the atmosphere (implemented by negative rather than positive emissions). BE CAREFUL here, as for a pre-industrial atmosphere with 278 ppm CO_2 , you do not have a lot more than ~ 600 PgC in there to begin with. So either: remove less than 600 PgC, or remove the carbon over a little longer than 1 year – implement this either by editing the forcing files directly, or set a scaling parameter for the duration.

(Hint: a forcing for removing CO_2 simply has the opposite sign to an addition (emissions) forcing.)

Again – view the time-series of ocean composition (e.g. DIC, ALK, $d13C$) as a function of time, plus mean sediment surface composition (wt% $CaCO_3$). Also view the sediment ‘cores’ and hence what in practice has been incorporated into accumulating sediments as a record of what is a very sharp perturbation at the ocean surface (and atmosphere).

How then is an event characterized by CO_2 removal from the system recorded differently from one characterized by CO_2 release? Are there different implications for constructing core age-scales and chronology, e.g. where in (core) ‘time’ does the excursion maximum appear to lie? Do all sediment locations show identical responses (i.e. does it matter what the initial wt% $CaCO_3$ is?).

Overview (and types of model output)

Time-slice output

- Frequency and timing of *time-slice* data saving
- Experiment end saving
- Seasonal/monthly data saving
- More frequent data saving

Time-series output

- Frequency and timing of *time-series* data saving
- Saving orbital insolation

Data field selection

Re-start files

Useful output

- Physics
- (Bio)Geochemistry
- Biology/Ecology

11. cGENIE.muffin model output

This section covers **cGENIE.muffin** saves data and how to ensure that the variables you want are saved and when you want.

- Overview.
- *Time-series* output.
 - *Time-series* file naming conventions.
 - Specifying frequency and timing of *time-series* data saving.
 - Seasonal/monthly data saving.
- *Time-slice* output.
 - *Time-slice* file naming conventions.
 - Specifying frequency and timing of *time-slice* data saving.
- Specifying which data fields to be saved in the *time-series* and *time-slice* format.
- *Re-start* files.

11.1 Overview (and types of model output)

The results of experiments are written to the directory:

```
~/cgenie_output
```

For any particular experiment, all saved model results, plus copies of input parameters and the model executable, are gathered together in a directory that is assigned the same name as the experiment (== *user-config* file name), e.g.:

```
EXAMPLE.worbe2.Ridgwelletal2007.SPIN
```

Every science module saves its results in its own individual sub-directory within the experiment directory. So for the module that calculates ocean biogeochemical cycles – **BIOGEM**, the results files will thus be found in:

```
~/cgenie_output/EXAMPLE.worbe2.Ridgwelletal2007.SPIN/biogem
```

Note that **ATCHEM** does not save its own results (**BIOGEM** can save information about atmospheric composition and air-sea gas exchange) while **SEDGEM** essentially saves results only at the very end of a model experiment (**BIOGEM** can also save the spatial distribution of sediment composition as time-slices as well as mean composition as a time-series). Furthermore, in order to attain a common format for both ocean physical properties and biogeochemistry, **BIOGEM** can save a range of ocean results in addition to temperature and salinity, such as: velocities, sea-ice extent, mixed layer depth, convective frequency, etc. Also note that **SEDGEM** saves data only at the end of an experiment¹.

Saving full spatial distributions for any or all of the tracers at each and every time-step is not practical, not only in terms of data storage but also because of the detrimental effect that repeated disk access has on model performance. Instead, **BIOGEM** saves the full spatial distribution of whatever tracer, flux, and/or physical properties of the system are required (how what fields are required is specified is discussed later), only at one or more predefined time points (in years). These are called '*time-slices*'. However, rather than taking an instantaneous snapshot, the time-slice is constructed as an average over a specified integration interval. The second main data format for model output is that of a '*time-series*' of change in a single (integrated) property of the Earth system. Model characteristics must be reducible to a single meaningful variable for this to be practical (i.e., saving the time-varying nature of 3-D ocean tracer distributions is not). Suitable reduced indicators include: the total inventories in the ocean and/or atmosphere of various tracers (or equivalently, the mean global concentrations / partial pressures, respectively), global sea-ice coverage. Like *time-slices*, the data values saved in the *time-series* files represent averages over a specified integration interval (one year by default). For both *time-slices* and *time-series* output, the files themselves are created during model initialization and are periodically updated (appended to) during the experiment. Hence, even before the experiment has finished they may contain data that is useful to view and can be used to check on the progress of an experiment.

ATCHEM

In the **ATCHEM** results directory, only the following file will be present:

1. `_restart.nc` – *Re-start* file – a snap-shot of the 2D distribution of atmospheric composition at the very end of the experiment. Not intended for user-access, although it can be plotted just like any normal *netCDF* format file.

¹With the exception of sediment core location environmental properties, which are saved more frequently.

BIOGEM

For **BIOGEM**, some or all of the following files will be present:

1. `_restart.nc` – *Re-start* file – a snap-shot of the 3D distribution of biogeochemical properties of the ocean at the very end of the experiment. Not intended for user-access, although it can be plotted just like any normal *netCDF* format file.
2. `fields_biogem_2d.nc` – 2-D fields of (mostly) ocean bottom, ocean surface, and sediment surface properties.² Also: water-column integrals of certain geochemistry diagnostics, air-sea gas exchange fluxes, atmospheric composition.
3. `fields_biogem_3d.nc` – 3-D fields ocean dissolved and particulate tracer properties.³
4. `biogem_series_*.res`⁴ – *Time-series* results files – globally and surface-averaged (and sometimes also benthic (bottom) surface averaged) property values as a function of time in plain text (ASCII) format.
5. `biogem_year*_diag_GLOBAL.res` – Miscellaneous global diagnostic information. It is saved at each requested time-slice with the file-name string containing the mid-point of the time-slice (as years). The diagnostics include:
 - time mid-point and integration interval
 - global ocean surface area and volume
 - mean global air-sea gas exchange coefficient (for CO₂)
 - mean atmospheric tracer concentrations plus total inventory
 - mean ocean tracer concentrations plus total inventory
 - mean plus total global productivity
 - mean plus total global sedimentation

SEDGEM

In the **SEDGEM** results directory, some or all of the following files will be written:

1. `_restart.nc` – *Re-start* file – a snap-shot of the 2D distribution of sedimentary properties at the very end of the experiment. Not intended for user-access, although it can be plotted just like any normal *netCDF* format file.
2. `fields_sedgem_2d.nc` – Contains 2-D fields of sediment surface and ocean bottom properties.⁵
3. `sedcore.nc` – *netCDF* format file containing the stacked records of accumulated deep-sea sediment composition.

The locations (if any) of sediment cores to be saved is specified in a plain text (ASCII) file pointed to by the string value of the *namelist* parameter `sg_par_sedcore_save_mask_name`⁶. In the mask file, a '1' indicates a location to save a sediment core at, and a '0' indicates that no sediment core should be saved at this location. This file must be present, so to save no sediment cores, simply populate the file with all zeros in an `xx by yy` grid.

²The mid-points at which time-slices are saved are specified as described above.

³The mid-points at which time-slices are saved are specified as described above.

⁴`.res` is a useful format for processing in **MATLAB**; for other programs, other extensions are needed. If using the Mathematica data processing scripts - see `genie-docs/cGENIE.AutomationScripts` - `.dat` is needed; this can be set with `gm_string_results_ext=".dat"`

⁵This data is saved only at the termination of an experiment (i.e., the *netCDF* file contains only a single time-slice).

⁶The location of this file is specified by the **SEDGEM** data input directory *namelist* parameter: `sg_par_indir_name` which by default is `~/genie-sedgem/data/input`.

4. `sedcoreenv_*` – These files contain pseudo time-series of surface sediment environmental properties at each of the requested sediment core locations (if any are chosen).
5. `seddiag_misc_DATA_GLOBAL.res` – A summary of mean global sedimentation, dissolution, and preservation fluxes, and surface sediment composition.
6. `seddiag_misc_DATA_FULL.res` – Surface sediment and bottom water properties at each and every sediment grid point.

ROKGEM

In the **ROKGEM** results directory, some or all of the following files will be written:

1. `fields_rokgem_2d.nc` – 2-D fields of (mostly) land surface, ocean surface, and atmospheric properties related to weathering.
2. `biogem_series_*` – Time-series results files.

11.2 *Time-slice* output

11.2.1 Frequency and timing of *time-slice* data saving

Rather than taking an instantaneous snapshot, the time-slice is averaged over a specified integration interval Δt (in years), defined by the parameter `bg_par_data_save_slice_dt`⁷. The model state is thus integrated from time $t_n - \Delta t/2$ to $t_n + \Delta t/2$. For instance, setting a value of $\Delta t = 1.0$ year results in all seasonal variability being removed from the saved time-slices, and successive time-slices then only reflect long-term (>1 year) trends in system state.

The mid-point years (t_n) for which time-slices should be saved are specified in a single column plain text (ASCII) file in the `cgenie.muffin/genie-biogem/data/input` directory, whose name is specified by the parameter `bg_par_infile_slice_name`⁸. For example, the default *time-slice* specification file `save_timeslice.dat` contains the specification⁹:

```
-START-OF-DATA-
0.5
1.5
4.5
9.5
19.5
49.5
99.5
199.5
499.5
999.5
1999.5
4999.5
9999.5
19999.5
49999.5
99999.5
199999.5
499999.5
999999.5
-END-OF-DATA-
```

where `-START-OF-DATA-` and `-END-OF-DATA-` are simply tags delineating the start and end of the time point data. Use of this particular specification lends itself to simple experiment run durations to be adopted (e.g., 10, 100, 10000 years). It provides a good generic starting point in that save frequency is faster to begin with (when environmental variables are more likely to be rapidly changing) and less frequently later (when environmental variables are unlikely to be changing rapidly and maybe converging to steady-state).

To change the time points used for *time-slice* data saving, either direct edit this file (less good), or create a new file (e.g. simply copy and rename `save_timeslice.dat`) with the required save

⁷An empty list is valid - time-slices will then be populated for you at an interval set by the time-slice integration interval. But if you really don't want any time-slices, just set the first (or only) time point to occur beyond the end year of the run.

⁸The location of this file is specified by the **BIOGEM** data input directory parameter: `bg_par_indir_name` which by default is `~/genie-biogem/data/input`.

⁹The order in which the time sequence is ordered (i.e., ascending or descending time values) does not actually matter in practice as long as the list of times is ordered sequentially. The list will be internally re-ordered if necessary according to the selection of 'BP' (the model running backwards-in-time) or not according to the logical value of the parameter `bg_ctrl_misc_t_BP`, which is `.false.` by default.

frequency and timing and saved to the `cgenie.muffin/genie-biogem/data/input` directory, with the parameter `bg_par_infile_slice_name` pointing to the new filename).

11.2.2 Experiment end saving

Just in case an experiment run duration is chosen such that there is no corresponding save point anywhere near the end of the run, a *time-slice* is automatically saved at the very end of an experiment regardless of whether one has been specified or not and with the same averaging as used for the specified *time-slices*.

11.2.3 Seasonal/monthly data saving

Time-slice (but not currently *time-series*) data can be saved seasonal or even monthly by selected by setting a single parameter rather than e.g. specifying a monthly or seasonal data save interval and editing the time-slice definition file with a series of min-points for months (or seasons). The way it works is that the overall averaging interval (parameter: `bg_par_data_save_slice_dt`)¹⁰ is broken down into sub-intervals of averaging. i.e., breaking down a year interval (the default) into 4 will give seasonal averaging. The parameter: `bg_par_data_save_slice_n` where *n* sets the number of time steps in each sub-interval of data saving and hence determines whether the averaging is e.g. seasonal or monthly. The slightly tricky part is to be sure of how many time steps in each year ;)

By default, **cGENIE.muffin** employs 96 time-steps per year for a 16-level ocean circulation model (**GOLDSTEIN**) and 48 for **BIOGEM**¹¹. Hence for a 16-level ocean configuration, seasonal data saving would be obtained with:

```
bg_par_data_save_slice_n=12
```

(12 **BIOGEM** steps per averaging interval out of a total of 48), and monthly averages with:

```
bg_par_data_save_slice_n=4
```

(i.e. 4 **BIOGEM** steps for each of the 12 monthly averaging intervals, giving of a total of 48).

For lower resolution configurations of **cGENIE.muffin**, **GOLDSTEIN** may be operating on 48 time-steps per year, and **BIOGEM** on 24 or even 12. As **cGENIE.muffin** starts up it will report the ocean and biogeochemical time-stepping, such as:

```
>> Configuring ...
    Setting time-stepping [GOLDSTEIN, BIOGEM:GOLDSTEIN]: 100 2
```

which specifies 100 time-steps per year for **GOLDSTEIN**, and 50 per year (100/2) for **BIOGEM** for the case of a 16 level ocean using the `runmuffin.t100.sh` run script. Note that for every year mid-point specified in the *time-slice* specification file, 4 or 12 (for seasonal and monthly, respectively) times as many time-slices will actually be saved.

¹⁰Default value = 999

¹¹Note that when running using `runmuffin.t100.sh`, 100 time-steps are taken in the ocean and 50 in **BIOGEM** for a 16 level ocean model.

11.2.4 More frequent data saving

Explicit frequent saving of fields or properties at specific locations can be done by setting a more higher save frequency of the time-slice data. However, because the 2D and 3D fields may contain a variety of unwanted variables in addition to the target one, save frequency is likely to be limited by the maximum netCDF file size that can sanely be manipulated. The practical maximum is around 100, depending on the number of types of data field selected to be saved. Several alternative options are available:

- (trivial) Make do with global or surface (or benthic) means in the time-series output.
- Cut down the types of data saved to the absolute minimum (see 'Data field selection' below).
- Save only 2D data. This can be accomplished by setting the parameter:

```
bg_ctrl_data_save_2d=.true.  
bg_ctrl_data_save_3d=.false.
```

(both are `.true.` by default). This disables the 3D data saving, although an empty netCDF file will still be created.

- Save the tracer fields in 3D, but at the save frequency of time-series data saving¹². This can be done by setting:

```
bg_ctrl_data_save_3d_sig=.true.
```

(by default, `.false.`).

¹²This is in addition to normal 3D saving at the time-slice data saving frequency

11.3 Time-series output

11.3.1 Frequency and timing of *time-series* data saving

For results *time-series*, a file containing a series of model times (t_n) at which data points need be saved is defined in the same way as for *time-slices*, with the filename specified by the parameter `bg_par_infile_sig_name`. Again, the data values saved in the time-series file do not represent discrete values in time but an average, calculated from time $t_n - \Delta t/2$ to $t_n + \Delta t/2$ as per the construction of time-slices. The averaging interval, Δt , is set by the value of the parameter `bg_par_data_save_sig_dt`. The format is also identical to before (with tags delineating the start and end of the list of mid-points). If there are less than two elements present in the list, a default frequency of data saving will be invoked, set equal the averaging interval, except in the situation that this results in an unreasonably large amount of data, when an order of magnitude (or more than one order of magnitude where necessary) fewer save points are assumed.¹³ The default setting:

```
bg_par_infile_sig_name='save_timeseries.dat'
```

provides for reasonably generic data saving, with the save frequency faster to begin with and becoming progressively less frequently later.

There is a related facility to `bg_par_infile_sig_name` for **SEDGEM** and **ROKGEM** in the parameter: `xx_par_output_years_file_0d`, where `xx` is `sg` for **SEDGEM** and `rg` for **ROKGEM**. These specify files in the `genie-*gem/data/input` directory and again contain a list of years for OD (time-series) output to be generated at. However, unlike **BIOGEM**, the data saved *do* represent discrete values in time and *not* e.g. annual averages.

11.3.2 Saving orbital insolation

For evaluating and checking orbital experiments in **cGENIE.muffin**, it is useful to be able to extract commonly used diagnostics of orbital variations in insolation, e.g. June 21 65N. **cGENIE.muffin** can be configured to save the *time-series* (at the standard save frequency for *time-series* output) at two different 'j' grid positions (latitude) – typically one Northern and one Southern hemisphere, and at specific points in the seasonal cycle. The relevant parameters are:

- `bg_par_sig_j_N` Sets the 'j' latitude grid point location for extracting the insolation. On a 18×18 grid, a value of 17 is as close as you are going to get to 65N.
- `bg_par_sig_j_S` As above, except for the Southern hemisphere (although there is nothing stopping you from choosing 2 Northern or 2 Southern hemisphere points, just what out for the automatic output column labelling that might cause confusion).
- `bg_par_t_sig_count_N` Sets the time-step in the **BIOGEM** tie-stepping cycle at which the insolation value is extracted. Care is needed here in distinguishing between the primary ocean (**GOLDSTEIN**) time-step, and **BIOGEM**, which is typically sub-stepped. For example, on an $18 \times 18 \times 8$ grid, the **GOLDSTEIN** time-step is 48 (per year), and **BIOGEM** time-steps every other **GOLDSTEIN** step, i.e. 24 per year. A value for this parameter of 12 is then approximately June 21.
- `bg_par_t_sig_count_S` As above, except for the Southern hemisphere.

The *time-series* output files is called: `biogem_series_misc_ocn_insol.res` (and has pretty self-explanatory columns).

¹³For historical reasons ... the maximum number of time-series (and time-slice) data points was set to 4096. This is set by the parameter `n_data_max` in `biogem_lib.f90` and can be altered if required.

11.4 Data field selection

Model output – both *time-slice* and *time-series* data are saved in blocks or by categories of model variables. For instance, all dissolved tracers in the ocean (3D netCDF *time-slice* and/or *time-series*), or all particle flux fields, all carbonate chemistry and associated variables, all surface sediment composition, etc etc. This still requires a multitude of parameters, one for each category and generally also one for each of *time-slice* and *time-series* data. In an attempt to simplify this, a single parameter, `bg_par_data_save_level`, specifying the sort and amount of data to save can be set instead.

The value of the `bg_par_data_save_level` save level parameter is given as an integer between 0 and 99, and has the following effect:

- 0 – Save nothing.
- 1 – Minimum – basic geochemistry only, i.e. ocean and atmosphere tracer fields (omitting e.g. the miscellaneous fields and time-series – see below).
- 2 – Basic output == basic geochemistry and physics.
- 3 – Basic + biology diagnostics, i.e. ocean and atmosphere tracer fields plus particulate flux fields and biological diagnostics such as limitations on export.
- 4 – Basic + geochemistry diagnostics, including output on air-sea gas exchange, ocean carbonate chemistry, and geochemical diagnostics such as remineralization rates and transformation, ocean pH field (3D), Fe cycle and speciation diagnostics (2D). In conjunction with the **ROKGEN** module, also: weathering fluxes.¹⁴
- 5 – Basic + biology + geochemistry diagnostics. A combination of 3 and 4.
- 6 – Basic + tracer diagnostics. Tracer diagnostics includes: N*, P* etc., water column inventories (2D).
- 7 – Basic + tracer + proxy diagnostics. Proxy diagnostics includes: ocean surface and benthic (and surface-benthic) tracers (2D). Also trace metals (e.g. Cd).
- 8 – Basic output + biology + geochemistry + tracer + proxy diagnostics.
- 9 – Basic output + full physics (e.g. all grid specifications and properties).
- 10 – Ocean acidification option == basic geochemical output fields plus all carbonate chemistry.
- 99 – Save everything.
- >99 – Use explicit user-specified settings for individual save categories. This is the default and is broadly consistent with previous version of the model.

All of options 2–11 save as *time-slices*:

- atmosphere tracer fields (2D)
- ocean tracer fields (3D)
- various miscellaneous diagnostics, including: ocean velocity (3D), overturning stream-function (2D), sea-ice extent and thickness (2D), incident radiation (2D), convection diagnostics (2D), air-sea gas exchange diagnostics (2D).
- core-top sediment composition fields (2D) (if **SEDGEN** is selected)

and as *time-series*:

- atmosphere tracer properties (as: atmospheric inventory (in *mol*) and concentration (mol kg^{-1}) or isotopic composition)
- ocean tracer properties (as: ocean inventory, plus mean (whole) ocean, or isotopic composition)

¹⁴The most common option.

- mean surface and benthic tracer properties
- various miscellaneous diagnostics, including: insolation, sea-ice extent, volume, and thickness; global overturning stream-function, ocean surface pH, land surface temperature, and Fe parameters.
- sediment (core-top) composition data (if **SEDGEM** is selected)

In addition, further output will be automatically added to the suite of saved data depending on the module selected and also for certain sorts of *forcing*.

11.5 Re-start files

Re-start files are saved in the results directories of each module. For **ATCHEM**, **BIOGEM**, and **SEDGEM**, these are in *netCDF* format.

For the climate modules of **cGENIE.muffin** (**GOLDSTEIN**, **GOLDSTEIN-SEAICE**, **EMBM**), *re-start* files can be selected to be saved in either plain text (ASCII) or *netCDF* format. ASCII format is the current default.

11.6 Useful output

What follows is a short summary of some of the output and how it can be used.

Note – depending on the specific model configuration (which model modules are selected) and selected tracers, as well as specific output choice, not all these variables and files will be present in the model output.

11.6.1 Physics

Filename	Data	Application
biogem_series_*.res		
atm_humidity	Mean surface humidity. (?)	(rarely used)
atm_temp	Mean surface temperature. (degrees C)	Climate and climate change, when a simple global diagnostic is needed.
misc_opsi	Global minimum (-ve) and maximum (+ve) overturning streamfunction, also reported for Atlantic and Pacific basins. Units of Sv. (For certain modern configurations.)	Simple diagnostic of large-scale ocean circulation. There is some relationship of the maximum (negative and positive) overturning to ocean ventilation.
misc_seaice	Sea-ice fractional cover (\%), thickness (m), and volume (m ³).	As a simple climate (change) diagnostic.
misc_SLT	Mean global surface land temperature. (C)	For calibrating and analysing global weathering rates.
ocn_sal	Mean global, and typically also surface and benthic, ocean salinity. (PSU)	For characterizing freshwater changes and salinity <i>forcing</i> impacts.
ocn_temp	Mean global, and typically also surface and benthic, ocean temperature. (degrees C)	Climate and climate change.

Table 11.1: Summary of the main (useful) *time-series* output for climate-only ('physics') investigations.

variable	variable (long name)	Description	Application
atm_humidity	specific humidity	surface humidity (?)	(rarely used)
atm_temp	surface air temperature"	surface air temperature (degrees C)	climate patterns and anomalies, comparison with terrestrial temperature proxies
grid_mask	land-sea mask	land-sea mask (n/a)	copy-paste-edit to create masks for data analysis
grid_topo	ocean depth	ocean depth (m)	
ocn_sur_sal	surface-water sal	surface ocean salinity (PSU)	diagnosing freshwater forcing impacts, regions of likely deep-water formation
ocn_sur_temp	surface-water temp	surface ocean temperature (degrees C)	diagnosing ocean circulation patterns, pole-to-equator temperature gradients, surface ocean temperature proxy comparisons
ocn_ben_temp	bottom-water temp	benthic temperature (C)	diagnosing ocean circulation patterns, benthic temperature proxy comparisons
phys_cost	convective cost	rate of convective adjustments anywhere in the water column (n/a)	diagnosing deep mixed layers (and light limitation of biology) and deep-water formation regions
phys_opsi	Global streamfunction	global overturning stream-function (Sv)	diagnosing large-scale ocean circulation patterns, sources of deep-water formation, deep ocean ventilation
phys_psi	Barotropic streamfunction	barotropic streamfunction (Sv)	diagnosing wind-driven ocean circulation patterns, effect of gateways
phys_seaice	sea-ice cover (%)	sea-ice cover (%)	(climate / sea-ice)
phys_seaice_th	sea-ice thickness	sea-ice thickness (m)	(climate / sea-ice)

Table 11.2: Summary of the main (useful, physics-focussed) 2D time-slice output.

variable	variable (long name)	Description	Application
ocn_sal	salinity	ocean salinity (PSU)	diagnosing ocean circulation patterns
ocn_temp	temperature	ocean temperature (C)	diagnosing ocean circulation patterns
phys_u	ocean velocity - u	Eastwards component of ocean velocity (m/s)	diagnosing ocean circulation patterns and currents
phys_v	ocean velocity - v	Northwards component of ocean velocity (m/s)	diagnosing ocean circulation patterns and currents
phys_w	ocean velocity - w	upwards component of ocean velocity (m/s)	diagnosing ocean circulation patterns (note that velocity is measured at the top of an ocean depth layer, hence n/a for the surface layer)

Table 11.3: Summary of the main (useful) 3D time-slice output.

11.6.2 (Bio)Geochemistry

Filename	Data	Application
biogem_series_*.res		
atm_pCO ₂	Global inventory (<i>mol</i>), mean concentration (<i>atm</i>) of atmospheric CO ₂ .	Drivers of and feedbacks with climate. Diagnostic of response to carbon emissions (and removal).
atm_pCO ₂ _13C	¹³ C inventory (<i>mol</i>) and δ ¹³ C of atmospheric CO ₂ .	Diagnostic of carbon emissions (and removal). Comparison with (terrestrial) proxy δ ¹³ C data.
atm_pO ₂	Global inventory (<i>mol</i>), mean concentration (<i>atm</i>) of atmospheric O ₂ .	Limited use. Checking on <i>restoring forcing</i> of atmospheric O ₂ . Impacts of C _{org} burial if included in model.
carb_sur_conc_*	Carbonate chemistry components (mean surface) (<i>molkg</i> ⁻¹).	Not generally useful.
carb_sur_H	Surface ocean mean [H ⁺] (<i>molkg</i> ⁻¹).	More useful is pH – reported under misc (see below).
carb_sur_ohm_arg	Mean surface aragonite saturation.	Ocean acidification impacts of CO ₂ release. Weathering impacts. Relates to carbonate production by (modern) corals, pteropods.
carb_sur_ohm_arg	Mean surface calcite saturation.	Ocean acidification impacts of CO ₂ release. Weathering impacts. Carbonate production by foraminifera and coccolithophorids.
diag_misc_specified_forcing_*	Applied flux <i>forcings</i> (<i>molyr</i> ⁻¹).	Whenever a <i>restoring</i> , or <i>flux forcing</i> is specified, the actual flux employed, is saved here. Useful for diagnosing the flux associated with a restoring forcing (e.g. allowing emissions flux associated with RCP (<i>restoring forcing</i>) scenario to be diagnosed.)
fexport_CaCO ₃	Total flux (<i>molyr</i> ⁻¹) and flux density (<i>molm</i> ⁻² <i>yr</i> ⁻¹), of CaCO ₃ export from the ocean surface.	Carbonate production. Impacts of ocean acidification.
fexport_POC	Total flux (<i>molyr</i> ⁻¹) and flux density (<i>molm</i> ⁻² <i>yr</i> ⁻¹), of POC export from the ocean surface.	Particulate organic matter export. Impacts of changes in nutrient supply and limitation.
misc_surpH	Mean surface <i>pH</i> .	Ocean acidification.
ocn_DIC_13C	Global inventory (<i>mol</i>), mean global, surface, and benthic δ ¹³ C.	Carbon release and removal. Surface-benthic – indicator or strength of carbon export and the biological pump, as well ocean ventilation.
ocn_O ₂	Global inventory (<i>mol</i>), mean global, surface, benthic concentrations (<i>molkg</i> ⁻¹) of O ₂ .	Indication of changes in ocean anoxia. (long-term) Imbalances between burial and weathering.
ocn_PO ₄	Global inventory (<i>mol</i>), mean global, surface, benthic concentrations (<i>molkg</i> ⁻¹) of PO ₄ .	Nutrient limitation. (long-term) Imbalances between burial and weathering.
ocn_TDFe	Global inventory (<i>mol</i>), mean global, surface, benthic concentrations (<i>molkg</i> ⁻¹) of dissolved Fe.	Nutrient limitation. (long-term) Imbalances between burial and weathering.

Table 11.4: Summary of the main (useful, plus notes on a few less used) *time-series* output for (bio)geochemistry (non ecological) investigations.

variable	variable (long name)	Description	Application
atm_*		Distributions of gases and isotopes.	Not useful as the atmosphere is well-mixed. The <i>time-series</i> outputs are simpler and more useful.
carb_ben_ohm_arg carb_sur_ohm_cal		Benthic aragonite and calcite saturation.	Impacts of ocean acidification of distribution of benthic organisms. Indicator of sediment preservation.
carb_sur_ohm_arg carb_sur_ohm_cal		Ocean surface aragonite and calcite saturation.	Impacts of ocean acidification of distribution of planktic organisms.
fseadair_pCO ₂	pCO ₂ : net sea->air gas exchange flux density	Air-sea CO ₂ gas exchange.	Indicator of air-sea gas disequilibrium, regions of out-gassing/in-gassing.
misc_pH misc_sur_ rCaCO ₃ toPOC	ocean pH CaCO ₃ to POC export rain ratio	Ocean surface pH. Particulate CaCO ₃ : POC export ratio from ocean surface.	Ocean acidification. Ocean acidification impacts.
ocn_sur_TDFe	surface-water TDFe	Ocean surface total dissolved Fe (<i>molkg</i> ⁻¹).	Patterns of nutrient uptake and limitation.
ocn_sur_TDL	surface-water TDL	Surface ligand concentrations (<i>molkg</i> ⁻¹).	(Stabilizes dissolved Fe, but so not useful itself.)
ocn_sur_PO4	surface-water PO4	Ocean surface [PO ₄] (<i>molkg</i> ⁻¹).	Patterns of nutrient uptake and limitation.
ocn_ben_PO4	bottom-water PO4	Benthic [PO ₄] (<i>molkg</i> ⁻¹).	Indicator of large-scale ocean circulation and ventilation.
ocn_ben_DIC_13C		Benthic δ ¹³ C.	Indicator of large-scale ocean circulation and ventilation. Model-data δ ¹³ C proxy comparison.
ocn_int_DIC	DIC water-column integrated tracer inventory	Pattern of water column integrated ocean DIC (i.e. dissolved carbon storage) (<i>molm</i> ⁻²).	Indicator of CO ₂ emissions storage and transport when used in difference/anomaly maps and calculations.

Table 11.5: Summary of the main (mostly useful) 2D time-slice output for (bio)geochemistry.

11.6.3 Biology/Ecology

Plotting with Excel

Plotting with Panoply

Issues with Panoply default settings

Basic plots – examples

Difference (anomaly) plots

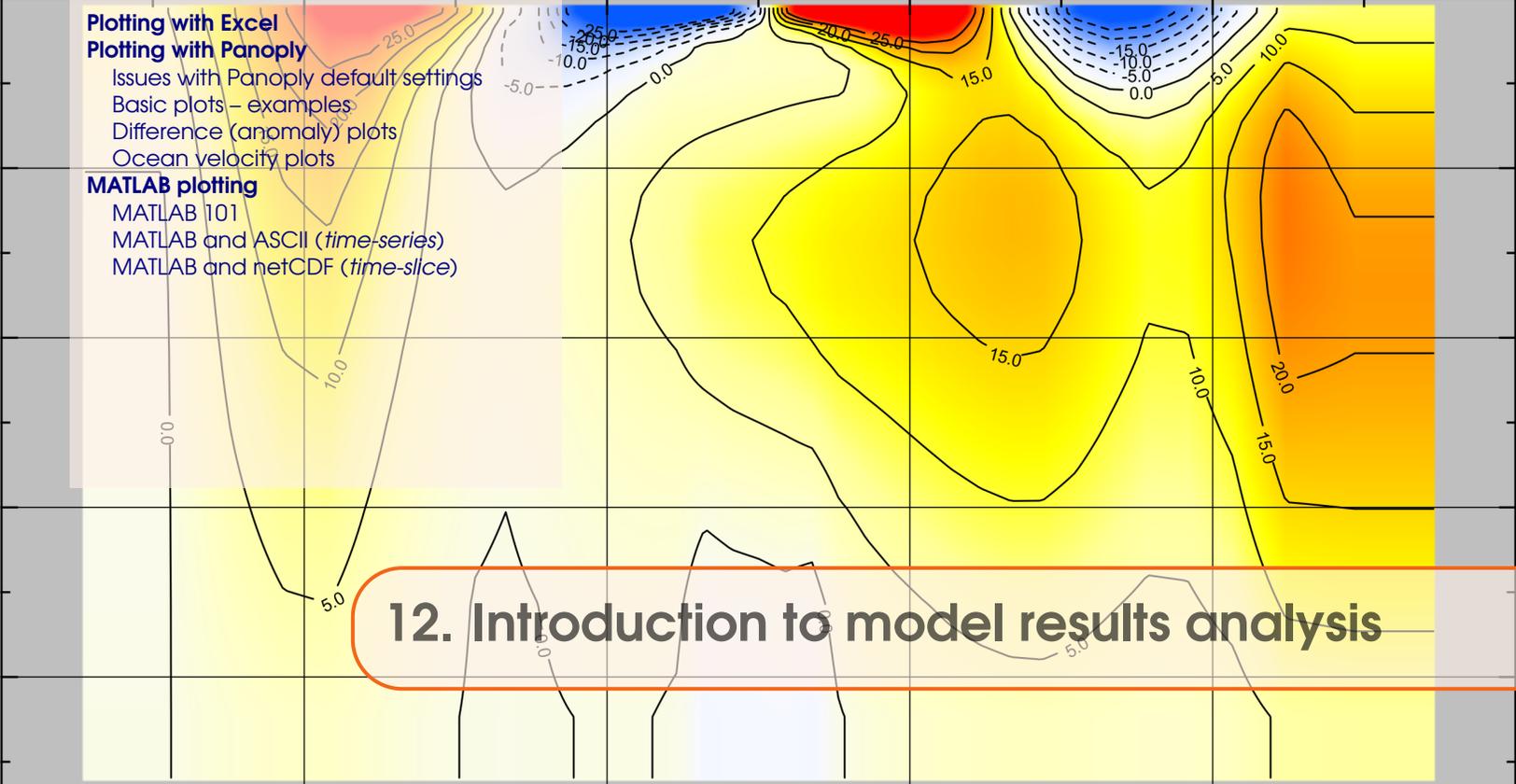
Ocean velocity plots

MATLAB plotting

MATLAB 101

MATLAB and ASCII (*time-series*)

MATLAB and netCDF (*time-slice*)



12. Introduction to model results analysis

The primary format for saving spatial (2- and 3-D) data is *netCDF* (network Common Data Form). More information on the netCDF format and the libraries necessary to compile the model can be found [here](#). The writing of netCDF follows roughly the **CF1.0 convention** (*NetCDF* Climate and Forecast (CF) Metadata Convention). The netCDF output is written for *BIOGEM* and *SEDGEM* separately and both modules have a flag that suppresses data file saving in ASCII format, with *netCDF* format being the default.

Under unix/linux, netCDF files can be interrogated with: `$ ncdump -h filename.nc` which will give you the header information of the file. The command is included in the netCDF library which has to be present to run the model anyway. It's useful to get the NCO software package helping to concatenate files or extract variables as shell command. A full list of available software to manipulate or graphically illustrate netCDF files can be found [here](#).

This Chapter covers how to visualize **cGENIE.muffin** output, with a particular emphasis on (netCDF) spatial data, via:

- **Panoply**

If you really really must insist on using Windozzzz, a 'recommended' viewer for netCDF is [Panoply](#) (see below). In fact, the can be run under linux and on the Mac (OS X) as well.

There is also [ncBrowse](#). Again, this will also run under LINUX and on the Mac (OS X).

- **MUTLAB**

You can also view *netCDF* files using **MUTLAB**, for which a number of plotting functions are provided. An advantage here is that the **MUTLAB** code can be hacked to produce much more powerful and bespoke analysis and plots.

12.1 Plotting with Excel

Just ... don't do it!¹

¹Obviously, there may be circumstances where it might be helpful to plot time-series output in **Excel**, although in practice, time-series output is much easier and faster to load in and plot with **MATLAB**.

12.2 Plotting with Panoply

In the following sub-sections are some pointers and examples of **Panoply** plotting.²

When you open the NetCDF file, you will be presented with a Datasets window (on the left hand side of the application window). This contains a list of all the variables available that you can display. You will find that the 'Long Name' description of the variable will be the most helpful to identify the one you want. Clicking (once) and highlighting an entry will display further information about that variable in the Variable window on the left hand side of the application window.

In a drop-down box at the bottom of the application window is an option for shortening the list of displayed variables in the netCDF file:

- Georeferenced variables – All the spatial (2 or 3D) variables.
- Plottable variables – As above, but now including axis definitions (which can be plotted as 1D lines).
- All variables – all of the variables.

Advisable, is to ensure that Georeferenced variables is selected.

To create a plot of a variable – simply double-click anywhere on the line containing that variable. A dialogue box will open with various options (Figure 12.1). For 3-D model output, you have the option of whether you want a 'Lat-Vert', 'Lon-Lat', or 'Lon-Vert' plot (for the 2-D fields, the only choice is 'Lon-Lat').

- 'Lat-Vert' plots – a common way of visualizing vertical distribution in the ocean. The default is for **Panoply** to display an average of all longitudes in a zonal mean. Un-ticking the Avebox in the Arraytab will enable you to specify a specific longitude section. Be aware that Panoply likes to plot the depth inverted by default ...
- 'Lon-Lat' plots – the classic 2D, top-down view of the ocean. There are multiple levels (depth layers) in the ocean of data that can be plotted, from the surface to the abyssal ocean.
- 'Lon-Vert' plots – (an uncommon option).

For all three: there may be multiple time-slices (i.e., you can plot data saved from different years).³

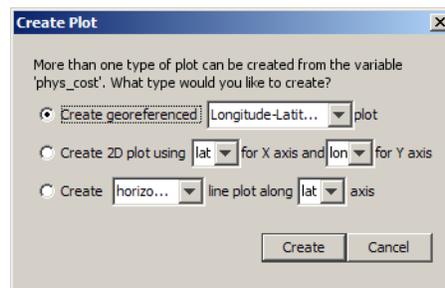


Figure 12.1: Panoply Create Plot dialogue window.

You can interpolate the data or not (often you may find that it is clearer not to interpolate the data but to leave it as 'blocky' colors corresponding to the resolution of the model), change the

²*WARNING* These instructions are strictly valid for older version of **Panoply** (ca. version 2.9.4), although some updates to the text have been made in light of version 4.6.2 ... so be aware that the plotting control buttons and options may have subtly changed in newer versions and the text no longer reflect the exact (current) options in **Panoply** ...

³Remember that the default, first time slice, will be the first once saved in the experiment. The last one saved and displayed, will reflect the end of your experiments.

scale and colors, overlay continental outline, change the projection, etc etc. Gray cells represent 'dry' grid points, i.e., continental or oceanic crust. To save plots in Panoply – from the file menu: File, then Save Image As ... and then select the location, filename, and graphics format.

12.2.1 Issues with Panoply default settings

The default settings in **Panoply**, i.e. those used when a plot is first created, can often mislead. In particular, note:

- Year (Arraytab)

The default is for the very 1st *time-slice* to be displayed rather than the experiment end. The first *time-slice* is numbered from 1 to however many total time-slices have been saved (displayed to the immediate right of the Year box), and it is this integer number that appears in the Year box – not the year of the data save. Instead, the mid-point year of the time-slice is displayed in a second box (labeled 'Year mid-point').

Different *time-slices* to be plotted can be selected by either clicking through the saved year count, or by selecting the save year mid-point from the drop-down list.
- Scale Range (Scaletab)

The color scale is auto-scaled so that the range always goes from the minimum to maximum displayed value. This can potentially mislead if save years and/or depth/latitude slices are scrolled through as the scale will be automatically adjusted to fit each plot in turn.

Confusion can also arise for fields with no variation, e.g. atmospheric trace gas concentrations or air temperature – the auto-scaled plot in these instances has a uniform color but with odd hatching as Panoply dutifully tries to achieve the impossible (creating a scale of multiple colors for a single value).
- Zonal averaging (Array tab)

Lat-Vert plots are displayed as a zonal mean by default. This is indicated by the tick in the Ave box (bottom RH corner). Un-ticking the Ave box releases the averaging with the first longitudinal value of the grid now displayed instead. Similar to how Panoply displays years – the longitudinal grid locations are counter from 1 to typically 36 (depending on the resolution of the ocean grid), with the longitudinal mid-point value in degrees East displayed to the right.

Different longitudinal sections to be plotted can be selected by either clicking through the grid point number count, or by selecting the longitudinal mid-point from the drop-down list.
- Scale bar tick marks (Array tab)

The tick labels on the color scale are displayed by default in the format: x.y . If the typical values of the variable are order e.g. 10^{-6} you will end up with value labels ranging from 0.0 to 0.0 ... This can be most easily resolved in one of two ways:

 - The format of the label can be changed by selecting a different option from the pull-down Tick Label Format box (default == %.1f). For instance, %.2e would give a display in the format x.xxEyy (or x.xxE-yy) or %.6f would give x.xxxxxx.
 - An alternative is to re-scale the values. This is done in the Scaling Factor box in which you set the scale factor in powers of 10. For example: setting -6in effect converts units of mol kg^{-6} to $\mu\text{mol kg}^{-6}$.

Simply be careful when opening a new plot that you are looking at what you **think you are looking at (or what you think you are looking at **is** what you are looking at).**

12.2.2 Basic plots – examples

12.2.3 Difference (anomaly) plots

It is possible to create an anomaly (difference) maps in Panoply which are essential when analyzing changes in a variable that may be small compared to the global variability in that variability. To do this:

- First, open the netCDF results file.
- Open the variable of interest, e.g., atm_temp (surface air temperature) in the 2D netCDF file.
- From the upper LH corner of the Dataset Browser window, from the drop-down menu, select the name of the plot you have just created (atm_temp in **field_biogem_2D** ...).
- From the upper LH corner of the Dataset Browser window, now click on the Combine Plot icon.

You now have a plot window that is displaying a difference map. By default, it is showing you the difference between two identical (in time) slices. The two different slices are labeled Array 1 (LH side) and Array 2 (RH side).

For instance, you can keep one array (Array 1) fixed to the initial (year 1 (centered on 0.5)) and vary the year in the second array (Array 2). Note that you can select in Panoply whether Array 1 - Array 2 is plotted, or Array 2 - Array 1, or various proportional or relative differences.

If switch off the auto-scaling feature (**Always fit to data**) you can center the scale so that no change is white, with positive deviations = red and negative = blue by clicking on Center on 0. This is something of a convention in the scientific literature.

The same variable in two different model experiments can also be opened up and analyzed combined:

- Start by opening up both required netCDF files.
- Open the variable of interest in one (either one) of the two 2D netCDF datasets.
- From the upper LH corner of the Dataset Browser window, from the drop-down menu, select the name of the plot you have just created.
- Now double-click in the variable in the 2nd netCDF dataset.

You now have a plot window that is displaying a difference map, but of the same variable between two different experiments, rather than two years of the same experiment.

12.2.4 Ocean velocity plots

By combining the two (horizontal) fields of ocean circulation, rather than a difference plot, Panoply can create a velocity plot. This is a great way of visualizing surface (and deeper) currents and circulation patterns. To do this:

- First, open the 3-D netCDF results file.
- Open either the phys_u ('ocean velocity - u') or phys_v ('ocean velocity - v') field and select a Lon-Lat plot.
- From the upper LH corner of the Dataset Browser window, from the drop-down menu, select the name of the plot you have just created.
- Now double-click on the other velocity variable (whichever of the u and v fields you did not open first).
- By default you get a difference map, which is pretty useless really. From the drop-down Plot menu box (which should be displaying 'Array 1 - Array 2' by default) select: **Vector Magnitude** (bottom of the list).

You now have a plot window that is displaying the ocean velocity field, with arrows indicating

the direction and speed (length of the arrow) together with an interpolated color background of the speed.

You can re-scale the velocity arrows to more clearly display the circulation pattern by altering the **Scale Length** value (**Contours & Vectors** tab). A value of 0.1 is a reasonable choice for surface currents. e.g. see Figure 12.2.

If you want to display deeper (in the ocean) current fields and/or different time-slices, take care that the depth level (/time-slice) in both LH and RH sides of the **Array(s)** panel must be changed to the same value. If displaying deeper current fields, then the velocity vectors will have to be further re-scaled (to a smaller value) in line with the lower velocities at depth compared to the surface.

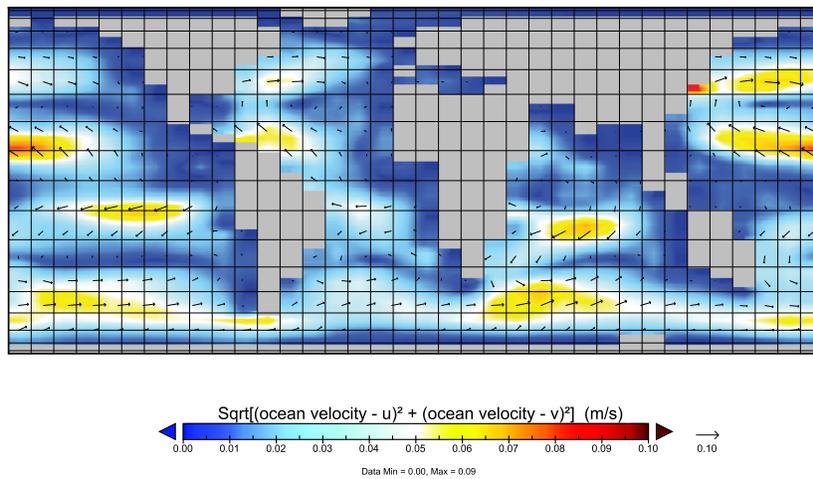


Figure 12.2: Example (modern) ocean surface velocity (current) map.

12.3 MATLAB plotting

12.3.1 MATLAB 101

If you need a tutorial on **MATLAB**, either as a refresher, or because you have not used the program before, work through the following sections of the **GEO111 course text**:

- All of Chapter 1 – that covers the very basics of using **MATLAB**, what variables are, including scales (e.g. single numbers), vectors (1D array), matrices (2D array) and higher order arrays, how to index and carry out basic manipulation of arrays, basic data loading and saving and plotting.
- Sections 2.1 and 2.2 of Chapter 2, which cover creating (script) programs – i.e. adding all your **MATLAB** commands to a file (a .m or 'm-file ') and running the file, and functions – programs (m-files) where one or more parameters might be passed into the function when it is called (e.g. at the command line), and potentially variables returned.
- In Chapter 3 – subsection 3.1.3 deals with reading in netCDF format data, which is the **cGENIE.muffin** format for spatial data. Section 3.2 deals with more advanced 2D plotting (also of direct relevance to **cGENIE.muffin** data processing and visualization using **MATLAB**).

12.3.2 MATLAB and ASCII (*time-series*)

The *time-series* (.res) output of **BIOGEM** are in a simple plain text (ASCII) format. These can be read in very easily in **MATLAB** using the load command. Note that a brief description of each column of data in the *time-series* files appears on the first first line of the file, and that prefixing this is a % symbol, that **MATLAB** ignores. Hence only the columns of data gets read in by default using load :) ⁴

For example:

```
co2=load('biogem_series_atm_p\{CO_{2}}\'.res', '-ascii');
plot(co2(:,1),1.0E6*co2(:,3));
```

loads in the atmospheric CO_2 time-series file (assigning it to the array variable co2), and then plots (as a line graph) the 3rd column (CO_2 concentration) vs. the first (time). Because concentrations are saved in units of atmospheres, a factor of 1.0E6 is applied to concert to μatm . Equivalently:

```
co2=load('biogem_series_atm_p\{CO_{2}}\'.res', '-ascii');
scatter(co2(:,1),1.0E6*co2(:,3));
```

12.3.3 MATLAB and netCDF (*time-slice*)

(See 3.1.3 in the **MATLAB programming text**.)

Basically – the only hard part, having opened the *netCDF* file is correctly deducing which dimension in an extracted data array is which of longitude, latitude, (sometimes depth), and time. Mostly this should be pretty obvious from inspecting the **MATLAB** Workspace window (assuming you have a column for Size selected to be displayed), or using the size command.⁵ Once you have done this, you can plot slices, scale data, average or otherwise process data, extract locations at specific locations, etc etc.

⁴There are also other **MATLAB** commands for reading in text data – refer to the **MATLAB programming text**.

⁵It also turns out that the order of dimensions for a variable read in by **MATLAB**, is the opposite of the order listed in the Variable window of **Panoply**.

As an example – in **MATLAB**, first either change directory to the biogem directory of a set of **cGENIE.muffin** results, or from where-ever you are, add a path to the location of the biogem directory. To open the 2D **BIOGEM** *netCDF* results file, type:

```
ncid = netcdf.open('fields_biogem_2d.nc', 'nowrite');
```

To extract a variable, you first need to find its ID from its name:

```
varid = netcdf.inqVarID(ncid, NAME);
```

In turn, you might need to use Panoply to display all the different variable names. Or, you can list all the variables and stuff in the netCDF file using `ncdisp`:

```
ncdisp('fields_biogem_2d.nc')
```

Having, by one means or another, identified the name of the variable you are interested in, you can recover its ID and then the data itself, for example, for `atm_temp` (surface air temperature):

```
varid = netcdf.inqVarID(ncid, 'atm_temp');
data = netcdf.getVar(ncid, varid);
```

In loading in the variable, end up with a multi-dimensional array – 2 spatial dimensions and if you have more than 1 *time-slice* of data saved, 1 temporal dimension (and if you loaded in the **BIOGEM** 3D *netCDF* file, you end up with a 4-dimensional array). **MATLAB** reports the size of the array in the Workspace window (depending on which column display options you have selected). In the example here, which took the experiment `EXAMPLE.worjh2.Caoeta2009.RCP6p0`, the array for atmospheric temperature is reported as $36 \times 36 \times 8$.⁶ so the last of the 8 time-slices would be accessed as:

```
data_last = data(:,:,8);
```

or

```
data_last = data(:,:,end);
```

For 3D file and e.g. many ocean variables, the array you obtain via `getVar` is 4D. Panoply reports this as having dimensions of (*time, zt, lat, lon*) (e.g. (time=13, zt=16, lat=36, lon=36)) but again, **MATLAB** reads it in the reverse order. For instance:

```
ncid = netcdf.open('fields_biogem_3d.nc', 'nowrite')
varid = netcdf.inqVarID(ncid, 'ocn_temp');
data = netcdf.getVar(ncid, varid);
```

If we then type:

```
>> size(data)
ans =
    36    36    16    13
```

⁶Although note that in **Panoply**, it is reported as `atm_temp(time=8, lat=36, lon=36)`.

we get an array orientated as $(lon, lat, zt, time)$.⁷ The final time-slice would hence then be accessed:

```
data_last = data(:,:,end);
```

with `data_last` now becoming a 3D array with dimensions (lon, lat, zt) .

There are three key things to remember at this point:

1. Firstly, the depth levels are read such that index 1 is the surface, and 16 is the deepest ocean depth level (in this case, otherwise it is 8).
2. For the lon-lat part – (lon, lat) equates to *rows* vs. *columns* in **MATLAB**, and hence if you were to plot e.g. the surface ocean slice:

```
imagesc(data_last(:,:,1));
```

you will end up with the plot on its side, with latitude along the x -axis and longitude on the y -axis.

You can exchange rows and columns in MATLAB with the transpose operator (see programming/MATLAB book):

```
imagesc(data_last(:,:,1)');
```

but ... this still leaves you with an up-side-down plot, because **MATLAB** reads from the first row down, whereas in latitude, you are expecting to read from -90 degrees up (towards the N pole). `flipud` accomplishes the final transformation:

```
imagesc(flipud(data_last(:,:,1)'));
```

3. The final complication is that `cGENIE.muffin netCDF` output uses a special value to represent an invalid or null number, e.g. for ocean temperature where the grid point is land (and an ocean temperature value would have no meaning). In the `netCDF` definition, Panoply is told what the special number is, hence it shows land in grey when plotting ocean variables. Panoply reports this as: `missing_value = 9.969209968386869E36`. There is no easy way (that I can see!) to get **MATLAB** to deal with this for you, so you need to search and replace this value, e.g.:

```
null=9.969209968386869E36;
data_last(find(data_last==null))=NaN;
```

which searches for this null value and replaces it with a `NaN` (so that now it can be simply plotted).

Once you are done accessing data, it is good practice to close the `netCDF` file after you are done with it:

```
netcdf.close(ncid);
```

The question then remains: what do you actually 'do' with it in **MATLAB**?

⁷ zt is the depth level in the ocean.

Plotting sections

For a quick look-see, `imagesc` (as per above) is handy.

For more advanced plotting (and for presentation) – refer to the [MATLAB programming text](#).

As per above, horizontal (lon-lat) fields can be extracted from 2D *netCDF* output by: `data(:,:,time)`

and from 3D by: `data(:,:,zt,time)`. Obviously, you could also extract lon-depth via: `data(:,lat,:,time)`

and lat-depth via: `data(lon, :, :, time)`.

Anomalies (with time) are extracted: `data(:,:,,time2)-data(:,:,,time2)`.

Typically, except a little trial-and-error in extracting the dimensions you want, and also in correcting the orientation of the matrix or resulting plot.

Calculating inventories

Often, the inventory (total mass or number of moles) of the ocean or atmosphere is useful to know, particularly as a function of time. For the ocean (or atmosphere) as a whole, the *time-series* output files report this (alongside the mean global concentration).

You can also calculate this with **MATLAB** from the *netCDF* output. Fields of ocean concentration are saved in the 3D output (and atmospheric concentrations in the 2D output). To convert concentration in the ocean, in units of mol kg^{-1} you'll need to know the mass of each ocean cell, in units of *kg*.

When saving using save option 9, or 99⁸, you save the 'physics' of the ocean model, which is actually mostly just the grid information, such as cell area, thickness, latitude and longitude edges and midpoints, depth edges and mid point. Also saved are the masses and volumes of the grid of cells. So to derive an array of cell tracer inventories from an array of concentrations, and the array of cell masses (variable `phys_ocn_M`), you'd write:

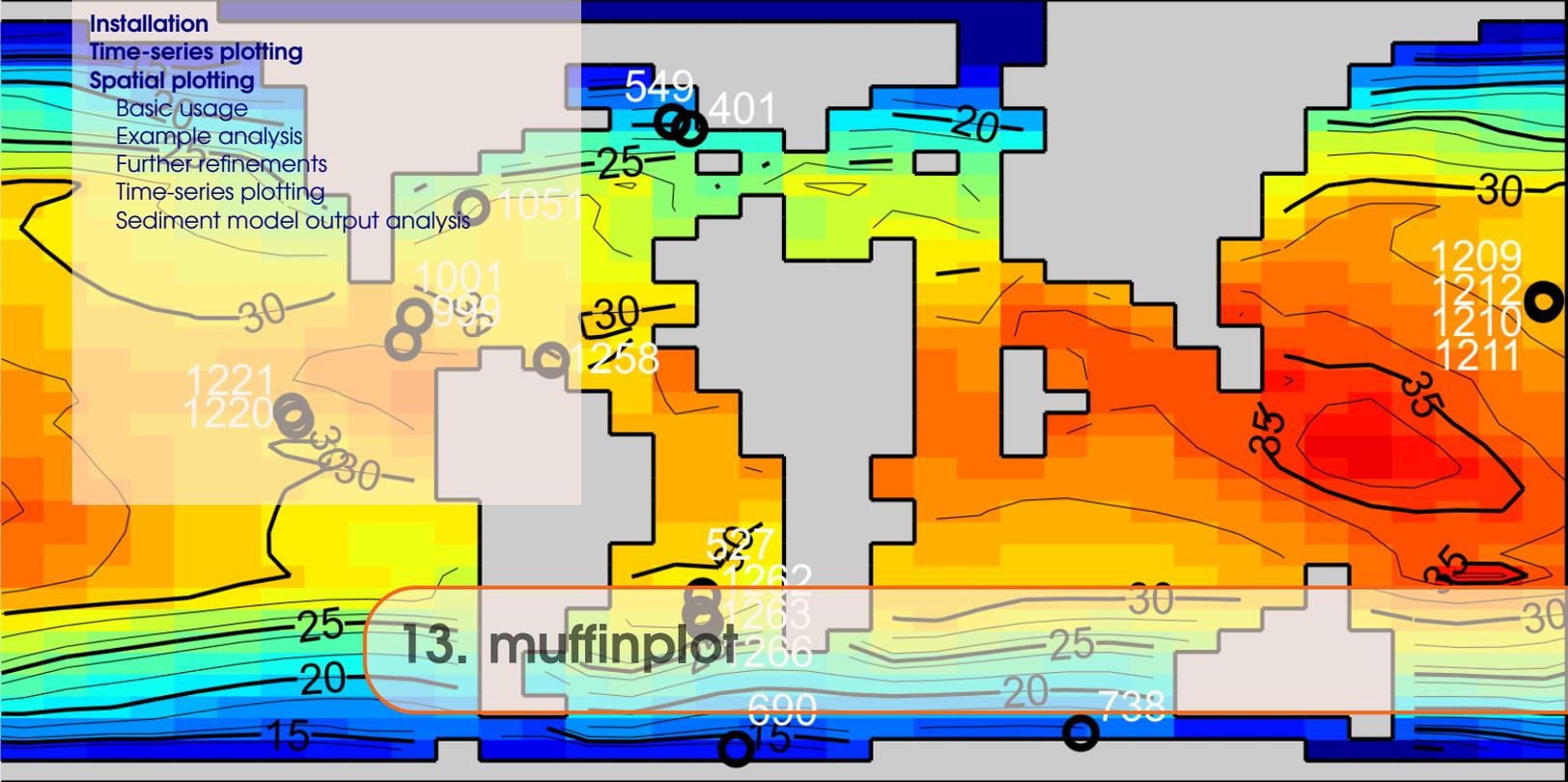
```
varid = netcdf.inqVarID(ncid,'ocn_temp');
data = netcdf.getVar(ncid,varid);
varid = netcdf.inqVarID(ncid,'phys_ocn_M');
mass = netcdf.getVar(ncid,varid);
inventory = data(:,:,,time).*mass(:,:,,time);
```

Before summing inventory to determine the global total inventory, you will, as before, have to deal with the null values (converting them to NaN) and then deal with the presence of NaNs in the array when summing ...

The advantage of doing the calculations in **MATLAB** (despite being provided with the global mean and inventory in the time-series files) is that you could calculate the inventory of a tracer (/substance) for just the ocean surface, or just a specific region of band of latitude. Or you could calculate the mean concentration for just a specific region of the ocean.⁹

⁸Parameter `bg_par_data_save_level` – see earlier.

⁹In calculating mean concentrations, you'll need to volume or mass weight the concentrations, and hence still need to use one of the physics variables.



The **muffinplot** suite of MATLAB functions provides a means of plotting a variety of output reproducibly (by means of saved parameter file) and with the potential for automation (i.e. automatically generating the same analysis for a large number of different experiments).

The functions comprising this software suite include:

- `plot_fields_biogem_2d`– lon-lat plots from the 2D **biogem** output.
- `plot_fields_biogem_3d_i`– lat-depth plots from the 3D **biogem** output.
- `plot_fields_biogem_3d_k`– lon-lat plots from the 3D **biogem** output.
- `plot_fields_ccd`– analysis of the 'CCD' (from both **biogem** 2D and **sedgem** 2D output).
- `plot_fields_sedgem_2d`– lon-lat plots from the 2D **sedgem** output.
- `plot_hisc_2d`– a generic color-coded histogram function.
- `plot_sedcore`– down-core plots from **sedgem** sedcore output.
- `plot_timeseries_biogem`– time-series plots from **biogem** time-series output.

At this current time, there is no facility for lon-depth plotting.

Most of the plots also perform additional functions (which can be generally disabled if not wanted), such as plotting and saving zonal or depth profiles, plotting difference maps, plotting and labelling data on maps and carrying out model-data fit statistics and plotting, extracting model values at data locations.

The following sections provide an overview and examples of such plotting and analysis.

13.1 Installation

muffinplot can be obtained from [github](#). If you do not have a **git** client on your computer (and hence can clone the repository locally), then simply download an archive of the code (from [clone or download](#) – pick Download ZIP).

When you unpack (or clone) **muffinplot**, you should see 3 directories – EXAMPLES, and MASKS, source, a series of .m files, and a single lonely .ps graphics file (colorscapes.ps). The .m files are split into filenames with or without the label SETTINGS – the ones without are code files (functions), and the ones with that string, contain parameter settings for plotting.

The plotting functions do not have to be run from the same directory that you are in – i.e. you can

```
>> addpath(PATH)
```

where PATH is the path from where you are, to the directory where **muffinplot** is installed. However, by default (the parameters can be changed if you wish), wherever you run the plotting function from, requires that you have a subdirectory called cgenie_output, where you will place the (complete) **cGENIE.muffin** experiment output directories (i.e. the contents of cgenie_output should look like the contents of cgenie_output on your cluster account¹). If you do any model-data analysis, by default, a directory DATA is also expected. Mask files reside in the MASKS subdirectory of the **muffinplot** installation, or in your current directory, or anywhere in the **MATLAB** path.

Results are saved to a subdirectory that by default is called PLOTS, and will be created for you if it does not already exist.

The plotting functions are run simply by typing their name and passing a list of parameters (comma-separated, with the complete list enclosed in parentheses). By default, the SETTINGS files need to be in the same directory as you are running the functions from, or in one of the **MATLAB** paths.

All the plotting functions provide some manner of 'help', that can be obtained by typing at the command line:

```
>> help FUNCTIONNAME
```

where FUNCTIONNAME is the function name (as per listed above).

13.2 Time-series plotting

The **muffinplot** function plot_timeseries_biogem.m provides a facility to plot **BIOGEM** *time-series* (.res) output.² You can use **MATLAB** help on the function name to detail the parameters that need to be passed (and examples).

The plot_timeseries_biogem plotting function plots a basic set of time-series variables by default. It then, enables a set up up to 3 additional variables to be plotted. It is also associated with a file of parameter values (plot_timeseries_SETTINGS.m by default) for fine-tuning plots.

The plotting function requires a list of parameters to be passed in the argument list, i.e.:

```
>> plot_timeseries_biogem(PAR1,PAR2,PAR3, ... PARn)
```

These are, in order:

¹Although you do not need to copy all the results over ... just the experiments that you wish to plot up.

²Obviously – there are lots of different and easy ways of plotting plain text output in the form of a simple column format.

1. PEXP1 – *string* → the (first) experiment name.
2. PEXP2 – *string* → is the name of the 2nd (optional) experiment. If no second experiment is selected, then a null string value must be passed, i.e., ''.
3. PTMIN – *real* → minimum plotted time (*x*-axis).
4. PTMAX – *real* → maximum plotted time (*x*-axis).
5. PDATA1 – *string* → time-series variable name for additional data to plot.
Omit the 'biogem_series_' and '.res' parts of the filename.
Leave blank, i.e., '', for no additional data panel.
6. PDATA1N – *integer* → the column number of the data in the time-series file.
7. PDATA2 – *string* → time-series variable name for additional data to plot.
8. PDATA2N – *integer* → the column number of the data in the time-series file.
9. PDATA3 – *string* → time-series variable name for additional data to plot.
10. PDATA3N – *integer* → the column number of the data in the time-series file.
11. POPT – *string* → The string for an alternative plotting parameter set.
If an empty (') value is passed as this parameter, then the default parameter set file is used.
12. PNAME – *string* → The string for an alternative filename.
If an empty value is passed as this parameter, then a filename is automatically generated.

13.3 Spatial plotting

Overview

4 of the **muffinplot** plotting functions provide spatial (2D) plotting capabilities:

- `plot_fields_biogem_2d`
Plot a 2-D field from: `fields_biogem_2d.nc`.
- `plot_fields_biogem_3d_i`
Plot a vertical-meridional (2-D) slice through the ocean (i.e., all cells have the same *i* (longitudinal) coordinate value) from: `fields_biogem_3d.nc`.
Options are provided for averaging longitudinally over a supplied mask, which may be the entire ocean and hence giving a global meridional cross-sectional mean, of a specific ocean basin, or may be a single cell 'wide' longitudinally and take a meandering path hence simulating an ocean transect. An option is also provided to overlay ocean circulation stream-functions.
- `plot_fields_biogem_3d_k.m`
Plot a horizontal slice through the ocean from: `fields_biogem_3d.nc`.
An option is provided for overlaying ocean circulation. Water column integrals can also be calculated and displayed.
- `plot_fields_sedgem_2d`
Plot a 2-D field from: `fields_sedgem_2d.nc`.

All 4 plotting functions can also overlay observed data and create difference (anomaly) maps – either between different experiments, time-slices, or variables, or between model and data and provide summary statistics regarding the difference.

Argument (parameter) list

All 4 plotting functions share exactly the same format of parameters³ passed in the argument list:

```
>> FUNCTIONNAME(PAR1,PAR2,PAR3, ... PARn)
```

i.e. take a (long!) list of parameters. These are (in order):

Firstly, a series of parameters for defining experiment, variable, and year:

1. PEXP1 – *string* – is the name of the 1st (main) experiment. A results directory with the same name must exist in the directory `cgenie_output`⁴.
2. PEXP2 – *string* – is the name of the 2nd (optional) experiment. If no second experiment is selected, then a null string value must be passed, i.e., ''.
3. PVAR1 – *string* – is the name of the 1st (main) variable. If no valid variable value is given, a list of valid variable names will be printed out.⁵
4. PVAR2 – *string* – is the name of the 2nd (optional) variable. If no second variable is selected, then a null string value must be passed, i.e., ''.
5. PT1 – *real* (or *integer*) – is the value of the 1st (main) time-slice. If no valid variable value is given, a list of valid variable names will be printed out.⁶
6. PT2 – *real* (or *integer*) – is the value of the 2nd (optional) time-slice. If no second time-slice is selected, then enter -1.⁷

Then there are 2 parameters for plotting sub-sets of the 2D or 3D data (essential for 3D data which cannot be usefully visualized in raw form):

1. PIK – *integer* – varies in its interpretation and is discussed below.
2. PMASK – *string* – is the name of an optional (2D) mask. A null string ('') must be passed if no mask is requested. A file with the same name (plus an extension `.dot`) must exist in the directory `MASKS`⁸. The interpretation of this parameter differs slightly between functions (below).

Next come options for plotting scale control:

1. PCSCALE – *real* (or *integer*) – is the scale factor for the plot. For example, to plot in micro molar (umol kg-1) units, enter; `1e-6`. The plot is auto-scaled if a value of zero (0.0) is entered.
2. PCMIN – *real* (or *integer*) – is the minimum scale value.
3. PCMAX – *real* (or *integer*) – is the maximum scale value.
4. PCN – *integer* – is the number of (contour) intervals between minimum and maximum scale values.

Finally, there are 3 parameters for: specifying discrete (observed) data to be plotted (and analyzed against model projections), for specifying the plotting parameter file to be used, and for substituting an alternative filename for all the output:

1. PDATA – *string* – is the filename containing the an overlay data set, which must be formatted as separated columns. The precise number and type of columns varies between different

³Parameters can be in for form of strings, in which case they must be given as a series of characters enclosed in inverted commas ''; as real numbers, e.g. `999.5` or `9.995E2`; or integers, e.g. `2`, `10`.

⁴Or alternative directory if the default file path settings have been changed.

⁵As a string, the value must be encased in inverted commas: ''.

⁶As **sedgem** does not save multiple and/or time-specific data, a dummy value (anything) is entered here.

⁷As **sedgem** does not save multiple and/or time-specific data, a dummy value (anything) is entered here.

⁸Or alternative directory if the default file path settings have been changed.

functions and also the plotting options chosen, and are hence discussed later. The full filename of this file must be given, including any extensions (e.g. `.dot`, `.txt`). This parameter must be passed as a *string*; leave blank, i.e., `''`, for no overlay data.

2. `POPT` – *string* – is the m-file filename (excluding the `.m` extension) containing the plotting options (`SETTINGS`). This parameter must be passed as a string; leave blank, i.e., `''`, in order to load the default file (`plot_fields_SETTINGS`).
3. `PNAME` – *string* – is the string for an alternative series of output filenames. This parameter must be passed as a string, e.g., `'experiment2'`. If an empty (i.e., `''`) value is passed to this parameter then the output filenames will be automatically generated.

The basic parameter list to all 4 plotting functions is hence:

```
>> FUNCTIONNAME(PEXP1,PEXP2,PVAR1,PVAR2,PT1,PT2,PIK,PMASK,PCSCALE,PCMIN,PCMAX,PCN,PDATA,POPT,PNAME);
```

Note that for `plot_fields_sedgem_2d` several of the parameters are redundant but must still be included (typically as zeros). This is in order to retain a common parameter list format between all the different plotting functions.

Function specific interpretation of PIK and PMASK

`PIK`, and to some extent, `PMASK`, have quite different interpretations depending on the particular plotting function used:

1. `plot_fields_biogem_2d`
 - (a) `PIK` – is the maximum depth (`k`) level that will be plotted, i.e. all depth levels deeper than `PIK` will be excluded. This is useful for plotting a variable only for the 'deep' ocean (rather than the ocean overlaying all ocean depths) for example. This value also provides an alternative way of creating a mask, and only values of `k` less than or equal to the passed value will be plotted.
 - (b) `PMASK` – is the name of an optional (2D) mask. A null string (`''`) must be passed if no mask is requested. (Shallow depths could also be excluded from the plot by means of a mask rather than setting `PIK`.)
2. `plot_fields_biogem_3d_k`
 - (a) `PIK` – the depth (`k`) level to be plotted. Note that the levels are numbered from a maximum value designating the surface, to 1 for the deepest ocean level. Typically, maximum values for the number of ocean levels are 8 (e.g. *Ridgwell et al.* [2007]) or 16 (e.g. *Cao et al.* [2009]).

Non ocean level `k` values have special meanings here:

- i. 0
 - A zero will result in a water column integral being plotted. With data, the model-data is carried out on the grid as a whole.
 - ii. -1
 - Will result in the benthic surface being plotted.
 - (b) `PMASK` – is the name of an optional (2D) mask. A null string (`''`) must be passed if no mask is requested.
3. `plot_fields_biogem_3d_i`
 - (a) `PIK` – the longitude-depth (`i`) slice through the ocean to be plotted.
 - Non longitude grid point `i` values have special meanings here:
 - i. 0
 - A zero will result in a zonal mean being plotted. With data, model-data comparison

is conducted at the specific data locations, rather than vs. a zonal mean model value.

ii. -1

I have forgotten what this does ...

(b) MASK – is the name of an optional (2D) mask. A null string (‘’) must be passed if no mask is requested.

For example: if the mask is of the entire ocean (mask_worbe2_ALL.dat), the result is a global meridional cross-sectional mean.

If the mask is just of a single basin such as the Atlantic (mask_worjh2_Atlantic.dat), the result is the Atlantic meridional cross-sectional mean.

Masks can also be constructed that are only a single cell wide longitudinally, but which take a meandering path following an ocean transect⁹.

The trivial usage would be to construct a mask consisting of a vertical line of 1s – the result is equivalent to setting an appropriate i value in PIK.

4. plot_fields_sedgem_2d.m is an exception as it does not (currently) use either parameter. PIK must be entered as 0 (any integer will do in fact), and PMASK as ‘’.

The mask itself (if PMASK contains a mask name) is a 2-D array of model grid points (on the **BIOGEM** grid) in the form of a simple ASCII file. A value of ‘1’ represents a vertical column of ocean cells to include, whereas a value ‘0’ will exclude all cells in the water column at that particular grid point. Examples of masks can be found in the MASKS subdirectory of **muffinplot**.

13.3.1 Basic usage

What follows are some basic and quasi random examples, just to illustrate a simple use of the three main plotting functions (1 function for the 2D netCDF output, and 2 functions for the 3D netCDF output).

1. Surface ocean temperature

Surface ocean temperature can be plotted in 2 ways – via the 2d plotting function (but only if the surface tracer properties fields have been saved, as these are optional), or via the 3d plotting function.

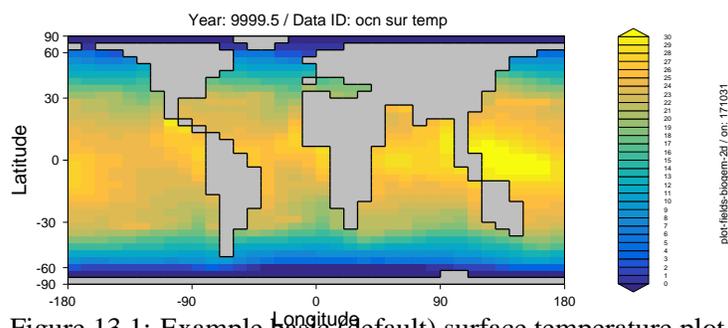


Figure 13.1: Example basic (default) surface temperature plot.

For example:

```
>> plot_fields_biogem_2d ...
('EXP1', '', 'ocn_sur_temp', '', 9999.5, -1, 16, '', 1.0, 0.0, 30.0, 30, '', '', 'example1a');
```

⁹e.g., as in: mask_worjh2_GEOSECS_WATL.dat

plots from the experiment EXP1, the variable `ocn_sur_temp` for time-slice 9999.5 (the mid-point time of the final year of a 10,000 year experiment). The color scale is from 0.0 to 30.0, with no re-scaling (1.0), and 30 color intervals in the scale. The default SETTINGS parameter file is used, and the default filename string replaced with `example1a`. The only other thing to note, is for parameter PIK, a value of 16 is set – corresponding to the ocean surface. See Figure 13.1.

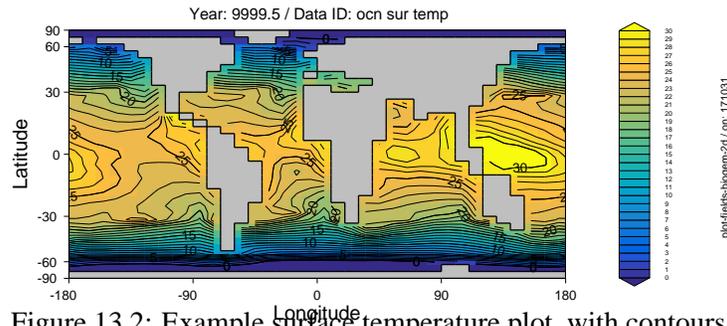


Figure 13.2: Example surface temperature plot, with contours.

To add contours, in the SETTINGS parameter file, adjust the following line:

```
contour_plot = 'y'; % ['y'] OVERLAY CONTOUR PLOT?
```

The results of this are shown in Figure 13.2. Refinements to the contouring can be done by changing the lines:

```
contour_mod = 1; % [ 1] NUMBER OF COLOR INTERVALS PER CONTOUR
contour_mod_label = 5; % [ 5] NUMBER OF LABELED CONTOURS PER CONTOUR
contour_label = 'y'; % ['y'] LABEL CONTOURS?
contour_dashneg = 'n'; % ['n'] PLOT NEGATIVE CONTOURS DASHED?
```

(these are the more commonly used refinements).

Alternatively, using 3d plotting:

```
>> plot_fields_biogem_3d_k ...
('EXP1', '', 'ocn_temp', '', 9999.5, -1, 16, '', 1.0, 0.0, 30.0, 30, '', '', 'example1c');
```

The only thing that changes here is the variable name – now `ocn_temp`, and the plot is identical to Figure 13.1.

2. Global zonal average temperature profile

To keep with ocean temperature, but now plot the global zonal mean (lat-depth) profile:

```
>> plot_fields_biogem_3d_i ...
('EXP1', '', 'ocn_temp', '', 9999.5, -1, 0, '', 1.0, 0.0, 30.0, 30, '', '', 'example2a');
```

where the only significant change compared to before, is setting a 0 for input parameter PIK. The results is shown in Figure 13.3.

3. Pacific dissolved oxygen profile

As per choosing ocean levels (k-values) in the lon-lat plotting, you can specify a specific longitude for creating a lat-depth section. e.g. Figure 13.4, created by¹⁰:

```
>> plot_fields_biogem_3d_i ...
('EXP1', '', 'ocn_O2', '', 9999.5, -1, 10, '', 1.0E-6, 0.0, 300.0, 30, '', '', 'example3a');
```

¹⁰Also turning on the contour plotting.

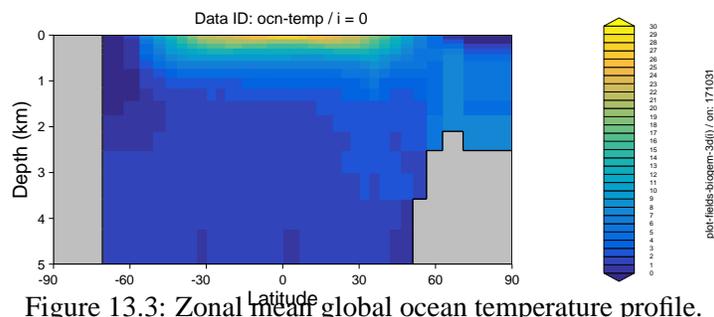


Figure 13.3: Zonal mean global ocean temperature profile.

is somewhere in the Pacific, along a line of longitude (whatever corresponds to $i=10$ on this GENIE grid ... I guess about 165W ...). Here, the variable to be plotted has also been changed – `ocn_O2`. Because of this the scale has also been changed – from 0 to $300 \mu\text{mol kg}^{-1}$, and because the *netCDF* variable units are mol kg^{-1} , a units scaling of $1.0\text{E}-6$ is specified for parameter `PCSCALE`.

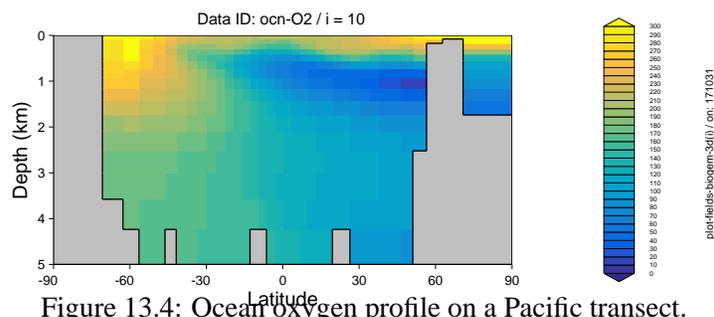


Figure 13.4: Ocean oxygen profile on a Pacific transect.

4. Atlantic zonal mean dissolved oxygen profile

So far, with the exception of plotting a gridded color field and a contoured field at the same time, all these examples can also be done in **Panoply**. One difference, is the ability in the **muffinplot** suite of **MATLAB** functions to apply masks – isolating geographical regions or even single points. In the `MASKS` directory, are a series of example ASCII mask files, mostly for the 2 (8- and 16-level ocean) modern published configurations of **cGENIE.muffin**. For instance, `mask_worjh2_AtlanticALL.dat` has all the grid points in the entire Atlantic basin assigned a value of 1, with 0 everywhere else. If we apply this first to the surface ocean dissolved oxygen field:

```
>> plot_fields_biogem_3d_k ...
('EXP1', '', 'ocn_O2', '', 9999.5, -1, 16, 'mask_worjh2_AtlanticALL.dat', 1.0E-6, 0.0, 300.0, 30,
... '', '', 'example4a');
```

we obtain Figure 13.5.

Now also applying a mask field to the zonal average:

```
>> plot_fields_biogem_3d_i ...
('EXP1', '', 'ocn_O2', '', 9999.5, -1, 0, 'mask_worjh2_AtlanticALL.dat', 1.0E-6, 0.0, 300.0, 30,
... '', '', 'example4b');
```

(Figure 13.6)

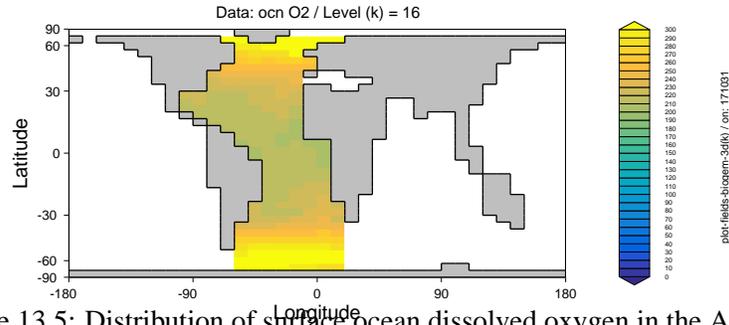


Figure 13.5: Distribution of surface ocean dissolved oxygen in the Atlantic.

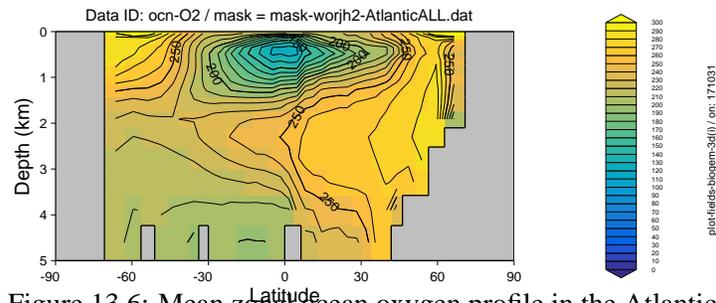


Figure 13.6: Mean zonal ocean oxygen profile in the Atlantic.

13.3.2 Example analysis

An set of `m-file` functions are provided that plot and analyse a basic set of experiments properties, with `make_analysis_ALL.m` given as a template for carrying out all the plotting. Obviously, the individual aggregate plotting functions can be edited, added to, or with unwanted or irrelevant plots, commented out or deleted – treat these all simply as templates for developing your own analysis strategy (as well as viewing the associated configuration files as illustrations of the function/use of some of the different further plotting options¹¹).

The aggregate plotting functions are as follows:

1. `fun_make_analysis_phys.m`

This encompasses a basic set of analyses of ocean circulation and climatology.

The script is written as a function, and requires two parameters to be passed as input:

- (a) The experiment name.
- (b) The (mid-point of the) year of the time-slice to plot.

In the example of an experiment called 'EXP1', and plotting the last annual time-slice (9999.5) from a 10,000 year model run, the function is hence called:

```
fun_make_analysis_phys('EXP1', 9999.5);
```

2. `fun_make_analysis_geo.m`

This encompasses a basic set of analyses of ocean (abiotic) geochemistry and ocean acidification related variables and metrics.

3. `fun_make_analysis_bio.m`

This encompasses a basic set of analyses of marine biological fluxes and biologically related

¹¹Also covered in a subsequent sub-sub-section.

properties.

4. `fun_make_analysis_eco.m`

This encompasses a basic set of analyses of marine ecological analysis

5. `make_analysis_ALL.m`

Aggregates the above functions.

Note that this particular m-file is a script file and has no inputs.

13.3.3 Further refinements

A number of additional options for exerting finer control over the plotting are provided as a block of parameters and (default) values in the m-file itself, in a section immediately after the commented help and change-log at the start of the m-file. Not all the options are relevant to all the plotting functions¹², but the full list (and then defaults in brackets []) is as follows:

1. `lon_min = -180; [-180] STARTING LONGITUDE FOR X-AXIS`
Sets the longitude of the left-hand edge of the plot.
2. `delta_lon = 90; [90] INCREMENT OF LONGITUDE ON X-AXIS`
Sets the longitude tick increment.
3. `contour_plot = 'n'; ['n'] OVERLAY CONTROL PLOT?`
Overlay line contours on the color block plot?
4. `contour_mod = 2; [2] NUMBER OF COLOR INTERVALS PER CONTOUR`
Number of color graduations per line contour.
5. `contour_mod_label = 4; [4] NUMBER OF LABELED CONTOURS PER CONTOUR`
Number of color graduations per labeled line contour.
6. `contour_label = 'y'; ['y'] LABEL CONTOURS?`
Label the line contours (frequency of labeled contours set by `contour_label`).
7. `contour_noneg = 'n'; ['n'] RESTRICT DATA PLOTTED TO > 0.0?`
Restrict the plotted values to non-negative? (Can be useful if slightly negative values exist as can occur during tracer transport associated with large concentration gradients.)
8. `plot_log10 = 'n'; ['n'] PLOT LOG10 OF THE DATA`
Plot data values as $\log_{10}(\text{value})$?
9. `contour_zero = 'y'; ['y'] PLOT ZERO CONTOUR`
Plot the zero contour?
10. `colorbar_old = 'n'; ['n'] PLOT 'OLD' COLORBAR`
Plot old style colorbar.
11. `data_offset = 0.0; [0.0] data offset (273.15 for K -> C)`
Introduce a data offset? This is useful for example for converting K to degrees C (removing the K value of 0 degrees C).
12. `data_ij = 'n'; ['n'] DATA as (i,j)?`
Overlay data in the form of (i,j) locations rather than longitude,latitude?
13. `data_ijk = 'n'; ['n'] DATA as (i,j,k)?`
Overlay data in the form of (i,j,k) locations rather than longitude, latitude, depth?
14. `data_ij_mean = 'n'; ['n'] average DATA by cell?`
Average overlay data per cGENIE grid cell rather than plotting raw locations.
15. `data_ijk_mean = 'n'; ['n'] average DATA by cell?`
Average overlay data per cGENIE grid cell rather than plotting raw locations.
16. `data_size = 25.0; [25.0] SIZE OF OVERLAY DATA POINTS`
Size of the overlay data points.

¹²See 'help' on a specific plotting function for details of the relevant options in the parameter block.

17. `data_anomaly = 'n'; ['n'] PLOT AS MODEL-DATA ANOMOLY ONLY?`
Plot data locations with the model-data anomaly rather than data value?
18. `data_only = 'n'; ['n'] PLOT ONLY DATA (no model values)?`
Plot only the overlay data locations (and not any model data)?
19. `data_site = 'n'; ['n'] PLOT DATA AS SITES (no data values)?`
Plot labeled site locations (no data value fill).
20. `plot_land = 'n'; ['n'] PLOT DATA OVER LAND?`
Plot data locations lying over land on the *c*GENIE grid (rather than screen out)?
21. `data_uv = 'n'; ['n'] overlay (u,v) velocity data?`
Overlay ocean current fields.
22. `data_uv_scale = 1.0; [1.0] scaling factor for vector length`
Scaling factor for velocity vectors.
23. `plot_opsi = ''; [''] PLOT OVERTURNING STREAMFUNCTION (basin)?`
Plot overturning streamfunction overlay?
24. `plot_opsi_min = -15; [-15]; plot_opsi_max = +15; [+15]; plot_opsi_dminor = 1; [1]; plot_opsi_dmajor = 5; [5]`
Controls on min, max and (major and minor) contour intervals.
25. `dscrsz = 0.60; [0.60] FRACTIONAL FIGURE WINDOW SIZE`
Adjustment factor of the fractional size (compared to the screen) of the figure window.

Further refinements: Examples

Examples:

1. To plot the positions (and labels) of data locations:

```
plot_fields_biogem_3d_k('cgenie_output', '120926.SPIN', '', 49999.5, -1, 'ocn_temp', '', '',
16, 1.0, 10.0, 40.0, 30, '', 'sites.dat')
```

 where the experiment name is 120926.SPIN, the mapped variable is `ocn_temp` (although no model field need be plotted – set by an option in the plotting function itself, and the file of data locations is `sites.dat`.

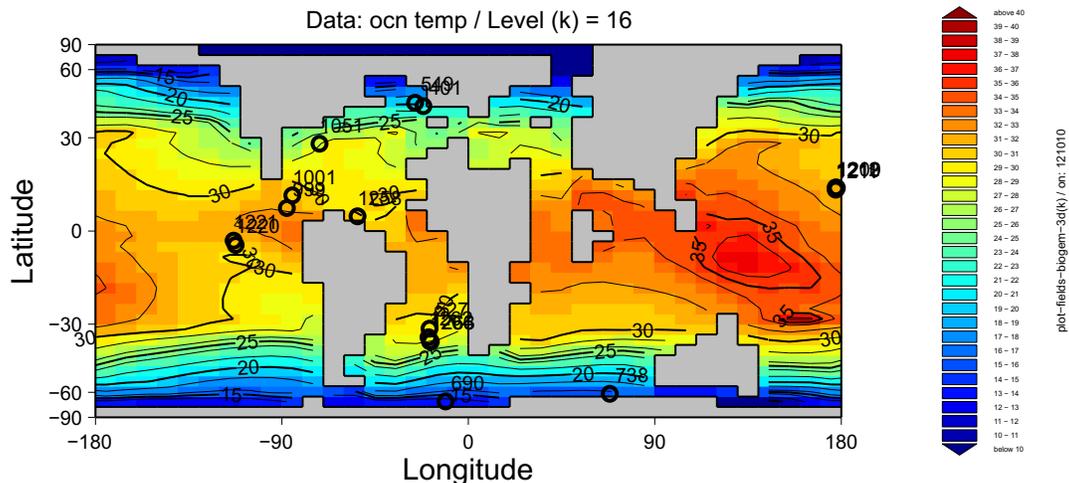


Figure 13.7: Paleocene-Eocene deep-sea sediment drill locations together with a contour-overlain map of surface temperature.

13.3.4 Time-series plotting

13.3.5 Sediment model output analysis

14. muffindata

14.1 Data re-gridding

14.2 Miscellaneous data processing

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Forcings questions



15. FAQ

aka: 'Has this dumb question been asked before?'

15.1 Installation related questions

Stack space

You may encounter issues with regards to the `ifort` Intel FORTRAN compiler (and maybe others), particularly when using SEDGEM because of the size of the arrays holding sediment information:

”The Intel® Fortran Compilers 8.0 or higher allocate more temporaries on the stack than previous Intel Fortran compilers. Temporaries include automatic arrays and array sub-sections corresponding to actual arguments. If the program is not afforded adequate stack space at runtime relative to the total size of the temporaries, the program will terminate with a segmentation fault.”

The (a?) solution is to increase the CPU stack space, Try:

```
$ ulimit -s unlimited
```

15.2 Cluster/queue questions

Do I have to submit experiments to the queue rather than running interactively?

Yes! Except for developing the model and debugging, testing new experimental designs, and forcing a re-compile. The number of instances of the model that can be run simultaneously interactively is limited by the number of processing cores on the head node. The more experiments that are run interactively, the slower everything will go. Additionally, if you even temporarily lose your Internet connection, an interactively-run experiment will die. The queue is there for your convenience, believe it or not ...

Can I leave all my experiment results on the cluster for ever?

No! Nothing is backed up on the cluster, and space is not infinite. So, periodically, transfer archived (.tar.gz) results off of the cluster and delete both the archive file and the results directory.

15.3 Help! My experiment has died ... why?

If, when using the `runmuffin.sh` shell script to run a **cgenie.muffin** experiment, it all goes horribly pear-shaped ...

1. The experiment dies absolutely immediately.
Check that the `runmuffin.sh` shell script has executable permissions. Also check that the directory you are trying to run the model from is the `genie-main` directory.
2. The experiment does not quite die immediately, but does not manage to stagger even as far as the line:

```
>> Here we go ...
```

before dropping dead. If so, there should be an error message telling you that a particular file or directory cannot be found. Check:

- All the files and directories you have specified exist.
- You have not omitted spaces where you should not have, nor added spaces where a `'_'` separator was required.
- You have not misspelt anything – a common cause of problems is in reading the number one (`'1'`) for the letter el (`'l'`), or *vice versa* in the computer font (`Courier`).

These first two sorts of pain and suffering are due to mis-configuration of the `runmuffin.sh` shell script. Also refer back to the overall sequence of configuring and running the **cGENIE.muffin** model shown in Figure 1.2.

15.3.1 Other sources of error

Other sources of error are due to the configuration of **cGENIE.muffin** (or more rarely, due to the model itself):

1. As **cGENIE.muffin** initializes, files may be reported as not being found. One possible cause of this is that `'~'` may not necessarily get expanded into the path of your home directory (e.g., `~/home/mushroom`). In this situation, `'~'` can simply be replaced with `'$HOME'`. Note that as well as making this substitution at the command line, the *user-config* file may also contain instances of `'~'` (such as in specifying particular *forcings*).
2. A missing/not found error can also arise with some compilers if one of the various ASCII input files to **BIOGEM** (or **SEDGEM**) does not have a blank line at the bottom (some vague quirk of the unformatted read used in the **FORTRAN** code). Check: the *user-config* file, and also any boundary condition files being requested.
3. Further trouble can occasionally arise when using **Windoz** and editing files (e.g., the *user-config* file) and it is possible to corrupt the format of the file. For what file(s) you have edited, use the command `dos2unix` to strip off **Windoz** formatting characters (which are invariably invisible in most editors). The syntax for this (or see the **linux** 'man' pages, or even Google it) is `$ dos2unix FILENAME`.
4. If the model starts running, but dies with a reported failure to solve the aqueous carbonate system, it may be that you need to force a re-compile (`$ make cleanall`). Running **cGENIE.muffin** with array dimensions which do not match the number of tracers selected is a common cause of failures to solve the aqueous carbonate system, as often calcium ion or other tracer concentrations become corrupted and get assigned nutty and all but impossible values.

Also, if a *re-start* is used which was generated with a different land-sea mask (and *base-config*) to the current experiment and associated *base-config*. Try running without the *re-start* and see if that solves it (identifies the source of the problem).

15.3.2 Meaning of specific error messages

'ERROR: path integral around island too long'

Such an error is possible when developing new or modifying existing continental configurations (and associated 'island' and 'path' definition files), but not in normal running of the model. First try a `make cleanall` and then try re-running. If the problem persists, it is possible that a key configuration file has accidentally/somewhat been changed. To check for this – do a `make cleanall`, and then from the `cgenie.muffin` directory:

```
svn status -u
```

Any file that you have modified is labeled with an `m`. Any new files on the server that you don't have will have a `*`. Files with a `?` are files that exist locally and are not on SVN (and can be ignored). If there is a file with an `m` that should not have been modified:

```
svn revert FILENAME
```

will re-set the file `FILENAME` (also include the relative path) it to the current SVN version status.

'ERROR MESSAGE: Particulate tracer CaCO3 ...'

I have been told 'ERROR MESSAGE: Particulate tracer CaCO3 does does not have the corresponding ocean tracer Ca selected' – is this a problem ... ?

No! You are simply being reminded that you have calcium carbon (CaCO_3) selected as a particulate tracer in the model, but although when it dissolves it releases Ca^{2+} (and removes Ca^{2+} when CaCO_3 is precipitated), you do not have Ca^{2+} selected as an explicit dissolved tracer in the ocean. This is not a problem as by far the most important effect on the carbon cycle of adding/subtracting Ca^{2+} is a change in alkalinity, which is implicitly account for. Only on **very** long time-scales, or in deep-time situations when the $\text{Ca}^{2+}/\text{Mg}^{2+}$ ratio was very different form today, might you need to select Ca^{2+} (and Mg^{2+}) as an ocean tracer.

15.4 General running and configuring experiments questions

When does the model need to be recompiled?

cGENIE.muffin will need to recompile in the following situations:

- You have just carried out one of the **cGENIE.muffin** tests, e.g., `make test` or `make testbiogem`.
- You have changed the dimension of the climate model grid (which also means an automatic change in the biogeochemistry modules), either horizontally (e.g., going from 36×36 to 18×18) or vertically (e.g., going from 8 levels in the ocean to 16).
- You have changed the number of selected ocean biogeochemical tracers in the *base-config* and hence changed the value of:

```
GOLDSTEINNTRACSOPTS='$(DEFINE)GOLDSTEINNTRACS=2'
```

(The latter two involve a change in compiled array dimension.)

In all three situations, the *base-config* is being changed (or should be¹). In running at the command line (i.e. interactively), the `runmuffin.sh` script detects the change in *base-config*, and automatically forces a re-compile for you. However, the compute nodes of the cluster do not have access to the **FORTRAN** compiler. As a sad and unfortunate consequence, submitted jobs cannot recompile modules and all science modules must be already compiled when a job is submitted.²

To recompile (and re link) the science modules – first, start an interactive run of the experiment you want to conduct. This will ensure that it is correctly compiled. This also serves as a visual check that you have requested a *user-config*, restart, etc that actually exists. Start the run for the length of time you intend to use when submitting the experiment as a job to the queue, but kill it (keyboard command: `Ctrl-C`) once it is compiled and you are happy that it is running OK (say, after 10 years). You can now be reasonably confident that the experiment is safe to submit the job to the cluster (and all files and inputs are as they should be).

If you have multiple experiments, all with the same resolution and number of tracers, you DO NOT need to re-run interactively or attempt to recompile. Also, you can add 'modules' and not recompile. i.e., you can interactively run an ocean -only carbon cycle. And then submit it. And then submit an experiment using **SEDGEM** as well. (Because when the model is compiled, ALL sciences modules are compiled, meaning that all there is to do is just link them, which does not require the (ifort) **FORTRAN** compiler.)

Refer to Figure 1.2 for the sequence of steps associated with configuring and running model experiments.

In the naming of different *forcing* specifications: what does 'yyyyz' mean?

A. The naming convention for *forcings* is that the (sub)directory name starts with the code for the continental configuration, if the *forcing* is tied to a specific continental configuration. For example: *forcings* with the string 'FeMahowald2006' relate to the prescription of a dust (Fe flux) field re-gridded from Mahowald *et al.* [2006]. When this has been re-gridded to the worjh2 continental configuration, worjh2 appears at the start of the name. If the *forcing* is independent of a specific continental configuration, such as restoring atmospheric CO_2 to a prescribed value (uniformly throughout the atmosphere), the string is 'yyyyz', as in e.g.: `pyyyyz_RpCO2_Rp13CO2`.

¹One could edit a *base-config* and re-ruin, but it is better to create a new *base-config* file if editing any of the settings, particularly those affecting array dimensions

²It is OK to change the flavor of GENIE as linking is done by the C compiler.

Can I make the model run faster?

sign You speed freak. Is this all you care about? What about the quality of the simulation - does that mean absolutely nothing to you? Oh well ... There is a bunch of stuff that slows **cGENIE.muffin** down that may not be absolutely essential to a particular model experiment. These include:

- The number of tracers - if you don't need 'em, then don't select 'em! Selected tracers are automatically passed to **GOLDSTEIN** and advected/convected/diffused with ocean circulation. Similarly, **BIOGEM** does a whole bunch of stuff with tracers, particularly those which can be biologically transformed. All this is numerically wasteful if you aren't interested in them. Equally importantly, the more tracers you have selected the more careful you have to be in configuring the model. Superfluous tracers therefore cost more configuration time and/or increase the change of a model crash.
- *Tracer auditing* - the continuous updating and checking global tracer inventories to ensure that there is no spurious loss or gain of any tracer (i.e., a bug) has computational overheads associated with it. Whether this checking is carried out or not is set by the value of the flag `bg_ctrl_audit`³.
- *Time-series* results saving. Model tracer (plus some physical) properties are brought continuously averaged in constructing *time-series* results files. Cutting down on *time-series* that you don't need will help minimize model run-time. The various categories of time-series that will be saved are specified by a series of namelist parameter flags. However, within each category (such as ocn tracers - `bg_ctrl_data_save_sig_ocn`) all properties will be saved - you are not given the option to save a defined sub-set (for example, *DIC* and *PO₄* in the ocean but not *ALK*).
- Time-slice results saving. If you have relatively few requested time-slices over the course of the model integration then this is unlikely to significantly impact the overall run-time (even will all possible data category save namelist flags set to `.true.`). However, note that if you have accidentally triggered the default time-slice saving interval (by having no data items in the time-slice specification file (`bg_par_infile_slice_name`)) you may end up with the model running about as fast as a 2-legged dog super-glued to a 10-tonne biscuit.
Note that time-series saving of data that is a 2-D average, such as atmospheric composition at the ocean-atmosphere interface, sediment composition at the ocean-sediment interface, or just ocean surface conditions, is less numerically demanding than mean values that have to be derived from a 3-D data field.
- Alter the degree of synchronicity between climate and biogeochemistry (see HOW-TO guide).

As a very rough guide, the impact on total run-time of making various changes to the model configuration are listed as follows. Numbers are given as a percentage increase in total model run-time (using the `/usr/bin/time` linux command). Tracers selected in the ocean are *DIC*, *ALK*, *PO₄*, *O₂*, *DOM_C*, *DOM_P*, *DOM_O₂*, as well as ¹³C isotopic components (*DIC_13C* and *DOM_C_13C*) (+ T and S). The corresponding tracers are present in the atmosphere and as particulates. The model is run for 10 years as a new run (i.e., not loading in a restart file):

- ADD auditing ⇒ +15%
- ADD time-slice saving ⇒ +20%⁴

³It is `.false.` by default.

⁴Because only a 10 year integration has been carried out with a time-slice saved at 10 years, the computational cost of

- ADD time-series saving $\Rightarrow +15\%$
- REMOVE ^{13}C isotopic species (= DIC and DOC ocean tracers) $\Rightarrow -10\%$ ⁵

You can also run at lower resolution. The basic configuration for a faster 'lego box' **cGENIE.muffin** configuration consists of a 18×18 model grid and an 8 level ocean. The continents are in a zonally-averaged configuration and there is no topography in the oceans.

The model is accelerated by:

1. it's low resolution
2. taking 48 instead of 96 ocean time-steps per year in the ocean
3. **BIOGEM** being only being updated every 4 rather than every 2 ocean time-steps.

In this configuration 100 years take about 40 seconds, 10 kyr would just take over an hour, and 100 kyr could be run overnight!

time-slice saving is disproportionately high as displayed. With a longer integration, the relative cost of saving a time-slice will fall. In contrast, the computational cost as a fraction of total run-time of time-series saving and auditing is likely to remain the same.

⁵The speed gained by removing two tracers is not proportional to the fractional decrease in number of tracers (in this example reducing from 11 to 9 the number of tracers in the ocean gives only a ca. 10% improvement in overall speed).

15.5 Climate-y questions

Can I disable climate feedback with CO_2 ?

Yes, for instance when you might want to compare the fate of CO_2 released to the atmosphere with climate (and ocean circulation and temperatures) not responding, vs. CO_2 in the atmosphere also driving changes in climate (and hence affecting the pathways and transformations of CO_2 , particularly in the ocean).

To specify no climate feedback, add to the *user-config* file:

```
# set no CO2 climate feedback
ea_36=n
```

as well as a radiative forcing scaling:

```
# scaling for atmospheric CO2 radiative forcing, relative to 278 ppm
ea_radfor_scl_co2=1.0
```

(a value of 1.0 giving no change in climate relative to the default).

How do I change the orbital configuration of cGENIE.muffin?

There is a complicated, time-dependent way of specifying how the orbital configuration changes with time. But sod this. There is also a simple⁶ way ... assuming that you only require a fixed (but different from modern) orbital configuration (and are running to steady-state).

The parameters:

- ea_par_orbit_osce=0.0167
- ea_par_orbit_oscsob=0.397789
- ea_par_orbit_oscgam=102.92

specify, respectively:

- eccentricity
- sine of obliquity
- longitude of perihelion

To specify a last glacial maximum like configuration, you would add (to the *user-config*):

- ea_par_orbit_osce=0.018994
- ea_par_orbit_oscsob=0.389911
- ea_par_orbit_oscgam=114.42

For comparison, the mid-Holocene (6 ka) parameter set is:

- ea_par_orbit_osce=0.018682
- ea_par_orbit_oscsob=0.408410
- ea_par_orbit_oscgam=0.87

(Taken from: [PMIP2](#) experiment boundary conditions.)

Can I do solar geoengineering (SRM) experiments?

No! Because you might destroy the planet.

No wait ... in the model (world) ... yes! But because of the absence of a dynamical atmosphere, options here are limited. However, modification of the solar constant, *a-la* 'giant mirrors in space' is possible. Sea-ice (surface) albedo can also be adjusted.

⁶.)

15.6 Ocean biogeochemistry questions

Can solubility related changes be separated from stratification and circulation changes?

With **BIOGEM** coupled to the climate model core of **cGENIE.muffin**⁷, a change in atmospheric CO_2 will induce a change in SSTs, which in turn affect the carbon cycle and feedback on CO_2 via changes in solubility and via changes in circulation (stratification) and thus biological productivity. There are times when it is helpful to separate out solubility related changes from circulation related changes. This equally applies to dissolved O_2 and CO_2 . The problem is that you need a change in climate and surface temperatures in the climate model in order to induce a change in circulation.

There is a way of having an altered climate and circulation, which then affects the marine carbon cycle, yet specify the SSTs that are actually seen by **BIOGEM** (and thus used in solubility calculations).

First of all, control the radiative forcing of climate internally in the **EMBM** rather than externally by the atmospheric CO_2 concentration calculated by **ATCHEM**. Turn off explicit CO_2 forcing of climate by setting: `ea_36='n'`. The namelist parameter `ea_20` will then dictate the EMBM radiative forcing: a value of 1.0 (default) gives no change in radiative forcing ($CO_2 = 278$ ppm), a value of 2.0 corresponds to the effect of doubling CO_2 , $\times 4 CO_2$, etc. Altering the value of `ea_20` thus lets you control climate (and circulation) without having to adjust CO_2 and the carbon cycle.

Next, SSTs in **BIOGEM** can be specified independently of the climate model. You achieve this by setting up a restoring forcing of ocean temperatures at the surface. Note that by default, prescribing SSTs (or SSSs) in **BIOGEM** does not propagate through to the climate model which does its own independent climate thing based on the value of `ea_20`. This allows you to retain the surface temperatures and thus solubility associated with a $\times 1 CO_2$ World, but have a warmer more stratified ocean (appropriate for a much warmer World).

What actually happens is that **BIOGEM** receives both the altered circulation field and the altered SSTs due to $\times 4 CO_2$, but sets its own SSTs internally rather than use those calculated by the climate model. Setting up the SST restoring is in principal just like for **PO4**. The values for the SST field you can simply copy and paste out of a prior $\times 1 CO_2$ experiment.

The converse experiment, is to have circulation and biological productivity not change, but explore the effect of changes in SST-driven solubility. i.e., to separate the solubility pump from circulation change effects on glacial CO_2 .

What is 'tracer auditing' – should I have it switched on?

When developing a new model parameterization, it is of fundamental importance that careful track is kept of the total tracer inventory of the system in the face of internal mass transfer and any inputs (e.g., prescribed restoring or flux boundary conditions) or outputs (e.g., sedimentation). No spurious gain or loss of tracer mass must occur as a result of bugs introduced to the code. The tracer inventories of the ocean can be periodically calculated and compared to that predicted to have occurred on the basis of any net input (or output) occurring in the intervening time to help catch bugs. The simplest implementation would be an audit carried out at system start-up (before any transformation of tracer mass has taken place), and at the very end (after the last transformation of the tracer fields). However, integrating over over an extended time period can lead to the excessive accumulation of numerical (truncation) errors. Instead, the audits are carried out periodically during the model run. The periodicity of tracer auditing follows the times specified for time-series data saving (i.e., at time listed in the file specified by `bg_par_infile_sig_name`).

⁷Namelist: `ea_36='y'`

The entire audit procedure is as follows:

1. First, an initial inventory is calculated, achieved by summing the product of the concentration of each (selected) tracer with the mass of each cell, across all wet cells.
2. During the model run, the net spatially- and time-integrated transfer of tracer mass arising from all transfers across the external reservoir boundaries is calculated.
3. At a periodic pre-defined time, the inventories are re-calculated. The difference between old and new inventories should be equal to the integrated net flux. If the relative difference between re-calculated inventory and estimated (on the basis of net flux) differs by more than a predefined threshold then an error message is raised (and the model halted if requested)
4. The integrated net flux variable is re-set to zero and steps (2-4) repeated.

In short – if you are not modifying the code then you can take it on trust(!) that the model distribution is free of (major) bugs and that spurious gain or loss of tracers does not occur. If you don't trust me ... then switch the auditing feature on.

Auditing is inactivated by default. To activate it:

```
bg_ctrl_audit = .true.
```

To adjust the threshold (relative) tolerance⁸:

```
bg_par_misc_audit_relerr = value
```

To halt the model⁹ if it fails the tracer drift tolerance:

```
bg_ctrl_audit_fatal = .true.
```

A secondary benefit of tracer auditing when running the model interactively, is that it reports back to you the maximum and minimum value of all the tracers (and locations of where this occurs), as follows:

```
>>> SAVING BIOGEM TIME-SERIES @ year 0.50 278.069 -6.501 16.522 3.843 18.543 ...
temp / min = 0.2713E+03 (18,36, 8) / max = 0.3030E+03 ( 4,18, 8)
sal / min = 0.3337E+02 (10,35, 8) / max = 0.3891E+02 (30,29, 8)
DIC / min = 0.1878E-02 (35,24, 8) / max = 0.2581E-02 (33,21, 1)
DIC_13C / min = -.4225E+00 ( 3,16, 3) / max = 0.2792E+01 (25,13, 8)
DIC_14C / min = -.1779E+03 (33,21, 1) / max = 0.2197E+02 (30,29, 8)
PO4 / min = 0.7071E-07 (29,28, 8) / max = 0.3806E-05 ( 3,16, 3)
O2 / min = -.4521E-04 (27,30, 5) / max = 0.3363E-03 (24,35, 8)
ALK / min = 0.2212E-02 (10,35, 8) / max = 0.2724E-02 (33,21, 1)
DOM_C / min = -.4159E-05 (21,34, 3) / max = 0.1517E-04 (32,25, 8)
DOM_C_13C / min = -.1000E+20 ( 1, 3, 2) / max = 0.5817E+01 (29,36, 8)
DOM_C_14C / min = -.1000E+20 ( 1, 3, 2) / max = 0.2236E+04 (29,36, 8)
DOM_P / min = -.3924E-07 (21,34, 3) / max = 0.1431E-06 (32,25, 8)
Ca / min = 0.9769E-02 (10,35, 8) / max = 0.1136E-01 (30,29, 8)
CFC11 / min = 0.0000E+00 ( 1, 3, 2) / max = 0.0000E+00 ( 1, 3, 2)
CFC12 / min = 0.0000E+00 ( 1, 3, 2) / max = 0.0000E+00 ( 1, 3, 2)
Mg / min = 0.5050E-01 (10,35, 8) / max = 0.5888E-01 (30,29, 8)
```

How do I do an ocean CO₂ injection experiment?

There is a hard way (but maximum flexibility), a less hard way, ... and an easy way. To cut the shit – what follows is the easy way!

First, you want to use the updated tracer forcing format:

```
bg_ctrl_force_oldformat=.false.
```

⁸By default, this is set to 1.0E-08.

⁹By default the model will continue running, even if there is an apparent spurious drift in tracer inventories occurring.

Put this line in the *user-config* file if it is not already there, perhaps under FORCINGS section.

You will need a *forcing* template for the CO_2 injection – `pyyyyz_FCO2_UNIFORM`. This is provided on mygenie.seao2.org. Download and unpack (`tar xzf pyyyyz_FCO2_UNIFORM.tar.gz`) from the directory: `~/genie_forcings`. As it stands, this is configured to stuff 1 PgC yr-1 of CO_2 into the ocean over the course of one year. The location of the CO_2 injection is some random default place that probably does not exist, which is not very good. So, you need to specify your ocean location. For this, add the following lines to a *user config* file:

```
bg_par_force_point_i=22
bg_par_force_point_j=33
bg_par_force_point_k=5
```

which corresponds to a cell in the N. Atlantic ($i,j = 22,33$) at an intermediate depth ($k=5$).

The i,j,k coordinates are counted from left-to-right with longitude: i , from bottom to top with latitude: j , and from top to bottom with depth for ocean level, k . The land-sea mask and maximum depth (lowest k integer) you are allowed can be got from the BIOGEM 2D netCDF, variable `grid_level`. This is a map of the 'k' values. >90 means land, for the 8-level ocean the ocean depths will be between 1 and 8. 8 being the surface. So the map is of the depth of the ocean and thus lowest k value you are allowed to use.

By default, using the CO_2 injection forcing template you will get 1 PgC emitted to the ocean, in the location you specify. You can scale the amount of carbon up via the namelist parameter:

```
bg_par_ocn_force_scale_val_3=xxx
```

where `xxx` is the multiple of 1 PgC you want to inject. NOT your favorite movie viewer rating. e.g., 100 PgC:

```
bg_par_ocn_force_scale_val_3=100.0
```

Note that 100.0 PgC is quite a lot of carbon to be injecting into a single location (cell) in the ocean model! By default, the time-scale of injection is set as 1 year. To increase the time over which the CO_2 injection takes place use the namelist parameter `bg_par_ocn_force_scale_time_3`, which simply scales the time interval. i.e.,

```
bg_par_ocn_force_scale_time_3=10.0
```

causes the CO_2 injection to take place over 10 years. But since the flux is in units of PgC per year, you will get 1000.0 PgC carbon total (10 years x 100 PgC yr-1). So a combination of both namelist scaling parameters (both flux scaling, and interval scaling) will be needed for the required total CO_2 injection.

Note that the integer at the end of the namelist parameter name corresponds to the index of the ocean tracer. 3 is DIC. 12 would allow you to inject alkalinity into the ocean (but the you would need to create additional forcing specification files).

The slightly harder way involves entering in the i,j,k location explicitly in the forcing configuration file `configure_forcings_ocn.dat`. Altering the magnitude and/or duration of the flux release requires editing `biogem_force_flux_ocn_DIC_sig.dat`.

The hardest way requires that two 3D fields explicitly specifying the spatial nature of the forcing flux are created and modified.

For these alternative options – see earlier section on tracer forcings (Section 4).

Can I do carbon dioxide removal (CDR) experiments?

Yes! See HOW-TO.

15.7 Questions of long-term carbon cycling and stuff

In GEMlite, does the adaptive step size control work with fixed/prescribed pCO_2 ?

If pCO_2 is fixed/restored, the answer is 'no' (ish). Or rather: you'll often get little difference compared to simply fixing the ratio of accelerated to non-accelerated time-steps. However, you will still get the advantage of adapting time-stepping depending on other changes to weathering (/sedimentation) that may have been prescribed. i.e. even with pCO_2 restored during 'normal' time-stepping, pCO_2 will change during the accelerated mode if weathering is significantly out of balance with sedimentation. The greater this imbalance, the greater the change in pCO_2 , and the sooner that time-stepping will be handed back to the normal (full updating) mode.

If you have prescribed changing pCO_2 , e.g. a continual ramp upwards, GEMlite is not appropriate in the first place, as the atmosphere is intrinsically assumed to be in equilibrium with the ocean surface and steady-state geochemical gradients in the ocean have been established. (This assumption is broken if CO_2 is rapidly invading the ocean.) Acceleration (and GEMlite) is also not appropriate if ocean circulation and carbon cycling have not yet been spun-up, unless at least 5 to 10 kyr of normal time-stepping forms part of the total spin-up including acceleration.

How can I diagnose changes in the carbon budget due to weathering/sedimentation?

The following example assumes that you are only running with $CaCO_3$ weathering (i.e. silicate weathering and outgassing are both set to zero). In this case the weathering flux of DIC into the ocean is equal to the Ca weathering flux. This is output as a time series in:

```
biogem_series_diag_weather_Ca.res
```

in units of moles per year.

The system is closed with respect to organic matter, so that all POC is remineralised and returned to the ocean. For this reason, the exchange of DIC between the ocean and the sediments is equal to the exchange of Ca. i.e. the exchange of one mole of C is always associated with one mole of Ca, as the system is only open with respect to $CaCO_3$. Therefore the net flux of DIC from ocean to sediments is equal to the difference between `biogem_series_focnsed_CaCO3.res` and `biogem_series_fsedocn_Ca.res`.

The net flux of DIC into the ocean from weathering and sediments is therefore equal to `weather_Ca + fsedocn_Ca - focnsed_CaCO3`.

15.8 Data saving questions

Why is the netCDF data saved at odd times?

There is a default sequence of points in time that **BIOGEM** will save data at. These points are specified in the file: `save_timeslice.dat` (which lives in `cgenie.muffin/genie-biogem/data/input`).¹⁰

This default sequence provides a useful generic starting point.

To specify different save points for an experiment:

1. Edit this file (not recommended).
2. Copy, or create a new file (with the same format). The file that **BIOGEM** uses for saving data is specified by the parameter:

```
bg_par_infile_slice_name='save_timeslice_historicalfuture.dat'
```

(in the example of a historical/future relevant series of save points being requested).

Note that always, at the very end of an experiment, data is automatically saved regardless of whether or not you remembered to specify a save point for the final simulated year.

Refer to the Chapter on **cGENIE.muffin** model output.

¹⁰As a default, the netCDF *time-slices* are saved as annual averages, centered on these points in time.

15.9 Forcings questions

Can I combine forcings together?

Yes ... but it is not quite as simple as in the *user-config* writing:

```
# specify forcings
bg_par_forcing_name="worjh2.FeMahowald2006"
bg_par_forcing_name="pyyyyz.FRpCO2_Fp13CO2"
```

in the example that you wanted to combine an atmospheric CO_2 emissions *forcing* with a surface ocean dust *forcing*, because only the last parameter value in a list of multiple definitions is used. i.e. the above is equivalent to just writing:

```
# specify forcings
bg_par_forcing_name="pyyyyz.FpCO2_Fp13CO2"
```

Instead, you need to create a new forcing (assuming the combined forcing you want does not already exist):

1. Copy/rename one of the two individual *forcing* directories. This will be your new *forcing* name.
2. In the example above – if you have copied the directory for `worjh2.FeMahowald2006`, you simply need to add in the specific atmospheric CO_2 emissions *forcing* forcing files contained in `pyyyyz.FRpCO2_Fp13CO2`, which are:
 - `biogem_force_flux_atm_pCO2_13C_sig.dat`
 - `biogem_force_flux_atm_pCO2_sig.dat`
 - `configure_forcings_atm.dat`

Note that more care has to be taken when combining *forcings* that include the same phase of tracer, i.e. atmosphere and atmosphere, or ocean and ocean. In this case, you need to open up the `configure_forcings_*.dat` file of one *forcing*, and copy the tracer selection line (or lines)¹¹ to the equivalent file in the new *forcing* directory.

Why does my ocean flux forcing does not do anything?

As always if you apply a flux forcing and nothing appears to happen, check:

1. The flux has not been scaled to zero ...
2. The spatial locations, where you expect the flux to be applied, and not on land ((i,j) location is a land, not ocean point), or in the ocean crust ((i,j) is ocean, but the layer chosen is deeper than the ocean floor at that location).
3. That the model run, in time, overlaps with the time-dependent forcing information. e.g. you might start a forcing at year 2010, but only run the model to year 2000 ...

Careful comparison, e.g. difference maps or simply looking at some global diagnostic output provided as in the time-series data format, will confirm whether the impact truly is zero, or just very small. If very small, your issue is mostly simply one of scaling and too small of a flux to make much impact.

¹¹These occur between a pair of tags:

```
-START-OF-DATA-
-END-OF-DATA-
```

Why does my ocean iron flux forcing does not do anything?

Start by referring to above (general flux forcing question).

However, there is a special point of failure of a forcing, unique to the iron system, because there are two different ways of representing *Fe* and *Fe*-related species in **cGENIE.muffin**:

1. In the basic, and original Fe scheme, there are three separate tracers represented in the ocean:

Fe – tracer number 09 – dissolved iron III (Fe).

FeL – tracer number 23 – ligand-bound Fe.

L – tracer number 24 – free ligand (iron binding).

In the forcing definition, a flux of Fe is selected in:

`configure_forcings_ocn.dat`

by:

`-START-OF-DATA-`

`9 f f 0.0 t t -1 01 01 01 '[Fe]'`

`-END-OF-DATA-`

Associated with this selection, is a file of time-dependent information for the forcing:

`biogem_force_flux_ocn_Fe_sig.dat`

and dependent on the nature of the forcing, potentially also a file containing a spatial pattern for the forcing, e.g.

`biogem_force_flux_ocn_Fe_SUR.dat`

Here: it is important to note that both file names contain the tracer short-name: **Fe**.

2. In a newer scheme, there are just 2 tracers:

TDFe – tracer number 90 – total dissolved Fe.

TL – tracer number 42 – total dissolved ligand.

(and e.g. free iron is derived by assuming an equilibrium partitioning based on the total iron and total ligand concentrations).

Why am I telling you all this? For example, configurations using **ECOGEN**, use the newer (two tracer only) representation of iron cycling, whereas in e.g. geoengineering examples, **BIOGEN** is using the older three tracer representation. If you then wish to configure **ECOGEN** to use forcings based on the geoengineering examples, you have to:

1. Firstly, in `configure_forcings_ocn.dat` change the selected tracer number from 9 to 90.
2. Secondly, rename the time-dependent information file, and if present, the spatial file, changing the Fe bit of the filename to TDFe.
3. Lastly, the parameter in the user-config that scales the forcing (if used), has a name that ends in the tracer number and needs to be changed, so rather than e.g.

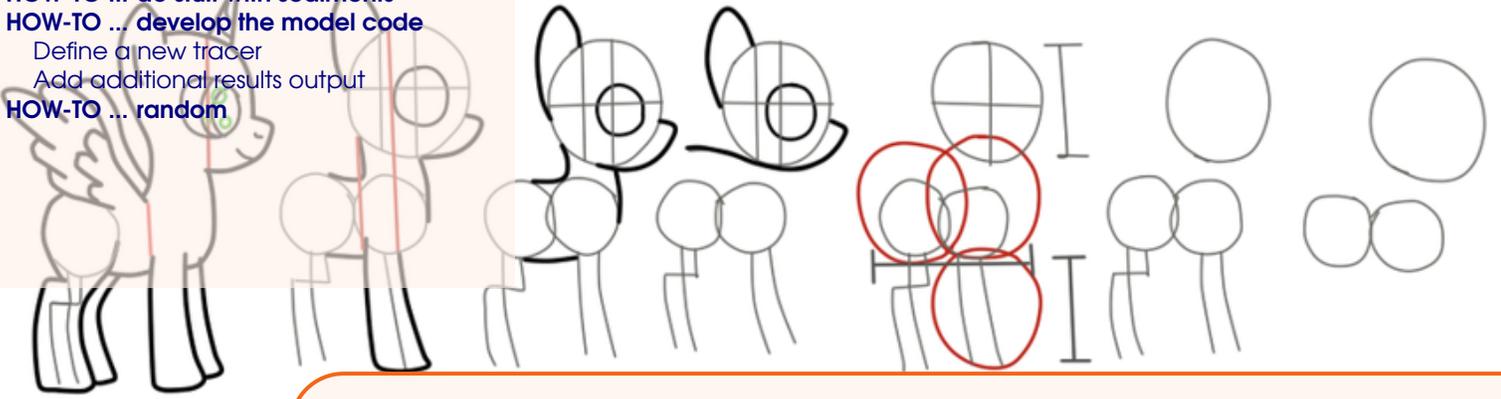
`bg_par_ocn_force_scale_val_9`

you would have:

`bg_par_ocn_force_scale_val_90`

If you select a tracer number in the *forcings* that does not exist in the ocean configuration you are using, such as the 'wrong' iron tracer – this is why the *forcing* appears not to do anything.

HOW-TO ... get started with cGENIE.muffin
HOW-TO ... (general)
HOW-TO ... change model output
HOW-TO ... screw with the climate system
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cycles
HOW-TO ... force the system
HOW-TO ... do stuff with sediments
HOW-TO ... develop the model code
 Define a new tracer
 Add additional results output
HOW-TO ... random



16. HOW-TO

What follows are potted HOW-TO instructions for doing things.
There is some overlap with the FAQ Chapter, so please read both!

16.1 HOW-TO ... get started with cGENIE.muffin

Install cGENIE

See: cGENIE *Quick-start Guide*. (Also refer to the READ-ME file for e.g., details of changes in configuring and running cGENIE compared to GENIE.)

Find configurations for cGENIE

A series of (example) cGENIE configurations are provided, many of which are detailed in full in the cGENIE *Tutorial* document. Example configurations comprise *base-config* and *user-config* files, plus any *forcings* needed.

- cGENIE *base-configs* are stored in:
/cgenie/genie-main/configs
and all start with 'cgenie_', for example:
cgenie_eb_go_gs_ac_bg_hadcm3l_eocene_36x36x16_2i_080928_BASE.config
- cGENIE user configs are stored in:
/cgenie/genie-userconfigs
- cGENIE forcings are stored in:
/cgenie/genie-forcings

16.2 HOW-TO ... (general)**Do some thing dumb**

Easy! Just close your eyes and change some parameter values at random. Better still, start using the model without reading the manual first ...

16.3 HOW-TO ... change model output

Set the frequency of time-series and time-slice output

See: *cGENIE user-manual* (section 5).

Diagnose orbital (insolation) changes

Two new misc category time-series files have been provided:

```
biogem_series_misc_ocn_insol.res
```

and

```
biogem_series_misc_ocn_swflux.res
```

with the the SW (shortwave) flux (`swflux`) being equivalent to the incident strength of solar radiation at the surface (`insol`) but accounting for the prescribed planetary albedo. Both variables are calculated and saved on a global mean ocean grid basis (2nd data column) and have units of $W\ m^{-2}$. In addition, to help diagnose orbital variability, `biogem_series_misc_ocn_insol.res` includes two further insolation variables (3rd and 4th columns). These reflect the strength of insolation at a single point in the annual cycle and at discrete latitudes (i.e. `j` grid indices). (The insolation at 2 different latitudes are saved so that both .e.g. N and S hemisphere insolation signals can be simultaneously recorded.)

Three new namelist parameters are provided to configure this:

1. `bg_par_t_sig_count` – which sets the BIOGEM 'time-step' in the annual cycle at which the insolation value will be saved. e.g. for 96 time-steps in the ocean physics, and a 2:1 GOLDSTEIN:BIOGEM gearing (the default for the 16 level configuration), there are 48 BIOGEM time-steps. (It is left to the user to work out which part of the annual cycle *cGENIE* starts at (i.e. time-step #1) – I haven't a clue ...)
2. `bg_par_sig_j_N` – sets the 'j' value for a northern hemisphere (but could be southern) snapshot.
3. `bg_par_sig_j_S` – sets the 'j' value for a southern hemisphere snapshot.

By default `bg_par_sig_j_N` is assigned a value of 2 and `bg_par_sig_j_S` a value of 1 (on account of the maximum grid size not being *a priori* known). The default setting of `bg_par_t_sig_count` is 1 – i.e. the first (BIOGEM) time-step in the annual cycle.

16.4 HOW-TO ... screw with the climate system

Set/un-set seasonal insolation forcing

Seasonal insolation forcing of the **EMBM**, **GOLDSTEIN** ocean, and sea-ice model, are set by the following parameters¹:

```
ea_dosc=.true.
go_dosc=.true.
gs_dosc=.true.
```

and are `.true.` by default. To set an annual average insolation forcing with no seasonality, simply set these to `.false.`.

Adjust solar forcing in a time-dependent manner

The value of the solar constant in cGENIE is set by the *namelist parameter*:

```
ma_genie_solar_constant
```

which by default is set to 1368 Wm^{-2} , i.e.:

```
ma_genie_solar_constant=1368.0
```

Specifying a different value for `ma_genie_solar_constant` in the *user-config* file allows the solar forcing of the EMBM to be altered. For example, to induce a 'snowball Earth' like state under a solar constant applicable to the late Neoproterozoic (some 6% less than modern) you would set:

```
ma_genie_solar_constant=1330.56
```

Modification of `ma_genie_solar_constant` can be turned into a time-dependent forcing of solar forcing, but only by frequent re-starting using a sequence of short model integrations.

Alternatively, a crude (temporary) hack is provided to allow a semi-continual adjustment of solar forcing. Whether you wish to vary the solar constant in a time-dependent manner is determined by the *parameter*:

```
bg_ctrl_force_solconst
```

By default this is set to `.false.`. By adding to the *user-config* file:

```
bg_ctrl_force_solconst=.true.
```

a time-varying change in the value of the solar constant will be imposed. For this, **BIOGEM** will expect the presence of a file called `biogem_force_solconst_sig.dat` in the forcing directory². This file must contain two columns of information: the first is a time marker (year) and the second is a paired value for the solar constant. In the current crude incarnation of this feature, the time markers (1st column) **must** correspond exactly to the time markers in the time-series specification file³. **cGENIE.muffin** will exit with an appropriate error message if this is not the case.

When using the time-varying solar constant hack, seasonal solar insolation is re-calculated each year with a call to `radfor(genie_solar_constant)`⁴ at the start of the time-stepping loop⁵. At

¹These are typically set in the *base-config* if needed (i.e. different from default).

²REMEMBER: The location of which is specified by the namelist parameter `bg_par_fordir_name`.

³REMEMBER: The filename of which is specified by the namelist parameter `bg_par_infile_sig_name`.

⁴`cgenie.muffin/genie-embm/src/fortran/radfor.F`

⁵`cgenie.muffin/genie-main/genie.F`

each time-marker, **BIOGEM** sets the value of `genie_solar_constant` equal to the corresponding value specified in `biogem_force_solconst_sig.dat`. Thus, regardless of how closely-spaced the time-marker years are, (seasonal) solar insolation is only adjusted every year. For a longer time-marker interval than yearly, no interpolation is performed on the series of solar constant values, and in this way time-dependant solar forcing currently differs from the calculation of other *forcings*.

A simple example file might look something like:

```
-START-OF-DATA-  
0.5  1367.0  
1.5  1366.0  
2.5  1365.0  
3.5  1364.0  
4.5  1363.0  
5.5  1362.0  
6.5  1361.0  
7.5  1360.0  
8.5  1359.0  
9.5  1358.0  
10.5 1357.0  
-END-OF-DATA-
```

which will decrease the value of the solar constant by 1 Wm^{-2} each year. Note that because the solar forcing is only updated each year (with the call to `rcdfor.F`), the first year will be characterized by climate with a solar constant of 1368 Wm^{-2} , the default. Although **BIOGEM** sets a new value of `genie_solar_constant` (1367 Wm^{-2}) mid way through the first year, it is only at the start of the second year that solar insolation is recalculated according to the reduction in solar constant.

Hacking the solar constant in a time-varying manner is, of course, a (albeit crude) way of addressing SRM geoengineering impacts.

16.5 HOW-TO ... configure ocean biogeochem. cycles

Configure an abiotic ocean

Biological productivity in the ocean can be completely turned off to create an abiotic ocean (why you would want to do this is another matter ... perhaps analyzing the solubility pump or a 'deep-time' and prior to significant marine life study ... (?)). The biological option is set by the *parameter*:

```
bg_par_bio_prodopt
```

which by default takes a value of "1N1T_P04MM" which selects the scheme described in *Ridgwell et al.* [2007a]. To have no biological production in the ocean, add the following line to the end of the *user-config* file⁶:

```
bg_par_bio_prodopt="NONE"
```

With this set, you do not have to specify any biological production or remineralization *namelist parameter* values in the *user-config* file.

Add a water mass age tracer

Water masses (and hence something of ocean circulation) can be tagged with a color (dye) tracer. However, on its own, this can tell you nothing about water mass age. A second color tracer can be added, however, and configured in such a way that by analysing the ratio of the two tracers, water mass age (time since a parcel of water last saw the surface ocean on average).

First off, you are going to need a *base-config* that defines both color tracers. An example of this (but with no biology or carbon cycle and a modern configuration) is:

```
cgenie.eb_go_gs_ac_bg.worjh2.rb
```

Obviously, this can be adapted or the 2 lines selecting the 'red' and 'blue' color tracers, copied over into a different *base-config* (but remembering that the total number of ocean tracers selected then increases by 2).

The way this is going to work is:

1. A restoring of red dye is applied evenly to the entire ocean surface. By itself, this will simply result in the ocean progressively filling up with dye until equal to the surface concentration and with no time information.
2. So a blue dye is also injected. The concentration of this is also restored at the surface. However, the surface concentration of the blue dye is scaled such that it reflects age in the model experiment. This counts 'down', such that at the start of the experiment the dye is at its highest concentration and hence representing the greatest amount of time (age). As the run progressive and time runs towards zero, so does the dye flux. i.e. for for an experiment running for 10,000 years, the concentration of blue at the surface linearly declines from 10,000 to 0.

Or alternatively:

3. So far, even with the blue dye reflecting 'time', remote parts of the ocean will not have received much dye, so even though the water should be 'old' and the surface concentration high, the concentration and hence 'age' in the deep ocean will still be low. So the red dye is used to normalize for the dispersion and dilution of the blue dye.

⁶Or edit the existing line under the section '-- BIOLOGICAL NEW PRODUCTION --'

In the original, and still valid way of doing this, 2 surface ocean *forcings* need be specified in order to create the combined age tracer. An example forcing is provided: *forcing*: pyyyyz.Rcolr_Rcolb, which is yhen configured, for a 10,000 year run in this example, by adding the following 2 parameter settings in the *user-config*:

```
bg_par_ocn_force_scale_val_49=10000.0
bg_par_ocn_force_scale_time_49=10000.0
```

(If the experiment duration is longer than 10,000 years, the parameter values need be adjusted accordingly.)

In this methodology, in the netCDF output, the concentration ratio of blue/red, should be 'age' – the mean time that a parcel of water was last at the surface.

This is all quite a *faff*⁷. So a simpler methodology has been implemented. As *base-config* file with both red and blue color tracers defined is still required, but now, rather than a set of *forcings* (and associated forcing configuration), a single parameter is added in the *user-config* file:

```
bg_ctrl_force_ocn_age=.true.
```

This will automatically create the age tracer and additional explicitly output (in netCDF) both the total age of a water parcel, as well as the age relative to the surface (ventilation age).

Example *user-config* files for both approaches are provided:

```
EXAMPLE.worjh2.NONE_age.SPIN
EXAMPLE.worjh2.NONE_colage.SPIN
```

Note that the automatic approach (EXAMPLE.worjh2.NONE_colage.SPIN) will handle experiments started from a *re-start* (but not for the manual approach). The only advantage to the manual approach (EXAMPLE.worjh2.NONE_age.SPIN), which is provided for backwards code/experiment compatibility, is that it is possible to specify a surface age for a specific region, e.g. North Atlantic, meaning that the ventilation age is the time since a parcel of water last saw the North Atlantic rather than anywhere at the surface (as in the automatic approach).

Prescribe the CaCO₃:POC export ratio

In the default⁸ 'biological' scheme in GENIE the CaCO₃:POC export ratio from the surface ocean in BIOGEM is parameterized as a power law function of the degree of ambient over-saturation w.r.t. calcite [Ridgwell *et al.*, 2007a,b]. The calculated CaCO₃:POC ratio will vary therefore both spatially, particularly w.r.t. latitude (and temperature), as well as in time, if the surface ocean saturation state changes. The latter can arise from climatic (temperature) or circulation changes, or through a change in the DIC and/or ALK inventory of the ocean (such as resulting from emissions of fossil fuel CO₂) or the re-partitioning of these species vertically within the ocean (e.g., as a result of any change in the strength of the biological pump).

There may be situations in which it is advisable to hold the CaCO₃:POC export ratio invariant. For instance, considering the current very considerable uncertainties in the impacts of ocean acidification on marine calcifiers [Ridgwell *et al.*, 2007a] the safest assumption is arguably to exclude any acidification impact on calcification and carbonate export. Specifying a spatially uniform value of the CaCO₃:POC ratio ratio (e.g. 0.25 or 0.3) also allows comparison with the results of early

⁷Google it.

⁸The default biological scheme is given by: `bg_par_bio_prodopt='1N1T_P04MM'`.

carbon cycle model studies. For deeper-time geological studies where little about marine carbonate production may be known *a priori*, a spatially uniform value represents the simplest possible assumption (e.g., *Panchuk et al.* [2008]).

BIOGEM can be told to use a prescribed (spatially and temporally invariant) 2D field of CaCO₃:POC export rain ratios (instead of calculating these internally as a function of ocean chemistry) by setting the 'Replace internal CaCO₃:POC export rain ratio?' *namelist parameter* flag to `.true.`:

```
bg_ctrl_force_CaCO3toPOCrainratio=.true.
```

You must also then provide a 2D data field that specifies the value of the rain ratio at each and every surface ocean grid point. The filename of this field is set by default to:

```
bg_par_CaCO3toPOCrainratio_file="CaCO3toPOCrainratio.dat"
```

and the file must be located in the 'BIOGEM data input directory'⁹, which by default is:

```
bg_par_indir_name=" $RUNTIME_ROOT/genie-biogem/data/input"
```

This 2-D field must be in the form of an ASCII file with space (or tab) separated values arranged in rows and columns of latitude and longitude. The format of the file must follow the GOLDSTEIN ocean grid with the first line being the most Northerly row, and the last line the most Southerly row of grid points. Data along a row is from West to East. The latitude of the first column of values must be consistent with the defined starting latitude of the model grid, which is specified by the namelist parameter `gm_par_grid_lon_offset`¹⁰. Examples are given in the code repository¹¹.

If you are using a uniform value, it is an easy enough job to create a 36×36 array of the value you so desire¹².

If you want to hold a previously-calculated (spatially variable) CaCO₃:POC field constant, then the easiest way to achieve this is to copy the information contained in the *time-slice* results field:

`misc_sur_rCaCO3toPOC` in the results netCDF file `fields_biogem_2d.nc`¹³. Because this is a 3D data field ($36 \times 36 \times 8$), carefully highlight just the surface ocean (2D) distribution (e.g., from the Panoply viewer) or extract from the netCDF file by some other means, and then copy and paste into:

`CaCO3toPOCrainratio.dat` (or whatever you have specified the filename as). When copying Panoply data, 'NaN's should be replaced by values of zero. Take care that the final (steady-state) time-slice is being copied and not the first (un-spunup) one ...

TIP: In order to quantify the importance of calcification feedbacks with CO₂ and climate, two model integrations are required: one with the CaCO₃:POC ratio held constant and the other with it allowed to vary, thereby allowing the effect of a changing CaCO₃:POC ratio on the system to be elucidated.

Implement an alternative fixed remineralization profile for POC (e.g. Martin curve)

There are several options for utilizing a fixed remineralization profile for POC, which by default is a double exponential (See: *Ridgwell et al.* [2007a]). The fixed remineralization profile scheme is set by the string parameter: `bg_par_bio_remin_fun`. By default, it has a value of 'efolding'. Currently available options are:

⁹`$RUNTIME_ROOT` being equal to `~/genie`.

¹⁰-260E by default

¹¹e.g., `~/genie/genie-biogem/data/input/CaCO3toPOCrainratio_worbe2_preindustrial.dat`

¹²It doesn't matter if you specify a value over land because only values associated with wet cells will be acted on.

¹³You must have the 'miscellaneous properties' time-slice save flag set to:

`bg_ctrl_data_save_slice_misc=.true.` (the default) for this field to be saved.

- Martin1987, which applies a globally-uniform power, set by:
`bg_par_bio_remin_martin_b`
 (which by default has a value of -0.858)
- Henson2012, which calculates the value of *b* according to sea surface temperature (SST):
 $b = (0.024 * SST) - 1.06$

To user either (on their own), all organic matter should be assigned to a single phase, with the 2nd (recalcitrant) fraction set to zero:

```
bg_par_bio_remin_POC_frac2=0.0
```

Note that these parameterizations can be combined with ballasting and will act on the 'free' POC phase (i.e. the one not controlled by the ballasting parameterization).

Implement particulate organic carbon 'ballasting'

The default particulate organic carbon (POC) ocean interior remineralization scheme is based on fixed, prescribed profiles of relative POC flux to depth (e.g. see: *Ridgwell* [2001]; *Ridgwell et al.* [2007a]). A 'ballasting' control on POC transport to depth can instead be implemented by:

```
bg_ctrl_bio_remin_POC_ballast=.true.
bg_ctrl_bio_remin_POC_fixed=.false.
```

The POC 'carrying coefficients' for CaCO₃, opal, and detrital (lithogenic) material are set by the parameters:

```
bg_par_bio_remin_ballast_kc
bg_par_bio_remin_ballast_ko
bg_par_bio_remin_ballast_kl
```

(for CaCO₃, opal, and lithogenics, respectively). Note that the ballast coefficient units are: g POC m⁻² yr⁻¹ (g ballast m⁻² yr⁻¹)-1 (i.e. **g g⁻¹**), which are internally converted to: mol POC m⁻² yr⁻¹ (mol ballast m⁻² yr⁻¹)-1 (i.e. **mol mol⁻¹**).

A fixed (in time), but spatially heterogeneous field can also be prescribed instead of global uniform values (akin to setting a pattern of the CaCO₃:POC export rain ratio (??)). The parameters setting whether to substitute a globally-uniform value with a specified pattern are:

```
bg_ctrl_force_CaCO3ballastcoeff=.true.
bg_ctrl_force_opalballastcoeff=.true.
bg_ctrl_force_detballastcoeff=.true.
```

and which by default are `.false.`. The patterns of carrying coefficient are determined by files read in from `cgenie/genie-biogem/data/input`. The filenames are specified by:

```
bg_par_CaCO3ballastcoeff_file
bg_par_opalballastcoeff_file
bg_par_detballastcoeff_file
```

(again, akin to the methodology for setting the CaCO₃:POC export rain ratio (??)).

Note that ballasting is combined with an e-folding (or other) fixed profile remineralization schemes¹⁴. Ballasting is calculated with respect to the 2nd (recalcitrant) fraction of POC only.

¹⁴Although in a sense, the remineralization of POC is not 'fixed' in that it does not have a predetermined profile but instead is set by the changing flux of CaCO₃, opal and lithogenic fluxes with depth, `bg_ctrl_bio_remin_POC_fixed` should still be set to `.true.` (the default).

The remaining POC export is degraded by an alternative algorithm, which by default, is an e-folding decay (see previously for more and alternatives). The fraction of initial export assigned to ballasting vs. 'free' POC is calculated according to the available exported ballast flux.

Prescribe biological export production

Two possibilities:

1. Via a full prescription of all particulate fluxes in surface ocean

Create a full set of particulate (sediment tracer) flux forcings fields for the surface ocean, one for each biologically-related sediment tracer selected in the model, including isotopes (and trace metals). Everything except for the surface layer can be left as a zero (0.0) in the two 3D spatial fields required for each tracer.

You must also create a set of dissolved (ocean) tracer flux forcings fields for the surface ocean, one for each dissolved tracer associated with the particulates and selected in the model (including isotopes etc). The dissolved tracer flux fields must be create so as to exactly cancel out the particulate fields to conserve mass. For most tracers this is trivial, i.e., the fields for P in particulate organic matter (sed_POP) need be associated with fields for dissolved PO₄ (ocn_PO4) which will simply be equal in magnitude but opposite in sign to POP. Complications start to arise for CaCO₃ and there is also the question of alkalinity changes associated with organic matter creation/destruction (via changes in NO₃), so this method, whilst the most flexible, is not without its complications and degree of tediousness (i.e. would not recommend).

2. By just prescribing just the POC flux

An alternative has been provided enabling a full biological productivity in the surface ocean, but controlled by prescribing just the particulate organic carbon export flux. This 'biological' scheme is selected with:
`bg_par_bio_prodopt="bio_POCflux"`
 What happens in practice is that the POC flux is used to calculate the equivalent PO₄ change in the surface ocean, and then this is passed to the biological scheme and export production calculated 'as usual'. (The POC flux forcing is set to zero once the associated PO₄ (uptake) flux has been calculated.)

A particulate (sediment tracer) flux forcing for POC in the surface ocean still has to be defined and selected, but no other *forcings* (including the associated removed dissolved tracers) are required. An example forcing configuration is given in `worjh2.FPOC_Caoetal2009` (and selected by:

```
bg_par_forcing_name="worjh2.FPOC_Caoetal2009"
```

) An example *user-config*: `EXAMPLE.worjh2.Caoetal2009_FPOC` illustrating this is provided.

NOTE: Take care with dissolved organic matter (DOM) production, as the specified POC flux is automatically increased to take into account DOM production. i.e. is 50% of export is specified to be partitioned into dissolved rather than particulate organic matter export, whatever is specified in the POC export forcing would be internally doubled, before being partitioned into POM and DOM. **NOTE:** Also take care with the units of the flux *forcing* to the surface layer in the ocean, which are in mol yr⁻¹. The main particulate flux output is in units of mol m⁻² yr⁻¹, but since cGENIE is invariably run on a equal area grid it is not difficult to convert export production densities to mol yr⁻¹ (you either need to divide the area of the Earth's surface by the number of grid points, or save the ocean grid information – see

netCDF save options in the User Manual). Alternatively, as of revision *r8825*, *cGENIE* saves the primary particulate flux fields also in units of mol yr⁻¹ (assuming you have the biological or full netCDF save options selected – see netCDF save options in the User Manual).

Be aware that if there is insufficient PO₄ to support the require POC flux, the entire POC flux will still be created, meaning that you may end up with regions of negative nutrient concentration.

Include a R-DOM cycle in the ocean

R-DOM degradation

The parameter: `bg_ctrl_bio_remin_RDOM_photolysis` determines whether RDOM degradation is restricted to the surface layer and occurs only by/associated with photolysis. It can be `.true.` or `.false.` and by default is set to:

```
bg_ctrl_bio_remin_RDOM_photolysis=.false.
```

When set `.true.`, RDOM degradation is set to zero everywhere in the ocean except the surface layer. Here, the lifetime (parameter: `bg_par_bio_remin_RDOMlifetime`) is modified in **inverse** proportion to the solar insolation integrated over the surface layer. (There is a field in the 2D netCDF of solar insolation at the ocean surface, and the average over the surface layer is approx 1/4 of this.) i.e., in lower latitude and higher insolation regions, the lifetime is shorter than specified by `bg_par_bio_remin_RDOMlifetime` (and by approx a factor of 1/4 of the solar insolation in W m⁻²).

Include a Cd cycle in the ocean

In order to run *cGENIE* with ocean cadmium cycle, the following *base config*: `cgenie_eb_go_gs_ac_bg_itfclsd_16l_JH_BASEFeCd` is provided.

A typical experiment command line, using the *user config* file: `EXAMPLE_worjh2_PO4Fe_Cd_SPIN` (also provided under SVN), would look like:

```
./runCCgenie.sh cgenie_eb_go_gs_ac_bg_itfclsd_16l_JH_FeCdBASE /
EXAMPLE_worjh2_FeCd_SPIN 11
To submit this job to the cluster (from $HOME):
qsub -q kitten.q -j y -o cgenie_log -S
/bin/bash subcgenie.sh cgenie_eb_go_gs_ac_bg_itfclsd_16l_JH_BASEFeCd /
EXAMPLE_worjh2_PO4Fe_Cd_SPIN 10001
```

Include an iodine cycle in the ocean

In order to run *cGENIE* with a marine iodine cycle, one of the following *base-configs* is needed:

```
cgenie.eb_go_gs_ac_bg.worjh2.BASEI
cgenie.eb_go_gs_ac_bg.worjh2.BASEI
cgenie.eb_go_gs_ac_bg.worbe2.BASESI
```

Of these, recommended is one of the 16-level ocean `worjh2` configurations¹⁵ as the oxygen minimum zones in the 8-level ocean `worbe2` configuration are much more poorly developed (see *Ridgwell et al.* [2007a]).

Example configurations of several different levels of complexity of iodine cycling are given in the follow sessions.

¹⁵The difference between the two `worjh2` configurations is that one (with the 'S' in 'BASESI' also includes a sulphur cycle.

Basic ('abiotic') iodine cycle

This section outlines the most trivial possible configuration of the marine iodine cycle, in which the only processes are:

1. Reduction of IO_3^- to I^- under dysoxic conditions
2. Re-oxidation of I^- to IO_3^-

An example experiment configuration *user-config* – `EXAMPLE.worjh2.P04Ibasic.SPIN` – is given. In this, the available parameters (listed under the heading '# -- MISC --' and '### IODINE CYCLE CONTROLS ###') controlling the iodine cycle are:

- # set no biological IO_3^- uptake
For simplicity, this experiment configuration sets the iodate uptake in organic matter associated with biological production at the ocean surface, to zero. The parameter `bg_par_bio_red_POC_POI` specifies the ratio of I to C in new production (the cellular quotient) and is set to zero.
- # select basic reduction and oxidation options
There are various alternative options for how IO_3^- is reduced in dysoxic conditions. The simplest parameterization is specified here;
`bg_opt_bio_remin_reduce_IO3toI='threshold'`,
in which a threshold of dissolved oxygen is prescribed. In any regions (i.e. model grid boxes) of the ocean in which dissolved oxygen concentrations fall are below this, IO_3^- is completely reduced to I^- .
Alternative options also exist for how the re-oxidation of I^- occurs. In the simple parameterization specified here;
`bg_opt_bio_remin_oxidize_ItoIO3='lifetime'`,
a fixed lifetime of I^- in the ocean is prescribed. Oxidation proceeds at this rate regardless of the oxygenation state of the ocean, but as long as sufficient oxygen to accomplish the reaction $2\text{I}^- + 3\text{O}_2 \rightarrow 2\text{IO}_3^-$ exists.
- # set [O₂] threshold (mol kg^{-1}) for (complete) reduction of I.
The parameter `bg_par_bio_remin_cO2_IO3toI` sets the dissolved oxygen concentration threshold (mol kg^{-1} , below which IO_3^- will be reduced.
- #set I lifetime (yrs)
Finally, the parameter `bg_par_bio_remin_Ilifetime` then sets the lifetime of I^- in years.

Model output is saved in the 'normal way' (refer to the User Manual) and amongst the ocean tracers ('ocn_*'), are the tracers of dissolved iodide and dissolved iodate (in units of mol kg^{-1}). Model output can also be contrasted with observed data re-gridded to the cGENIE (worjh2) grid. A typical command-line launching of a model experiment (10000 years integration in this case) would be:

```
./runmuffin.sh cgenie.eb_go_gs_ac_bg.worjh2.BASEI /
EXAMPLE.worjh2.P04Ibasic.SPIN 10000
```

'Biotic' component to the iodine cycle

This section outlines the next component of the marine iodine cycle, involving phytoplankton, with the two processes:

1. uptake of IO_3^-
2. remineralization/release of I^-

The way this cycling in the model works is as follows: First, as plankton biomass is created, alongside C and P (and if selected N, Fe and other trace elements), IO_3^- is taken up by the cell, in a specified proportion to carbon (see below). Of this biomass, a proportion is assumed to be exported in particulate organic matter (POM) form beneath the euphotic zone, with the remainder converted to labile dissolved organic matter (DOM) (see: *Ridgwell et al.* [2007a] for details). When either POM or DOM is remineralized and elemental constituents released, rather than returning IO_3^- back to solution, it is assumed that the dissolved iodine is in the form of I^- . (This also requires an accounting of release of O_2 , as IO_3^- is taken up by the cell and assume to be instantaneously internally reduced.) The result is: (i) a progressive transformation $\text{IO}_3^- \rightarrow \text{I}^-$ in the surface ocean as DOM is continually created and destroyed, and (ii) a 'nutrient-like' enrichment of I^- in the sub-surface as a consequence of the remineralization of POM.

An example experiment configuration *user-config* – EXAMPLE.worjh2.P04Ibio.SPIN – is given. In this, the available parameters (listed under the heading '# -- MISC --' and '### IODINE CYCLE CONTROLS ###') controlling the iodine cycle are:

- # select option for no watercolumn reduction
For simplicity, this experiment configuration sets the reduction of iodate under low oxygen conditions in the water to zero (i.e. leaving the biological pump as the only source of I^-);
`bg_opt_bio_remin_reduce_IO3toI='NONE'`
- # set biological IO_3^- uptake
Set a cellular I:C quotient. Here, a value of $1.0\text{E-}4$ is given as an example;
`bg_par_bio_red_POC_POI = 1.0E-4`
- # set I lifetime (yrs)
Finally, the parameter `bg_par_bio_remin_Ilifetime` then sets the lifetime of I^- in years. (Here, for illustrative purposes, the lifetime is increased to 10 years compared to 1 year in the basic Example.)

Determine the CH_4 flux required to achieve a particular atmospheric p CH_4 value

Unlike the concentration of CO_2 in the atmosphere, which if restored to a chosen value during a *spin-up* experiment, will remain at that value in a *continuation* experiment (if no other perturbation of the carbon cycle or CO_2 emissions have been prescribed), CH_4 in the atmosphere decays with a lifetime of ca. 8 years (with a small fraction dissolving in ocean surface waters and being oxidized in the ocean). Hence, atmospheric CH_4 restored to a particular value in a spin-up, requires that restoring to be maintained in any *continuation* experiment or CH_4 will quickly decay to zero. However, doing this (*restoring* CH_4 concentrations), prevents the effect of CH_4 emissions on being assessed (as the atmospheric composition is being held constant).

An alternative would be to carry out the *spin-up* experiment with no *restoring* of atmospheric CH_4 (or *restoring* to zero), and then run the *continuation* experiment with no CH_4 restoring. This would enable e.g. CH_4 emissions experiments to be carried out and the change in atmospheric CH_4 in response to be simulated. The problem here is that the lifetime of CH_4 in the atmosphere scales with the CH_4 concentration. So in starting with no CH_4 in the atmosphere, the CH_4 lifetime is relatively short, and the response to CH_4 emissions will be underestimated.

What is in effect 'missing' are the (natural) sources of CH_4 to the atmosphere such as wetlands, which at steady state, provide a CH_4 flux that balances the oxidation rate of CH_4 in the atmosphere (and ocean). cGENIE has a *parameter* for this: `ac_par_atm_wetlands_FCH4` (mol yr^{-1}) (with the isotopic composition of this source set by: `ac_par_atm_wetlands_FCH4_d13C`). All that then

remains is to determine the flux of CH₄ that balances the rate of oxidative loss for the desired atmospheric CH₄ concentration. To do this:

1. Carry out a *spin-up* with atmospheric CH₄ *restored* to the desired concentration.¹⁶
2. Determine the total loss rate of CH₄ (including both atmospheric oxidation and invasion (and subsequent oxidation) into the ocean) – this is recorded in the *time-series* results file: `biogem_series_focnatm_pCH4.res`¹⁷.
3. Set the *parameter* `ac_par_atm_wetlands_FCH4` equal to this value.

An example of a spin-up in which a prescribed ('wetland') flux of CH₄ to the atmosphere is set, is described in:

cGENIE.Examples – *spin-up* example `EXAMPLE_p0055c_P04_CH4_SPIN2`

Modify the ocean inventory of a tracer of a *re-started* experiment

There are three different ways in which for a closed system, the inventory of a tracer can be modified:

1. Add a flux forcing, to the ocean surface or the ocean as a whole. The tracer change is then the total global flux times the duration of the forcing. Note that the forcing can be positive or negative (to effect a decrease in the tracer inventory)
2. There is a parameter that add to or subtract from, the tracer inventory at the very beginning of an experiment (and assuming it is running on from a *re-start*). The parameter name is `bg_ocn_dinit_nn`, where `nn` is the tracer *number* (which can be found in the parameter list table PDF, or by inspection of the file `tracer_define.ocn` (for ocean, dissolved tracers) in the directory `cgenie.muffin/genie-main/data/input`. For example:
`bg_ocn_dinit_8=1.0E-6`
 will add 1 μM kg⁻¹ of PO₄ (#8 is the tracer number of dissolved phosphate), uniformly to the ocean. Note that a negative value will result in the subtraction of a uniform concentration form every grip cell in the ocean (meaning that care has to be taken to ensure that negative numbers to not appear following subtraction).
3. There is a variant to the concentration adjusting parameter that is enabled by setting the parameter `bg_ctrl_ocn_dinit` to `false` (it is `true` by default). `bg_ocn_dinit_nn` now acts as a scaling factor that is applied to the tracer concentration field. The new concentration field is equal to the old concentration field (from the *re-start*), times $(1.0 + \text{bg_ocn_dinit_nn})$, e.g.:
`bg_ctrl_ocn_dinit=.false.`
`bg_ocn_dinit_8=0.5`
 will result in a 50% increase in the concentration of dissolved phosphate everywhere in the ocean (and a value of 1.0 doubles concentrations). Conversely, a value less than one will result in a proportional reduction everywhere, e.g.:
`bg_ctrl_ocn_dinit=.false.`
`bg_ocn_dinit_8=-0.2`
 generates a 20% decrease everywhere.

Obviously, if the experiment is not being run from a re-start, or is being run from a re-start which

¹⁶For an example: see experiment `EXAMPLE_p0055c_P04_CH4_SPIN` described in *cGENIE.Examples*.

¹⁷Second column (the value in units of mol yr⁻¹)

does not include the particular tracer, then the initial value of the trace can be set. The parameter name is `bg_ocn_init_nn`, where `nn` is the tracer *number*.

Determine the atmospheric radiocarbon flux required to achieve a steady state ^{14}C system

First, you need to spin the system with the atmosphere restored to the e.g. pre-industrial $\Delta^{14}\text{C}$ value desired. There are two ways then to diagnose the equivalent ^{14}C flux:

1. Sum the total mol inventory of $\Delta^{14}\text{C}$ in the ocean (as DIC) and atmosphere (CO_2), both of which can be found in the relevant time-series output. The radiocarbon decay rate multiplied by the (steady state) inventory then gives the total decay, which of course is equal to the cosmogenic input flux needed to maintain steady state.
2. Rather simpler, enabled by some recent output code changes, is simply to read off the reported atmospheric flux forcing that is being applied to restore the atmosphere $\Delta^{14}\text{C}$ value. The relevant time-series output file is:

```
biogem_series_diag_misc_specified_forcing_pCO2_14C.res
```

Once the balancing ^{14}C flux value has been obtained, the namelist parameter controlling the rate of cosmogenic ^{14}C production (by default, zero): `ac_par_atm_F14C`

16.6 HOW-TO ... *force* the system

Configure an abiotic ocean

Note: All the *forcings* described here assume a newer (simplified) methodology for prescribing *forcings*.¹⁸ This methodology is enabled by setting:

```
bg_ctrl_force_oldformat=.false.
```

This is set automatically as part of the `runmuffin.sh` shell script¹⁹.

Taking the example of the ocean (dissolved tracers): flux and restoring *forcings* are defined in the *forcings* specification file: `configure_forcings_ocn.dat`. As detailed in the notes to this file, there is a flag (COLUMN #6) which sets the spatial attributes of the *forcing* as follows:

- 3 – '3D' – force the entire ocean volume uniformly
- 2 – '2D' – force the entire ocean surface uniformly
- 0 – '0D' – force a specified point
- 1 – 'SURFACE' – force with/to specified surface ocean pattern
- 2 – 'BENTHIC' – force with/to specified benthic pattern
- 3 – 'LEVEL' – force a specific layer in the ocean with/to specified pattern²⁰
- 4 – 'SURFACE+BENTHIC' – simultaneously force with/to specified (and sperate) surface and benthic pattern

The default (3) is that the forcing is applied uniformly to the entire (3D) ocean volume.

Options 3, 2, and 0, as: uniform 3D²¹ (volume), uniform 2D (surface), and point forcing, respectively, require no additional (spatial) information and only an additional file specifying the time-dependent information for each forcing need be provided:

```
biogem_force_flux_ocn_xxx_sig.dat
```

for flux forcings, and

```
biogem_force_restore_ocn_xxx_sig.dat
```

where: xxx represents the mnemonic of the tracer (e.g., DIC is dissolved inorganic carbon. CH4 is methane, etc.).

Options -1 (SURFACE), -2 (BENTHIC) -3 and -4, require a 2D field to be provided, in addition to the time-dependent information for each forcing. The grids for both are the same – i.e., all 'wet' grid points (non dry land) in the model. The filename for these 2D files is of the form:

```
biogem_force_flux_ocn_xxx_SUR.dat
```

for flux forcings, and

```
biogem_force_restore_ocn_xxx_SUR.dat
```

with BEN for the equivalent BENTHIC, and LEV for LEVEL *forcing* selections.

¹⁸For details of the 'old', fully 3D spatially-explicit forcing methodology, refer to the *cGENIE user-manual*.

¹⁹(but it not the actual default *namelist parameter* setting)

²⁰For `k=k_max`, this duplicates the SURFACE forcing.

²¹Note that here: '3D' does not mean a spatially explicit 3D pattern and hence the original ('old') way of specifying *forcings*, but instead: that the forcing is applied uniformly in 3D space (i.e., is in effect a volume *forcing*).

Applying a geoengineering 'liming' flux to the ocean surface

There are two different methodologies provided for: (1) applying a surface flux of alkalinity (and Ca, and DIC) with a uniform or spatial pattern, regardless of atmospheric pCO_2 and emissions, and (2) applying a surface flux calculated internally to meet some objective – here, the value of atmospheric pCO_2 at any point in time.

1. The *forcing*:

`pyyyyz_RpCO2_Rp13CO2_FRALK_FDIC_F13DIC_FCa`

provides a facility for applying alkalinity (ALK) (as well as Ca, and DIC, if selected) to the ocean surface concurrently with a prescribed time-history of restoring of atmospheric pCO_2 (and associated $\delta^{13}C$), while

`pyyyyz_FpCO2_Fp13CO2_FRALK_FDIC_F13DIC_FCa`

is similar, except with a prescribed time-history of flux (emissions) of CO_2 (and associated isotopic composition) to the atmosphere. By default in each, only an ALK flux to the ocean surface is selected. The other three ocean tracers listed in `configure_forcings_ocn.dat` are set to 'f' for flux forcing. As provided in these examples, atmospheric pCO_2 is held at 278 ppm for the restoring, and emissions of 1 PgC yr⁻¹ ($8.3333e+013$ mol yr⁻¹) for the flux forcing version.

By default, *forcings* are set such that the ALK (and Ca, and DIC if selected) is applied uniformly over the ocean surface (the '2' in COLUMN #06). Point sources can be specified by changing this to a '0', or an explicit spatial pattern with a '-2' (which must then be provided in a separate file).

2. The *forcing*:

`pyyyyz_FRpCO2_Fp13CO2_FRALK_FDIC_F13DIC_FCa`

differs firstly in specifying both a CO_2 emissions flux and restoring value for atmospheric pCO_2 . For alkalinity (ALK), both a flux and restoring of the ocean are selected.

Basically, what happens here is when a flux + restoring of ocean ALK is selected together with flux + restoring of pCO_2 , ALK additions to the ocean is made in order to try and maintain the prescribed history of atmospheric pCO_2 , regardless of the CO_2 emissions also specified. If at any one time-step atmospheric pCO_2 is higher than the target value, ALK is added to the ocean with the flux specified in the ALK flux forcing. If atmospheric pCO_2 is lower than the target value, no ALK is added or taken away, unless the parameter:

`bg_ctrl_force_invert_noneg=.true.`

is set, which enables a negative ALK flux to be applied²².

Note that the restoring value of ALK has no meaning and the value set in:

`biogem_force_restore_ocn_ALK_sig.dat`

is not important. Both atmospheric CO_2 flux forcing and restoring forcing time-series are specified 'as normal', and may constitute an SRES CO_2 emissions scenario and RCP based pCO_2 time-history, respectively, for example. For d13C, only the d13C of emissions is specified²³.

If an atmospheric CO_2 emissions is not required, simply set the value in the time-series file to zero. Note that no scaling of the atmospheric CO_2 forcing²⁴ must be applied because it will scale both restoring and emissions ...

²²This is likely pretty un-physical for most applications, hence the default is `.false.`

²³A restoring must **not** also be set.

²⁴e.g. `bg_par_atm_force_scale_val_3`

If the model cannot quite attain the pCO_2 target specified, you probably do not have a sufficiently large ALK flux specified. Conversely, if the pCO_2 target is significant over-shot (technically: under-shot) then you might have prescribed too large a flux.

Prescribe a time-varying history of radiocarbon production in the atmosphere

The original way for doing this was to create a CO_2 flux forcing and give its $\delta^{14}C$ signature an extremely positive value. However, inputs and outputs of isotopic compositions in *cGENIE* are limited to being between -999 and +999 per mil. Prescribing a small flux of CO_2 to the atmosphere with a $\delta^{14}C$ of +999 per mil will not give an equivalent flux as if it were pure ^{14}C (hence needing the calculation of the equivalent total CO_2 flux with a +999 per mil signature).

There is now a simple hack (not yet extended to flux forcings of the ocean) to direct *cGENIE* to interpret the value specified as its $\delta^{14}C$ forcing signature as an absolute flux (in units of mol yr^{-1}) rather than convert from a per mil notation. *cGENIE* knows to use this alternative if the flux of CO_2 specified in the CO_2 flux forcing is **identical** to the value given for ^{14}C . i.e. in order to prescribe a production rate of ^{14}C in the atmosphere sufficient to balance ocean-atmosphere carbon cycling (in the *Cao et al.* [2009] configuration of the model), instead of setting the atmospheric ^{14}C production parameter:

```
ac_par_atm_F14C=0.387E+03
```

One would now set an identical value of 0.387E+03 in both *forcing* files:

```
biogem_force_flux_atm_pCO2_14C_sig.dat
biogem_force_flux_atm_pCO2_sig.dat
```

(or equivalently, re-scale a unit flux forcing given in the *forcing* files using the `bg_par_atm_force_scale_val_5` parameter in the example of ^{14}C (and `bg_par_atm_force_scale_val_3` for bulk CO_2)).

Selection of the required *forcings* in `configure_forcings_atm.dat`²⁵ then looks like this:

```
-START-OF-DATA-
03 f 0.1 t t F 2 01 01 '[carbon dioxide (CO2)]'
05 f 0.1 t t F 2 01 01 '[14C CO2]'
-END-OF-DATA-
```

Now that ^{14}C production is being specified explicitly by means of a *forcing*, one can easily then implement a time-dependent change in ^{14}C production.

Prescribe an injection of radiocarbon-dead DIC

First ... a *base-config* with ^{14}C tracers is needed, e.g.,:

```
cgenie_eb_go_gs_ac_bg_itfclsd_161_JH_ANTH
or
```

```
cgenie_eb_go_gs_ac_bg_itfclsd_161_JH_ANTHFe
```

(with Fe co-limitation of marine biological productivity).

Then, in the *user-config*, an appropriate *forcing* needs to be specified, e.g.:

```
pyyyyx_FDIC_F13DIC_F14DIC
```

and under the heading `-- FORCINGS --`, might look something like:

- For *forcing* selection:

```
bg_ctrl_force_oldformat=.false.
bg_par_forcing_name="worjh2_FDIC_F13DIC_F14DIC"
```

²⁵There is no need to specify a ^{13}C flux (ignore any warnings at start-up). Indeed, formally, there is no ^{13}C associated with ^{14}C production from N_2 . Actually ... DO NOT add a ^{13}C forcing, just in case ...

which prescribes a forcing of DIC plus its (¹³C and ¹⁴C) isotopes to the ocean (somewhere or everywhere).

- Then:

```
bg_par_ocn_force_scale_val_03=0.0833e15
```

sets the flux (mol yr⁻¹), which is equivalent to 1 PgC yr⁻¹.

- To scale the isotopic composition:

```
bg_par_ocn_force_scale_val_04=-60.0
```

```
bg_par_ocn_force_scale_val_05=-999.0
```

for example gives -60 per mil for ¹³C like methane and ¹⁴C that is pretty isotopically dead²⁶.

- By default in the *forcing*, the duration of the emission 1 year, and can be re-scaled (e.g., to 1000 years duration) by:

```
bg_par_ocn_force_scale_time_03=1000.0
```

```
bg_par_ocn_force_scale_time_04=1000.0
```

```
bg_par_ocn_force_scale_time_05=1000.0
```

- Finally – the emission location is specified by, e.g.:

```
bg_par_force_point_i=18
```

```
bg_par_force_point_j=26
```

```
bg_par_force_point_k=7
```

Alternatively, the DIC release can be made over the entire ocean floor or simply sections (or depth intervals) of the ocean floor instead of a point source.

Prescribe a spatial map of benthic tracer release

Flux *forcings* to the ocean with a variety of different spatial attributes can be specified in the *forcings* specification file: `configure_forcings_ocn.dat`. As detailed in the notes to this file, there is a flag (COLUMN #6) which sets the spatial attributes of the *forcing*:

- 3 – '3D' – force the entire ocean volume uniformly
- 2 – '2D' – force the entire ocean surface uniformly
- 0 – '0D' – force a specified point
- 1 – 'SURFACE' – force with/to specified surface ocean pattern
- 2 – 'BENTHIC' – force with/to specified benthic pattern
- 3 – 'LEVEL' – force a specific layer in the ocean with/to specified pattern²⁷
- 4 – 'SURFACE+BENTHIC' – simultaneously force with/to specified (and sperate) surface and benthic pattern

Options -1 through -4 require a 2D field to be provided. The grids for both are the same – i.e., all 'wet' grid points (non dry land) in the model. Templates for these can be created as follows:

1. Open up the *BIOGEM* results file: `fields_biogem_2d.nc` (any experiment).
2. Display the variable: `grid_mask`.
3. Select the Array 1 tab (to display the actual gridded values rather than the color-coded map); highlight the grid of values and then copy-and-paste to a text editor.
4. You should have a grid of values, with a '1.0' representing ocean, and 'NaN' land. The NaNs can then be search-and-replaced to '0.0' and you have a grid valid for either the entire surface ocean or entire benthic surface.

²⁶Note this is on the scale of d14C not D14C

²⁷For k=k_max, this duplicates the SURFACE forcing.

From here: 1s can be replaced by 0s to remove unwanted locations.²⁸

In the forcing configuration file, if the COLUMN #5 flag ('scale flux forcing of tracer?') is set to 't', then the flux applied at each selected location is scaled such that the total applied flux is equal to that given in the *forcing* time-signal file.²⁹

²⁸This can be quite time-consuming and tedious and there is no particular short-cut :(

²⁹The values in the forcing map need not be all 1.0 of course.

16.7 HOW-TO ... do stuff with sediments

Spin-up the full marine carbon cycle including deep-sea sediments

By a 2-step process³⁰:

1. First-guess closed system *spin-up*

As of code release **r4211**, it is possible to carry out the initial spin-up, with a solute input to the ocean via rivers, but also at the same time, with the system configured closed, i.e.,:

```
bg_ctrl_force_sed_closedsystem=.true.
```

The weathering flux is subtracted from ocean cells overlying the sediments to balance the global budget and ensure a closed system. This subtraction involves partitioning the total global weathering flux between each ocean floor cell with a subtraction in proportion to the estimated $CaCO_3$ preservation and burial rate. To utilize this methodology now requires that the **ROKGEM** module is used, i.e., a *base config* such as:

```
cgenie_eb_go_gs_ac_bg_sg_rg_itfclsd_161_JH_BASE
```

A first guess for the weathering flux must now be prescribed. This could be derived from a previous closed system model experiment with no weathering flux specified (diagnosing weathering from total global $CaCO_3$ burial as described earlier), or from the literature, e.g., *Ridgwell* [2007] cites $20 \text{ Tmol HCO}_3^- \text{ yr}^{-1}$, an equivalent $CaCO_3$ weathering rate of 10 Tmol yr^{-1} :

```
rg_par_weather_CaCO3=10.00E+12
```

The following *user config* file

```
EXAMPLE_worjh2_P04_S36x36_SPIN
```

can be used for the closed system spin-up.

To launch an experiment, type (all in one line; notes space separators between line items in this document format):

```
./runCCSgenie.sh cgenie_eb_go_gs_ac_bg_sg_rg_itfclsd_161_JH_BASE /  
EXAMPLE_worjh2_P04_S36x36_SPIN 20001
```

To submit to the cluster type:

```
qsub -q kitten.q -j y -o cgenie_log -S /bin/bash subcgenie.sh  
cgenie_eb_go_gs_ac_bg_sg_rg_itfclsd_161_JH_BASE /  
EXAMPLE_worjh2_P04_S36x36_SPIN 20001
```

20,000 years is probably about the minimum practical *spin-up* time. Primarily – you are looking for convergence in the mean wt% $CaCO_3$ value (averaged sediment composition), which is recorded in the *BIOGEM time-series* file:

```
EXAMPLE_worjh2_P04_S36x36_SPIN
```

2. Open system spin-up

The last stage is an open system spin-up as described previously. The prescribed weathering flux (`rg_par_weather_CaCO3`) is revised and set equal to the diagnosed global $CaCO_3$ burial rate ('Total CaCO3 pres (sediment grid)') as reported in the **SEDGEM** module results file:

`seddiag_misc_DATA_GLOBAL.res`. In addition, an *open system* must now be specified in the *user config*:

³⁰This is a revised methodology compared to that described in the GENIE-1 HOW-TO.

```
bg_ctrl_force_sed_closedsystem=.false.
```

50,000 years is probably about the minimum practical *spin-up* time. Again – you are looking for convergence in the mean wt% $CaCO_3$ value and it is up to you to judge how long the *spin-up* needs to be.

There is still some departure of ocean Ca and ALK inventories during the revised multi-stage spin-up compared to observed (and the initialized values), but this is substantially reduced compared to the original 2-part spin-up methodology as well as to a single spin-up methodology.

TIP: Having completed the full marine carbon cycle spin-up, it is recommended that the $CaCO_3$: POC rain ratio is set invariant – see earlier HOW-TO. If the default $CaCO_3$ parameterization setting is retained, CO_2 -calcification feedback as described in *Ridgwell et al.* [2007b] is enabled.

NOTE: There is no climate feedback by default. To run experiments with feedback between CO_2 and climate, add:

```
ea_36=y
```

at the end of the *user-config*.

Run the sediments at higher resolution (as compared to the ocean grid)

By default (as set in the *base-config* file in `~/cgenie.muffin/genie-main/configs`) the **SEDGEM** sediment grid is configured at a resolution of 36×36 (and on an equal area grid), by:

```
SEDGEMNLNSOPTS='$(DEFINE)SEDGEMNLNS=36'
SEDGEMNLATSOPTS='$(DEFINE)SEDGEMNLATS=36'
```

Several data input files are required by **SEDGEM** consistent with the specified grid:

- A mask, which specifies the sediment grid locations (if any!) at which sediment cores (see: *Ridgwell* [2007]) are to be generated at:

```
sg_par_sedcore_save_mask_name="sedgem_save_mask.36x36"
```

The example provided on SVN contains some illustrative locations set (by a '1') for cores to be generated.

- The required sediment grid topography (bathymetry):

```
sg_par_sed_topo_D="sedgem_topo_D.36x36"
```

This particular grid is derived from observed bathymetry and excludes sediment locations shallower than the surface ocean layer (of the 8-level model) as described in *Ridgwell and Hargreaves* [2007].

As described in *Ridgwell and Hargreaves* [2007], **SEDGEM** can be sub-gridded to a resolution of 72×72 (equal area). The following *namelist* parameter additions are necessary to the *user-config* file:

```
SEDGEMNLNSOPTS='$(DEFINE)SEDGEMNLNS=72'
SEDGEMNLATSOPTS='$(DEFINE)SEDGEMNLATS=72'
sg_par_sed_topo_D="sedgem_topo_D.72x72"
sg_par_sedcore_save_mask_name="sedgem_save_mask.72x72"
```

NOTE: Carbonate chemistry stability problems (= model crash) may occur in the 16-level configuration in conjunction with 72×72 resolution sub-gridded sediments. Who knows why?! :(

Include shallow water depositional systems

By default, the entire seafloor grid is considered as 'deepsea' and sedimentary diagenesis options are provided accordingly. In practice, because the simple diagenesis options available, such as for CaCO_3 (e.g. Archer [1991]; Ridgwell [2001]; Ridgwell *et al.* [2003]) tend not to be applicable to shallower water and particularly high organic carbon delivery (especially in the case of CaCO_3 diagenesis) environments, the deep-sea grid is restricted. This can be prescribed 'hard', by defining the sediment grid only at deeper depths and classifying shallowed ocean grid points as invalid (a value of 0.0) in the SEDGEM depth definition file³¹ (e.g. worbe2.depth.36x36x08). Or, and more flexibly, the entire ocean grid can be defined as potentially valid (e.g. worbe2.depth.36x36x08.ALL) and SEDGEM directed to treat any grid points lying shallower than a specific depth as shallow water sediments. This depth cut-off is set via the parameter `sg_par_sed_Dmax_neritic` (in units of m below the ocean surface). A typical value is 176.0³², which for an 8-level configuration will exclude the shallowest ocean depth level from the deep-sea grid, and for a 16-level ocean will exclude the two shallowest ocean depth levels.

Sediments that are not classed as 'deepsea', are automatically classed as 'mud', i.e. potentially detrital and organic carbon rich. Such sediment locations currently have no option for carbonate burial associated with them. Instead, a limited range of extremely simple and crude assumptions regarding organic carbon preservation (and P regeneration) are provided.

Locations at which carbonate can be produced and preserved are classified as 'reef' locations. Note that the assumption here is that carbonate is precipitated benthically and buried, rather than produced pelagically and settle to the seafloor. A mask needs to be provided³³ in order to define the cells which are *reef* rather than *mud*. The mask consists of the sediment grid, with values being either 1.0 (*reef*) or 0.0 (other). Grid points that are shallower than the depth cut-off and not associated with a value of 1.0 in the corresponding reef mask file, are classified as *mud*.

It can be a little tedious (not really *that* tedious) to create a reef mask that exactly matches all the shallow grid points, so a parameter is provided to force all non *deepsea* grid points to be *reef*. This is achieved by setting:

```
sg_ctrl_sed_neritic_reef_force=.TRUE.
```

This parameter can also be used to force a pattern of *reef* cells as defined by the reef mask, even if they lie deeper than the depth cut-off value (`sg_par_sed_Dmax_neritic`).

Additional information and tips on setting up shallow water sediment grids can be found in some of the EXAMPLES (e.g. EXAMPLE.p0251b.PO4.SPINO).

Set a specific ocean chemistry or saturation state

In the absence of a significant pelagic plankton derived deep-sea carbonate sink, the global burial sink of CaCO_3 is likely to have been dominated by shallow water depositional environments. cGENIE can represent something of these systems and the fundamental dynamical difference between deep-sea pelagic and shallow water CaCO_3 sinks (the former is predominantly controlled by preservation and input of C_{org} , whilst the latter is primarily a function of primary production by benthic calcifiers) by specifying particular shallow water grid cells as 'reefal' (see: 16.7). Also in contrast

³¹Set by parameter `sg_par_sed_topo_D_name` that must point to a file in `genie.muffin/genie-sedgem/data/input`.

³²Note that the simple (esp. CaCO_3 diagenesis schemes) arguably only applicable below depths of ca. 1000 m, although even this is complicated by varying patterns of productivity across the ocean.

³³The parameter name to set the reef mask is `sg_par_sed_reef_mask_name`.

to the deep-sea pelagic CaCO_3 sink, which is mechanistically simulated (both export production and particularly early diagenesis and CaCO_3 dissolution within the sediments), the shallow water CaCO_3 sink in cGENIE is heavily parameterized and is treated as little more than a function relating deposition of CaCO_3 (in units of $\text{mol cm}^{-2} \text{ yr}^{-1}$) at each and every designated reefal grid point. A parameter is provided:

```
sg_par_sed_reef_CaCO3precip_sf
```

that scales the CaCO_3 burial flux so as to achieve some desired global value, such as to balance the global weathering rate. Unfortunately, the value of this parameter is not *a priori* known. Its value (and hence sink strength) also depends on the ambient saturation state. Hence, from the outset, the carbonate saturation properties of the ocean surface must be assumed and set. For the modern ocean, this arises naturally from a system initialized with observed concentrations of ALK, DIC, $[\text{Ca}^{2+}]$, etc. and (preindustrial) atmospheric $p\text{CO}_2$, and in conjunction with an adequate simulation of the large-scale productivity of the ocean and recycling in the interior ocean.

For deeper in the geological past and particularly in the absence of an effective deep-sea pelagic CaCO_3 buffer, ALK and DIC in particular *a priori* not known, although either sensitivity experiment assumptions of the CO_2 concentration required to generate a specific climate and/or proxy data, can give some ideas of $p\text{CO}_2$. It is beyond the scope of this HOW-TO to discuss what geological constraints can be applied in constraining surface ocean saturation (and hence, in conjunction with a given weathering flux, to identify the value of the scaling parameter) but having identified a number (as calcite or aragonite), there are three possible ways of setting an appropriate ocean chemistry:

1. To achieve a specific mean ocean surface saturation state: mean ocean ALK and DIC values can be set consistent with assumptions regarding $p\text{CO}_2$ and $[\text{Ca}^{2+}]$. The parameters specifying the initial mean ocean ALK and DIC concentrations ($\mu\text{mol kg}^{-1}$) are:

```
bg_ocn_init_12=2.3630E-03
bg_ocn_init_3=2.244E-03
```

for their respective modern mean ocean values.

In order to estimate appropriate past concentrations for a given $p\text{CO}_2$ and $[\text{Ca}^{2+}]$, trial and error can be employed, aided by use of a carbonate calculator program such as CO2SYS, although bearing in mind that the saturation conditions calculated in CO2SYS will be w.r.t. the surface, while the 2 model initialization parameters are setting the mean ocean composition – ocean circulation and particularly biological productivity in the open ocean will create an offset between mean surface and bulk ocean properties. Once a reasonable ocean saturation state has been employed, the reefal CaCO_3 depositional parameter can be played with to (re)balance weathering and global burial, although if the initial guess is too far off, this will need to be briefly iterative (as localized CaCO_3 removal and burial will affect the local saturation state and hence in turn burial).

Conclusion: do-able, but tedious.

2. An alternative methodology, somewhat akin to how the system with a responsive deep-sea pelagic CaCO_3 buffer is configured, is to set the system 'open', and allow the balance between weathering and shallow water CaCO_3 burial to set ocean chemistry (whilst restoring $p\text{CO}_2$ to a required value). This is potentially even more tedious, because the value of the scaling parameter is not known. In fact, it is the scaling parameter that will set the mean ocean surface saturation in order to balance weathering and shallow water CaCO_3 burial.

An acceleration technique can be used (see: 16.7) but the process is still iterative (and tedious).

A further provision is hence made in the SEDGEM module that will flux force the ocean with Ca (and also DIC if requested) at each and every reefal cell if the local saturation state falls below a specified target value. In this:

```
sg_ctrl_sed_neritic_reef_force=.TRUE.
```

will turn 'on' the provision of a forcing of ocean chemistry towards a local saturation target,

```
sg_par_sed_ohmegamin
```

sets the saturation threshold, and

```
par_sed_ohmegamin_flux
```

specifies the flux in units of mol Ca²⁺ cm⁻² per time-step at each and every reefal grid point. Setting:

```
ctrl_sed_forcedohmega_ca=.false.
```

will enable the corresponding fluxes of ALK and DIC to be applied.

This methodology can be accelerated as per Section 16.7 but with the downside that what is being set in practice is the minimum saturation at any reefal grid point – i.e. one should obtain a reefal grid point, typically at relatively high latitudes, with ambient chemistry close to the specified saturation, with all other grid points characterized by higher saturation. There is no simple way of deriving the global mean surface saturation in advance.

Having forced ocean chemistry (e.g. just by altering the Ca²⁺ and ALK input to the ocean) and restored to a specified atmospheric *p*CO₂ value, the value of the reefal CaCO₃ burial scaling parameter can be adjusted in order to balance weathering and global burial. A little iteration may probably be required to get a good balance as the ompeting flux input and CaCO₃ removal are very localized (at reefal cells).

Conclusion: do-able, but does not give a simple-to-predict mean global saturation.

3. With a desired global mean surface saturation value in mind, cGENIE can be configured to determine the appropriate ocean ALK and DIC value directly.

In brief – a mean global saturation target value is set by the parameter:

```
bg_par_force_invert_ohmega
```

ALK, and if requested: DIC (and isotopes) and Ca, is fluxed evenly throughout the ocean if the current ocean surface saturation value falls beneath the target, and not fluxed at all otherwise³⁴.

An aragonite saturation value can be forced towards rather than calcite by setting:

```
bg_ctrl_force_ohmega_calcite=.false.
```

The setup for this is a little involved and involves specifying a particular forcing that cGENIE identifies as requiring a saturation target to be matched and requires some additional configuration.

An example *user-config* to spin-up the ocean to a specified saturation value is provided:

³⁴A negative flux of ALK can be enabled, which would act to reduce saturation, by setting the following:
bg_ctrl_force_invert_noneg=.false.

EXAMPLE.p0251b.P04.SPINO

and described in the Examples chapter. A typical configuration would like like the following:

```
bg_par_forcing_name="pyyyyz.RpCO2_Rp13CO2_FRALK_FDIC_F13DIC_FCa"
bg_par_atm_force_scale_val_3=2800.0E-06
bg_par_atm_force_scale_val_4=-6.5
bg_par_ocn_force_scale_val_3=1000.0E12
bg_par_ocn_force_scale_val_4=0.0
bg_par_ocn_force_scale_val_12=2000.0E12
bg_par_ocn_force_scale_val_35=0.0
bg_par_force_invert_ohmega=10.0
```

of which the first line sets the forcing (provided), the second line specifies the atmospheric $p\text{CO}_2$ value to be restored to (and the 3rd its isotopic composition). Because the ocean is either fluxed or not, depending on surface saturation compared to the target, how aggressively the ocean is fluxed is set by lines #4 (for DIC) and #6 for ALK. The values shown here³⁵ are approximately $\times 10$ global weathering for reference. The last line is the required mean ocean surface saturation state. (Lines #5 and #7 set the isotopic composition of injected DIC, and any associated Ca flux, respectively.)

In terms of methodology:

- (a) This saturation restoring would be carried out in a closed system for typically 10 or 20 kyr, depending on whether an initial *spin-up* was being used and how far off any guess as to initial ALK and DIC is.

Depending on the reported global CaCO_3 burial rate³⁶, the reefal CaCO_3 burial scaling parameter is adjusted. This is also a good time if climate-dependent feedback is to be used, to adjust e.g. the baseline mean global temperature.

- (b) The saturation restoring *forcing* is replaced by a simple atmospheric $p\text{CO}_2$ *forcing* (or none at all), and a short, perhaps just 1 kyr experiment is carried out with an open system. Fine-tuning of the CaCO_3 burial scaling parameter is carried out (plus any fine-tuning required to set an exact initial weathering flux).
- (c) A long, open-system *spin-up*, likely accelerated (Section 16.7) 3rd spin-up phase is conducted with no forcing.

Example *user-configs* of this sequence are provided (EXAMPLE.p0251b.P04.SPIN*). Note that the 3rd phase of spin-up could be carried out with a prescribed $p\text{CO}_2$ *plus* associated isotopic composition if not already done so (e.g. as part of the phase #1 of the spin-up).

Conclusion: work-able.

Specify a particular carbonate mineralogy

Firstly – pelagic carbonate production and deep-sea sedimentary diagenesis is inherently assumed to be (all) calcite. Currently there is no alternative option, nor mixed phase (including high-Mg calcite) assemblage option.

Shallow water (neritic) carbonates, however, can be specified as either calcite (the default) or aragonite, the difference being simply in which saturation state is used to calculate precipitation and burial rate. Reefal carbonate precipitation assumed in the form of aragonite is simply set by:

```
sg_par_sed_reef_calcite=.false.
```

³⁵These values represent the *total* flux that distribution throughout the ocean relative to grid cell volume.

³⁶SEDGEM file seddiag_misc_DATA_GLOBAL.res

Obviously, the same scaling value as per for calcite cannot be used (aragonite will require a higher scaling to achieve the same global CaCO_3 depositional flux).

In terms of a spin-up to a specified saturation state (see: 16.7) – an aragonite, rather than calcite (the default) saturation value can be forced towards rather than calcite by setting:

```
bg_ctrl_force_ohmega_calcite=.false.
```

Set up a (silicate) weathering feedback

To create a temperature (only) dependency for the weathering of carbonate and silicate rocks, the following two parameter values need to be set (for carbonate and silicate weathering, respectively):

```
rg_opt_weather_T_Ca=.true.
rg_opt_weather_T_Si=.true.
```

A reference temperature governs the dependency of weathering on climate change and modifies the solute fluxes from weathering scaled (non-linearly) to the deviation of mean global land surface temperature from the reference temperature. The mean global land surface temperature is given in the **BIOGEM** *time-series* file `biogem_series_misc_SLT.res` and is set equal to the reference temperature parameter:

```
rg_par_ref_T0.
```

The baseline (unmodified) solutes fluxes from terrestrial weathering are set by the parameters:

```
rg_par_weather_CaCO3
rg_par_weather_CaSiO3
```

for carbonate and silicate weathering, respectively. At steady-state (no climate perturbation), these must sum up to the total weathering flux which is given by the total global CaCO_3 burial flux found in the file `seddiag_misc_DATA_GLOBAL.res` (in the `genie-sedgem` results sub-directory). To balance the silicate weathering, volcanic CO_2 out-gassing is then assigned a value equal to silicate weathering flux by setting the parameter `rg_par_outgas_CO2`.

There are different ways in which the total weathering flux (equal to total CaCO_3 burial at steady state) can be split between carbonate and silicate weathering and hence a value for volcanic CO_2 out-gassing assigned:

1. All solutes could simply be assumed to be derived from carbonate weathering (which is the default assumption in open system experiments without a weathering feedback), e.g.:

```
rg_par_weather_CaCO3=10.0E+12
rg_par_weather_CaSiO3=0.0
```

for a hypothetical example with 10 Tmol yr^{-1} total global CaCO_3 burial. No volcanic CO_2 out-gassing should then be prescribed.

2. Secondly, silicate and carbonate weathering could be assumed to be split evenly, e.g.:

```
rg_par_weather_CaCO3=5.0E+12
rg_par_weather_CaSiO3=5.0E+12
```

Volcanic CO_2 out-gassing needs to be set equal to the baseline silicate weathering flux:

```
rg_par_outgas_CO2=5.0E+12
```

3. Thirdly, volcanic CO_2 out-gassing could be assumed, which then constrains the silicate flux, with the carbonate flux being whatever is required to make the total Ca^{2+} weathering flux equal to global CaCO_3 burial.

A difficulty arises because setting the reference temperature `rg_par_ref_T0` equal to the mean global land surface temperature only gives rise to weathering fluxes exactly equal to the reference parameter values (`rg_par_weather_CaCO3` and `rg_par_weather_CaSiO3`) for a non-seasonally forced climate. Because weathering is non-linear in climate (here just temperature), a seasonally forced configuration of the model will give rise to slightly modified weathering fluxes. The result is a slight drift in atmospheric $p\text{CO}_2$ and climate, even after a fairly long (100s of kyr), or alternatively, a slightly different steady state $p\text{CO}_2$ value (for a ca. 1 Myr *spin-up*).

The reason is that long-term climate and hence $p\text{CO}_2$ is controlled by the silicate weathering component of total weathering, not total weathering. Under a seasonally forced rather than annual average climate, the non-linearity in the weathering response to temperature deviations means that silicate weathering will generally slightly exceed volcanic CO_2 out-gassing. Atmospheric $p\text{CO}_2$ and with it global temperatures will hence be gradually drawn-down until net (carbonate) carbon removal exactly matches the rate of new (volcanic) carbon input

The imbalance between silicate weathering and volcanic CO_2 out-gassing is given in the BIOGEM output:

`biogem_series_misc_exweather_Ca.res`

which records the absolute excess of weathering compared to out-gassing, in $\text{mol Ca}^{2+} \text{ yr}^{-1}$ as well as a percentage. In an *open system* (not a closed one!), this excess needs to be adjusted 'close' (how close? sub 1 percent or $0.1 \text{ Tmol Ca}^{2+} \text{ yr}^{-1}$, certainly) to zero. This adjustment is done by running a short (a few or 10s of years) experiment, reading off the excess weathering value, and doing one of the following:

1. Adjust the reference temperature parameter `rg_par_ref_T0` – to a lower value if there is an excess of silicate weathering over out-gassing, or
2. Adjust the reference silicate weathering parameter `rg_par_weather_CaSiO3`, or
3. Adjust the volcanic CO_2 out-gassing flux parameter `rg_par_outgas_CO2`.

An exact adjustment can be made for the second and third possibilities, with the easiest being to reduce the out-gassing flux value by the amount of excess weathering. For the first option, a couple of iterations will typically be required in order to determine the new reference temperature value that gives rise to a silicate weathering balance. The second option is the recommended one, with the third being the least recommended. Regardless of which of the 3 options, the total weathering flux, given in the BIOGEM time-series file `biogem_series_diag_weather_Ca.res`, will now be slightly different from the required burial flux. This either requires that the value of the reference carbonate weathering parameter (`rg_par_weather_CaCO3`) is now slightly adjusted, or that a slightly different global carbonate burial flux is allowed.

If the carbon isotopic signature of volcanic CO_2 out-gassing is assigned a value of e.g. -6.0‰ . The $\delta^{13}\text{C}$ of weathered CaCO_3 is then simply set in order that inputs equal the mean $\delta^{13}\text{C}$ of carbonate burial, which as given in the **BIOGEM** *time-series* file:

`biogem_series_sed_CaCO3_13C.res` For example, assuming equal fluxes of 5 Tmol yr^{-1} for both weathering components and for volcanic CO_2 out-gassing (at -6.0‰) and assuming a mean carbonate burial $\delta^{13}\text{C}$ of 3‰ , requires:

```
rg_par_outgas_CO2_d13C=-6.0
rg_par_weather_CaCO3_d13C=12.0
```

Of course, in reality organic carbon burial is important and including it would enable a much more realistic value of weathered carbonate $\delta^{13}\text{C}$ to be set ...

Accelerate the weathering-sedimentation mass balance ('GEMlite')

Also see: The FAQ Chapter.

A (pseudo) module is provided: 'GEMlite' which provides a means of much more rapidly solving the weathering-sedimentation mass balance – i.e. the long-term (>10 kyr) carbon cycle processes and feedbacks. The motivation behind GEMlite is the stark disparity between the time-scales of ocean circulation and biological pump (ca. 0.1-1000 years) and those of sedimentation and weathering (2-20 kyr) and particularly the silicate weathering feedback (>100 kyr). This makes running cGENIE to an open system steady state (with or without the silicate weathering feedback) challenging. Is there any way of 'accelerating' the calculation of the 'long tail' [Archer et al., 2009] of the CO₂ curve (e.g. in response to fossil fuel CO₂ emissions)?

The philosophy is as follows: the long-term weathering-sedimentation processes are effectively just an imbalance between the supply of solutes via weathering and preservation and burial of esp. carbonates in deep-sea (and shallow) marine sediments. For a small imbalance between weathering and sedimentation, atmospheric pCO₂ and climate (and hence the solute flux when including weathering feedbacks) will only change very slightly. For long intervals characterized by only a small imbalance in weathering-sedimentation the key assumption is made: Ocean circulation and the biological pump, and hence the *gradients* of dissolved species in the ocean can be considered *invariant*. Hence, for the purpose of solving weathering-sedimentation over an intervals of time: The ocean can be treated as a *single box*. It further assumes that: The ocean is initially in equilibrium with the atmosphere (w.r.t. CO₂). (This latter assumption does place important limitations on under what circumstances GEMlite can be employed to accelerate experiments.)

This is what GEMlite does – it solves for weathering-sedimentation and applies the mass difference *uniformly* throughout the ocean (as if it were a single box), hence preserving the tracer gradients in the ocean. It also (optionally) calculates and re-partitioning of carbon between ocean and atmosphere. Because ocean circulation and the biological pump etc. do not have to be re-calculated, the accelerated quasi box-model phase can be calculated very considerably faster than the 'full' model. Obviously, if atmospheric pCO₂ and hence climate are changing at an appreciable rate then the assumption of invariance in ocean tracer gradients breaks down and it is not 'safe' to apply the accelerated calculation. Similarly, appreciable changes in nutrient inventories will affect the biological pump and hence also change tracer gradients.

The key to employing GEMlite, in addition to knowing when it is appropriate/not appropriate to employ it, is to decide what balance of accelerated (GEMlite) time-stepping vs. normal (full system update of ocean circulation, biological pump, etc.) time-stepping to employ. This division is implemented by creating a sequence of accelerated vs. non-accelerated time-stepping. This can be done in one of two ways:

1. Fixed sequence.

By default, GEMlite will employed a fixed, pre-determined sequence of accelerated vs. non-accelerated time-stepping. The parameters to specify this sequencing are:

`ma_gem_notyr` – which sets the number of years (the assumed time-step of GEMlite) for 'normal' time-stepping.

`ma_gem_yr` – which sets the number of years for accelerated time-stepping.

For instance: if `ma_gem_notyr=50` and `ma_gem_yr=50`, you would have a sequence with 50 years of full updating, followed by 50 years of accelerated.

For instance: if `ma_gem_notyr=10` and `ma_gem_yr=90`, you would have a sequence with 10 years of full updating, followed by 90 years of accelerated.

etc.

Note that the GEMlite cycle phase of 'normal' time-stepping is **always** done first.

Also note that choosing e.g. `ma_gem_notyr=10` and `ma_gem_yr=100`, while appearing a desirably simple ratio, would result in the change-over point in cycle phase (to accelerated) occurring at the end of year 10, 120, 230, 240, etc. – something that might affect/influence your choice of data saving pattern (i.e., the sequence of time-points for time-series and time-slice data saving).

By default, the parameter values are: `ma_gem_notyr=999999` and `ma_gem_yr=1` meaning that in practice you will never get to the end of the 'normal' time-stepping phase. Note that these parameters are **integers** (setting real numbers, e.g. `1.0E6` will not work ...).

2. Adaptive sequencing.

Here, GEMlite attempts to be clever and optimizes the ratio between the duration of each phase of the cycle.

The motivation for this is that often in model experiments, environmental parameters will tend to change faster at the beginning of an experiment compared to towards the end. Fossil fuel CO₂ release and its long tail of declining pCO₂ is a good example of this. Obviously this complicates the choice of a (fixed) ratio of cycle phases – 100:100 (or more likely: 1000:1000) might not lead to too much degradation of the simulation, but you would only gain a speed advantage of x2 for the experiment as a whole, which if 100-1000 kyr in total duration, is still going to be long. On the other hand: 10:90 would give you a factor almost x10 increase in overall speed, but would seriously degrade the simulation during the initial, rapidly changing environment following CO₂ release.

Adaptive sequencing adjust the time-stepping via 2 criteria:

- In the normal time-stepping phase, if the rate of change of pCO₂ is **more than** a specified threshold over any one year, then the total duration of this phase is extended by one year.
- In the accelerated time-stepping phase, if the total change in pCO₂ since the last normal phase is **less than** a specified threshold, then the total duration of this phase is extended by one year.

The result is that the phase durations are always a minimum of the values set by `ma_gem_notyr` and `ma_gem_yr`. If it is 'unsafe' to switch to accelerated mode, because pCO₂ is changing rapidly, then the model stays in normal mode. If it is safe to stay in the accelerated mode, because pCO₂ has not changed much in total during the phase, then the model stays in the accelerated phase.

The parameter names and default values for the two thresholds are:

- `ma_gem_adapt_dpCO2dt=0.1` (ppm yr⁻¹)
- `ma_gem_adapt_DpCO2=1.0` (ppm)

but these will not necessarily be the ideal of any particular experiment (and some trial-and-error may be called for).

Adaptive time-stepping is enabled by setting:

`ma_gem_adapt_auto=.true.`

(by default it is `.false.`).

The switching between normal (non accelerated) and accelerated phases is saved in a time-series file:

```
biogem_series_misc_gemlite.res
```

As a further refinement, the accelerated phase can be set to be relatively short to begin with, but gradually increasing in length. The parameters controlling this are:

`ma_gem_yr` – the initial accelerated phase duration

`ma_gem_yr_max` – the maximum accelerated phase duration

`ma_gem_adapt_dgemyr` – the (minimum) fractional increase in duration each cycle (or 1.0 yr, whichever is greater)

A reasonable set of parameters:

```
ma_gem_notyr=10
ma_gem_yr=10
ma_gem_yr_max=990
ma_gem_adapt_dgemyr=0.05
ma_gem_adapt_dpCO2dt=0.10
ma_gem_adapt_DpCO2=0.01
ma_gem_adapt_auto=.true.
ma_gem_adapt_auto_unlimitedGEM=.false.
```

Finally ... you will need a *base-config* that has GEMlite enabled. This actually requires nothing more than the addition of a couple of lines (to a *base-config* file):

```
ma_flag_gemlite=.TRUE.
```

which can go e.g. near the start of the file under # GENIE COMPONENT SELECTION. Plus:

```
ma_kgemlite=xx
```

which can go e.g. under # TIME CONTROL AND TIME-STEPPING.

Here, `xx` will depend on the time-step assumed in the *base-config*. This is likely to be either 96: the standard for most *base-configs*, or 48: for low resolution and faster model configurations, which typically have `.t48` in their filename. By convention, I name *base-configs* including GEMlite with `_gl`,

e.g. `cgenie_eb_go_gs_ac_bg_sg_rg_gl.p0000c.BASESLi.t48.config`

but you can name it `BobTheLeglessPony` for all I care.

The **most important** thing is to ensure you are not seriously degrading model fidelity (of carbon cycle simulation) by your adoption and configuration of GEMlite.

Test different assumptions of how the time-stepping phases are scheduled and compare (of possible) against a full experiment in which GEMlite is not used.

It is important to recognize that when the model switches into the GEM phase, it assumes all ocean tracer gradients are fixed, and updates only ocean composition as a whole according to weathering vs sedimentation imbalance (and also tries to re-equilibrate ocean and atm). As part of this, the flux to the sediments is taken from the average of the last year of the preceding normal phase, and fixed. This also means that the $\delta^{13}\text{C}$ of the CaCO_3 deposited to the sediments is fixed ... even if the ocean $\delta^{13}\text{C}$ is being updated and changing ... So, basically you lose the feedback that leads to $\delta^{13}\text{C}$ converging as sinks balance (weathering and volcanic) inputs³⁷.

The solution is to not run in the GEM phase for such long intervals – instead giving the normal

³⁷Adjusting the fluxes themselves during the GEM intervals would break the underlying assumption inherent in the acceleration approximation.

phase a chance to make a brief update of ocean gradients and also d13C of export flux. BUT, if pCO₂ hardly changes, cGENIE runs the risk of staying in the GEM phase for ever (ish)!

A further option:

`gem_adapt_auto_unlimitedGEM`

sets whether GEM is allowed an unlimited phase duration or not. By default it is `.false.`. This means that the maximum GEM duration is limited to the normal `gem_yr` parameter. Also, if excessive (pCO₂) drift occurs, the model will immediately switch to the normal phase.

By default then:

- `gem_notyr` specifies a MINIMUM duration for a normal phase..
- `gem_yr` specifies a MAXIMUM duration for a GEM phase.

Values of `gem_yr` much less than 100 are not advisable as you will not reestablish a new equilibrium gradient of tracers in the ocean in that time.

16.8 HOW-TO ... develop the model code

Best not to. But ...

Add 'name-list' (run time) parameters

In order to create a new 'namelist' parameter, i.e. a parameter whose value can be set in a *user-config* file, you need to edit a total of 4 files:

1. *_lib.f90

Add an entry in the relevant module library file, which for BIOGEM would be:

```
biogem_lib.f9038.
```

The parameter must be defined (with an appropriate type) and added to the NAMELIST section at the top of the file. Simply follow the format of the existing entries and add to the most appropriate section of parameter categories. Note that the parameter name appears *twice* – one in defining its type, and once in adding the the parameter NAMELIST.

2. *_data.f90 For completeness, there is an entry in the subroutine that reports the selected parameter options upon model start-up (if this reporting is selected in the first place ...), which for BIOGEM, is subroutine:

```
sub_load_goin_biogem
```

that lives in:

```
biogem_data.f90.
```

Again – simply follow the format of existing entries for creating a new one. Again: add in an appropriate section of parameter categories to prevent future coders going mad looking for something.

3. Add a new definition (including brief description and default value) in the xml definition file: definition.xml

that can be found in the directory:

`cgenie.muffin/genie-main/src/xml-config/xml` You will have to scroll down to find the section for the appropriate module, and then within that, the section for that category of parameter. Simply follow the format of the existing entries.

4. <FILE>.f90 (or <FILE>.f77)

Finally, edit into the appropriate FORTRAN source file the code that incorporates the parameter that you wish to use. D'uh!

Having ensured the model compiles and seems to do what you wish it do to – run a standard test to confirm that nothing obvious has been unintentionally affected by the change. To do this, simply type `make testbiogem` and confirm that the test passes ...

16.8.1 Define a new tracer

You probably should not be doing this ... but ... just in case ...

Basic definition procedure

1. The starting point to adding a new tracer in *cGENIE* is to add the its definition to the relevant tracer definition file.

There are three tracer definition files that live in `cgenie.muffin/genie-main/data/input`:

- `tracer_define.atm`
- `tracer_define.ocn`

³⁸`cgenie.muffin/genie-biogem/src/fortran`

- `tracer_define.sed`

that house a list of atmospheric (gaseous), oceanic (dissolved), and sediment (solid) tracer definitions. Each file has a similar format, with a series of columns³⁹ holding information on:

#01 The short (mnemonic) name for the tracer. This is used in creating the output filenames and netCDF variable names. In theory, this could be anything you like, but to a limit of 16 characters.

#02 The identifier (index) of the tracer. This must be a unique number – one number for each tracer.

#03 The 'dependency' of the tracer. For instance, an isotope depends on the bulk (and lower mass) tracer. A scavenged element depends on bulk organic matter. For a bulk tracer⁴⁰, its dependency is itself.

The dependency is used in the code to automatically determine any tracer that it depends on and in the case of isotopes, to calculate the δ value.

#04 Is the tracer variable 'type', used internally to determine what to 'do' with a specific tracer:

'1' → assigned to primary biogenic phases; POM (represented by POC), CaCO₃, opal (all contributing to bulk composition)

'2' → assigned to abiotic material (contributing to bulk composition); det, ash

'3' → assigned to elemental components associated with POC; P, N, Cd, Fe

'4' → assigned to elemental components associated with CaCO₃; Cd

'5' → assigned to elemental components associated with opal; Ge

'5' → assigned to elemental components associated with det; Li

'7' → assigned to particle-reactive scavenged elements; ²³¹Pa, ²³⁰Th, Fe

'8' → assigned to carbonate 'age'

'9' → assigned to the fractional partitioning of biogenic material (for remineralization purposes)

'10' → assigned to misc / 'inert'

'>10' → assigned to isotopic properties: 11==¹³C, 12==¹⁴C, 13==¹⁸O, 14==¹⁵N, 15==³⁴S, 16==³⁰Si, 17==¹¹⁴Cd, 18==⁷Li, 19==¹⁴⁴Nd, 20==⁴⁴Ca, 21==⁹⁸Mo, 22==⁵⁶Fe.

#05 Tracer long name. Cannot be more than 128 characters in length.

#06 Units.

#07 Minimum valid value for the tracer. Values lower than this are classed as NaN in the netCDF output.

#08 Maximum valid value. Values higher than this are classed as NaN in the netCDF output. Hence, the first thing to do is to add an entry for the required tracer(s) to the relevant file(s), using the above information and keeping a consistent format and convention with the existing tracers.

If you have an elemental or isotopic tracer that is taken up by a growing phytoplankton cell and incorporated structurally into POM, you will also need to define equivalent dissolved (and recalcitrant) dissolved organic matter tracers (DOM and RDOM). If the tracer is associated with organic matter (or other particles), then you require a scavenged particulate tracer but no corresponding dissolved tracer. For something like iron, which is both incorporated into the

³⁹The meaning of the columns is also summarized at the end of the file.

⁴⁰The elemental components of organic matter (P, N, etc) count as bulk tracers for the purpose of dependency.

cell, and scavenged, you need both types of particulate tracer plus a set of dissolved organic matter tracers ...

2. A set of tracer-related definitions also exists in:
`cgenie.muffin/genie-main/src/fortran/cmngem/gem_cmn.f90`
 Requiring some editing-attention here is⁴¹:
 - ca. L24-26** Three parameters define the main tracer array sizes. (Unfortunately this information has to be duplicated elsewhere due to the peculiarities of the GENIE code structure – see below.) Their values must be equal to the total number of tracers defined in the tracer definition files.
 - ca. L72-261** So as to simplify referencing the tracers in the code, each tracer is assigned a simple mnemonic. This mnemonic may or may not be the same as the short name defined in the tracer definition files. With a little bit of code, the tracer definition short name could have been turned into an index value, but to date this has not been implemented. However, while slightly tedious to set up, created fixed and compile-time rather than run-time mnemonic assignments is going to be somewhat faster as well as making the code slightly more compact.
 - ca. L630-642** Definition of the isotope standards. Only if creating a new isotope system does this need to be edited.
 So effectively, just the total number of tracers, and the addition of the tracer mnemonic name, has to be edited in this file.
3. An extremely unfortunate fact of GENIE code structure life is that the total tracer numbers are defined a second time in:
`cgenie.muffin/genie-main/genie_control.f90`
 at ca. L144-146, and need to be edited consistent with the values in `gem_cmn.f90` (see above). Why ... ? Please don't ask.
4. Equally, or arguably even more annoying and opaquely justified, is the need to edit a number of entries in:
`cgenie.muffin/genie-main/src/xml-config/xml/definition.xml`
 A number of sets of runtime parameters exist with one entry per tracer. Given that all runtime parameters must be defined in the xml definition file, means that every time a new tracer is created, a number of xml entries must be created⁴² :(The approximate line occurrence of this outrage against all good programming practice, are as follows:
 - ca. L397-1159** The tracer arrays holding information about which tracers are selected.
 - ca. L2538-3271** Tracer arrays for the initial value of ocean (dissolved) tracers, as well as a tracer value modification array.
 - ca. L4343-5234** Tracer arrays for atmospheric (gaseous) and ocean (dissolved) tracers governing tracer forcings (value and tie-scale).
 - ca. L5292-5370** Tracer array for the initial value of atmospheric (gaseous) tracers.
 To edit in new entries – simply follow the format of the last entry.
 In addition – at the start of each array definition (xml tag starting `<ParamArray`), the array size is defined (`extent=`). This also needs to be edited to reflect the inclusion of additional tracers.
5. Even more annoying, if possible, is the need to then edit the array entries in the Python xml parameter translation script:
`cgenie.muffin/genie-main/config2xml.py`

⁴¹Watch out that the line numbers may have changed somewhat ...

⁴²Note that there are no sediment (solid) tracer arrays for either initial composition or forcings.

It should be obvious where there additional entries corresponding to the new tracers need to be added. Just be careful of the formatting (the last entry in the list of tracers for each array not having a ', ' terminating the line. Otherwise, simply follow the existing format.

That is it for the basic tracer definition. The model should now compile without error (and still pass its `make testbiogem`) although you'll need to do a `make cleanall` first. However, whilst 'knowing' about the possibility of the new tracers, at this point you have not actually selected any for an experiment and no initial conditions will be read in, nor relevant output created.

Testing

Having checked the model compiles and passes the basic BIOGEM test, the next step is to check that the new tracer(s) is initialized correctly (atm and ocn tracers) and that appropriate output is generated.

1. Create a new *base-config* by taking an appropriate/comparable existing *base-config*⁴³ and modifying it.

The first modification is to increase the number of ocean tracers, defined on the line headed by:

```
# Set number of tracers
```

incrementing the total by the number of additional tracers to be included in the new *base-config* as compared to the original one.

The second modification is to select the additional tracers – simply add appropriate entries to the list of selected tracers (the tracers are selected by setting the parameter value to `.true.` as by default the are `.false.`).

The final modification, in the case of atmospheric (gaseous) and ocean (dissolved) tracers is to set an initial value. If you do not do this, by default, concentrations are initialized to zero.

2. Now go create a *user-config* file. Copy an EXAMPLE config file – one corresponding to the unmodified *base-config*, if possible. You should rename it, and although no modifications are required to the parameter settings in order for the model to run, to ensure all possible output is produced, set:

```
bg_par_data_save_level=99
```

3. Run a brief experiment and check that the tracer appears in the output – both time-series and netCDF. For ocean tracers, the concentration field should progressively look like salinity as concentration changes at the surface will be influenced by P-E. (Remember that at this point, no other transformations or changes of tracer have been defined – just that there is a tracer and it is initialized to a certain value.)

Adding (and testing) a basic tracer biogeochemical cycle

Now for the trickier part, assuming you do not just want a simple passive tracer (there are plenty of 'color' tracers defined already!) and assuming you have got the tracer already successfully configured and running as a simple passive tracer (i.e. previous steps).

The example will be for an ocean tracer that is incorporated into particulate organic matter (POM) (and hence creating an associated sedimentary tracer) during biological productivity. Other and much more fun and entertaining complexities will apply if e.g. the ocean tracer exchanges with the atmosphere and hence is associated with an atmospheric tracer.

1. The relationships between different sorts of tracers, e.g. dissolved and gaseous, and dissolved and solid, are defined in subroutine `sub_def_tracerrelationships`⁴⁴

⁴³`/cgenie.muffin/genie-main/configs`

⁴⁴`cgenie.muffin/genie-main/src/fortran/cmngem/gem_util.f90`

If there is an equivalent dissolved organic matter tracer corresponding to the particulate organic matter one, the relationship between POM and DOM (and also RDOM) also needs to be defined.

Typically, the relationship between a particulate and dissolved inorganic, or particulate and dissolved organic will be 1.0, but depending on the species concerned, it may be 2.0 (or its reciprocal) and/or negative. These values can be modified later if necessary, and this occurs depending on the redox state of the ocean in⁴⁵:

```
sub_data_update_tracerrelationships
```

- For a dissolved inorganic species being taken up biologically, a 'Redfield' like ratio is defined and used to relate the cellular quotient of the tracer versus carbon⁴⁶. An array (`bio_part_red`) stores the relationship of every particulate tracer to every other particulate tracer. It is hence mostly zeros, except for the ratios of the particulate tracers to carbon in both organic matter and CaCO_3 (and for that matter, opal)) and of the isotope ratios of species to their bulk equivalent. The array is (re)populated each time-step in `sub_calc_bio_uptake`⁴⁷, typically by directly applying a globally applicable ratio that is read in at run-time (and hence requiring a new *namelist* parameter to be defined), by some function of ambient environmental conditions that is often a modification of the run-time parameter.

If steps #1 and #2 have been completed correctly, there should now be a biological cycle of the new tracer, with it being taken up at the ocean surface and incorporated into POM with a specific ratio compared to carbon (set by the new *namelist* parameter) and resulting in depletion of the inorganic dissolved tracer at the ocean surface. Conversely, there should be elevated concentrations of the inorganic tracer at depth, mirroring the pattern of e.g. $[\text{PO}_4]$. Creation and subsequent remineralization of a corresponding tracer incorporated into DOM (and RDOM if selected) should occur automatically. However, the recommended first step in testing the newly defined biogeochemical cycle is to disable all DOM formation, by setting:

```
bg_par_bio_red_DOMfrac=0.0
```

Also recommended is to enable 'auditing' of all the tracer inventories to ensure that tracers are not being spuriously created or destroyed by:

```
bg_ctrl_audit=.true.
```

Scavenged tracers are automatically remineralized along with the corresponding parent particulate tracer by default⁴⁸. However, there is no scavenging or creation of scavenged tracers by default. A call would need to be added to⁴⁹:

```
sub_box_remin_part
```

(ca. L2961), e.g. following the examples of Fe scavenging and H_2S reaction with organic matter (treated as a form of 'scavenging' for simplicity of code structure), plus a corresponding subroutine added in which is the scavenged particulate tracer concentration is calculated and the removal of the corresponding dissolved tracers set.

⁴⁵`cgenie.muffin/genie-biogem/src/fortran/biogem_data.f90`

⁴⁶A carbon currency is used in the model rather than phosphate, despite the classic Redfield ratio being defined relative to a phosphate quotient of 1.0

⁴⁷`cgenie.muffin/genie-biogem/src/fortran/biogem_box.f90`

⁴⁸They can instead be set to remain in the scavenging medium by setting a non zero value of `par_scav_fremin`, which sets the fraction of the scavenged tracer that is remineralized along with the degraded parent particulate tracer.

⁴⁹`cgenie.muffin/genie-biogem/src/fortran/biogem_box.f90`

16.8.2 Add additional results output

If a new data field can be derived from an existing field, then creating additional results saving is relatively straightforward because no new time-averaging has to be carried out (i.e. you create the new field based on annual (or sub-annual) averages that are already calculated and available. (If not – refer to the 'full' data saving sub-sections.) Note that new tracers are automatically saved.

Data saving 101

For new *time-slice* saving, code needs to be added to either⁵⁰:

```
sub_save_netCDF_2d_USER
```

or:

```
sub_save_netCDF_3d_USER
```

depending on whether the field is 2D or 3D, respectively. Add the code to the end of the subroutine (as marked). Follow the format of previous data saving as far as possible. For general format is:

- Add a conditional to define under what circumstances, particularly selected save options, the data is saved. DO NOT save it by default ... unless it is of vital importance to the future of the planet. Refer to the user-manual for the categories of save options, investigate what options are used for similar data fields, and use your common sense ...
- Specific a units name for `loc_unitsname`, or set to 'n/a' if not applicable (or non-dimensional).
- Initialize (zero) the local 2D (`loc_ij`) or 3D (`loc_ijk`) (depending on the data field) data array.
- Calculate the data, employing a nested loop if necessary (i.e. simply matrix math cannot be employed) and assign to the local data array.
- Add a call to `sub_adddef_netcdf` (don't ask questions – follow the general format).
- Add a call to `sub_putvar2d` (ditto).

For new *time-series* saving ... it is sort of both more and less complicated :o) The new code goes in `biogem_data_ascii.f90` but in two different places: `sub_init_data_save_runtime` and `sub_data_save_runtime`. For former creates the (ASCII) file and adds a header (to define the columns), and then closes the file. The latter opens the now existing file, writes the output, including the (*time-series* save point) time, and the closes the file. The code needed (again – follow the generla format and best to add the new code to the end of the subroutines) is hence ...

In `sub_init_data_save_runtime`:

- Add a conditional to define under what circumstances, particularly selected save options, the data is saved. DO NOT save it by default ... unless it is of vital importance to the future of the planet. Refer to the user-manual for the categories of save options, investigate what options are used for similar data fields, and use your common sense ...
- (If multiple different variables stored in or based on the same array, set up a loop.)
- Create the filename to be sued (`loc_filename`) via a call to `fun_data_timeseries_filename`. All files are `*_series` with the specific variable type and variable after, or if not a specific variable type, then `_misc`.
- Create the header text. If you start with a '%' then it is all the more MUTLAB friendly.
- Follow the sequence format of: CHECK / OPEN / CHECK / WRITE / CHECK / CLOSE / CHECK (yawn) ...

In `sub_data_save_runtime`:

⁵⁰`biogem_data.netCF.f90`

- Add the same conditional as used in `sub_init_data_save_runtime`.
- (If multiple different variables stored in or based on the same array, set up a loop.)
- Construct the local filename *exactly* as before (`sub_data_save_runtime`) or it will not find the file you have created ...
- Calculate the data value(s) (`loc_sig`).
- CHECK / OPEN / CHECK / WRITE / CHECK / CLOSE / CHECK (yawn) ...

16.9 HOW-TO ... random

Set up carbon dioxide removal (CDR) geoengineering experiments

There are various ways implement different carbon dioxide removal strategies in cGENIE. For instance – additions of dissolved iron and phosphate can be implemented as simple flux forcings to the ocean surface, as described in the Tutorial exercises. Similarly, ocean 'liming' can be implemented as a flux *forcing* of alkalinity to the surface (with or without associated Ca²⁺ and with or without additional CO₂ emissions to the atmosphere due to the creation of lime). There is also the facility for automatically calculating the liming required for a specific policy target (atmospheric CO₂ history or desired mean ocean surface pH or saturation state). This is described in a subsequent HOWTO. This section describes a framework created for applying additional geoengineering modifications and particularly ones that cannot be implemented as a simple flux *forcing*.

Geoengineering with ... 'pipes'

Pipes are parameterized following *Yool et al.* [2009] (Yool, A., J. G. Shepherd, H. L. Bryden, and A. Oschlies (2009), Low efficiency of nutrient translocation for enhancing oceanic uptake of carbon dioxide, *J. Geophys. Res.*, 114, C08009, doi:10.1029/2008JC004792.) This is selected by setting:

```
bg_opt_misc_geoeng='pipes'
```

(Currently, there is no other option and anything passed other than a value of 'pipes' results in the default: no geoengineering.) A series of option then control the working of the pipes:

- A mask file is provided to designate the grid points of the ocean with pipes in them, via:

```
bg_par_misc_2D_file
```

with a default of 'misc.dat'. The default file location is:
cgenie.muffin/genie-biogem/data/input.

This file is treated in a similar way to the normal 2D *forcing* files. The values at each grid point can be scaled via the parameter: `bg_par_misc_2D_scale`. The units are m³ per year. e.g. setting `bg_par_misc_2D_scale=1E13`, assuming values of 1.0 or 0.0 in `misc.dat` will create an annual vertical advective flux at each grid point equivalent to 30% of the volume of the surface cell (3.2E13).

- The ocean depth level associated with the base of the pipes is set via:

```
bg_par_misc_kmin_pipe=12
```

- Three parameters are then provided to control what is advected, with a number of combinations of tracers possible (useful for diagnosing the relative importance of e.g. nutrients vs. respired CO₂ vs. temperature and salinity (and hence ocean circulation changes)):

```
- # pump T and S?
bg_ctrl_force_GOLDSTEInTS=.false.
```

```
(the default) prevents T and S being advected.
- # ONLY pump T and S?
bg_ctrl_force_GOLDSTEInTSonly=.true.
```

results in **only** T and S being advected. This requires that `bg_ctrl_force_GOLDSTEInTS=.false..`
Its default is `.false..`

```

- # pump no DIC?
  bg_ctrl_misc_geoeng_noDIC=.false.
  prevent DIC from being advected.

```

The following combinations are then valid (shown commented out (#) are the settings that are the same as the default settings and hence do not need to be re-defined, although it would not hurt to):

1. `bg_ctrl_force_GOLDSTEInTS=.true.`
`#bg_ctrl_force_GOLDSTEInTSonly=.false.`
`#bg_ctrl_misc_geoeng_noDIC=.false.`
 results in everything being advected.
2. `#bg_ctrl_force_GOLDSTEInTS=.false.`
`#bg_ctrl_force_GOLDSTEInTSonly=.false.`
`#bg_ctrl_misc_geoeng_noDIC=.false.`
 results in everything (nutrients, DIC, ALK, isotopes, etc.) except T and S being advected.
3. `#bg_ctrl_force_GOLDSTEInTS=.false.`
`#bg_ctrl_force_GOLDSTEInTSonly=.false.`
`bg_ctrl_misc_geoeng_noDIC=.true.`
 results in everything except T and S *and* DIC being advected (i.e. just nutrients, alkalinity, isotopes etc.).
4. `bg_ctrl_force_GOLDSTEInTS=.true.`
`bg_ctrl_force_GOLDSTEInTSonly=.true.`
`#bg_ctrl_misc_geoeng_noDIC=.false.`
 results in only T and S being advected.

Combination #1 is arguably the only realistic setting – the others being for diagnosing how the model works and the primary controls on the effectiveness or otherwise of pipes, only. The difference between #2 and #1 indicate the importance of changes in ocean circulation driven by T and S, which can also be assessed in isolation via option #4. The difference between option #3 and #2 indicates the importance of the respired CO₂ 'leak' in the effectiveness of ocean pipes. (Note that there is no option for removing DIC only and e.g. advecting T and S and nutrients etc etc.)



17. Examples



Bibliography

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